ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH

NNOVARE
ACADEMIC SCIENCES
Knowledge to Innovation

Vol 9, Issue 2, 2016

Online - 2455-3891 Print - 0974-2441 Research Article

HISTOLOGICAL EFFECT OPTIMIZATION COMBINATION OF *BACILLUS SPHAERICUS* 2362 AND *BACILLUS THURINGIENSIS* SUBSP. *ISRAELENSIS* IN MIDGUT OF *CULEX QUINQUEFASCIATUS* RESISTANCE

RITA TJOKROPRANOTO1*, TRI HANGGONO ACMAD2, ENDANG SUTEDIA3, SUSY TJAHJAN11

¹Department of Parasitology, Maranatha Christian University, Bandung, Indonesia. ²Department of Biochemistry, Padjadjaran University, Bandung, Indonesia. ³Department of Skin & Venereology, Padjadjaran University, Bandung, Indonesia. Email: rita_tjokropranoto@yahoo.com

Received: 08 January 2016, Revised and Accepted: 19 January 2016

ABSTRACT

Objective: The objective of the study was to analyze pore-forming at *Culex quinquefasciatus* resistant larvae midgut and analyze optimization ratio after treatment using combination *Bacillus thuringiensis var. israelensis* (*Bti*) and *Bacillus sphaericus* (*Bs*) 2362.

Methods: This research was an experimental study. *C. quinquefasciatus* larvae were divided into 10 groups. The Group I until VII had treatment by the various concentration of combination *Bti* and *Bs* 2362, treatment Group VIII as a positive control, Group IX as a negative control, and Group X as a single *Bs* 2362. All of the treatment groups were examined for the histological effect of *C. quinquefasciatus* larvae midgut by hematoxylin eosin. The lowest lethal concentration 50% (LC₅₀) was a standard optimization ratio of combination *Bti* and *Bs* 2362. LC₅₀ was analyzed by probit.

Conclusion: Combination of Bti and Bs 2362 was shown pores at C. quinquefasciatus larvae midgut, and optimization ratio was shown in Group I.

Keywords: Bacillus thuringiensis var. israelensis, Bacillus sphaericus 2362, Culex quinquefasciatus midgut.

INTRODUCTION

Mosquitoes are vector many kind mosquito-borne diseases such as malaria, filariasis, dengue, yellow fever, chikungunya, and multiple viral encephalitis [1-3]. *Culex pipiens quinquefasciatus* transmit of filariasis, West Nile Virus, and Japanese Encephalitis [4]. Approximately 250 million people have been the world burden of lymphatic filariasis [5]. Almost all of province in Indonesia endemic of lymphatic filariasis such as North Sumatera, South Sumatera, Bangka Belitung, Papua, East Kalimantan, Central Java, Tangerang and more than 17 districts of West Java [6]. Therefore, adequate mosquito control strategies are important for interrupt mosquito-borne diseases transmission [7].

Chemical insecticides have been used during four decades, extensive used of synthetic insecticides have caused the development of resistance in mosquito, environmental pollution, harmful effects on beneficial non-target animals, food chain contamination [1,5,8]. Therefore, we need the alternative environment-friendly control agents, the bacterial larvicide that can be save used such as *Bacillus sphaericus* (*Bs*) and *Bacillus thuringiensis serovar israelensis* [1,9].

Over the past two decades, biological larvicide has been used as a mosquito vector control programs in the world [10,11]. The continuous exposure of *Bs* has been result in development of moderate to the high-level resistance of *C. quinquefasciatus* [8]. *Bacillus thuringiensis subsp israelensis* produced crystal protein toxin (Cry11Aa, Cry4Aa, and Cry4Ba), cytolitic protein toxin (Cyt1Aa) [12]. Cyt1Aa have strong binding affinity to the apical brush border of midgut epithelial cells of dipterans, which the Cyt1Aa toxin have the ability to perforate cells membrane [13]. Cry4Ba toxin and Cry11Aa toxin were shown to form pores in the cells membrane of dipterans [14]. Mixture combination of *Bs* with purified Crys of Cyt1Aa at a 10:1 ratio were completed suppressed resistance in *C. quinquefasciatus* population in the field [15]. Combination of Cry powders Cyt1Aa with *Bs* powders were resulting

toxicity was much greater than *Bs* alone against *Aedes aegypti* [16]. *B. thuringiensis* Cry and Cyt toxins known as pore-forming toxins [12]. The aim of this study was to analyze pore-forming at *C. quinquefasciatus* resistant larvae midgut after treatment using optimization combination *Bacillus thuringiensis susp israelensis* and *Bs* 2362, which we used a new combination *Bs* 2362 and *Bacillus thuringiensis var israelensis* (*Bti*), following the single *Bs* 2362 have not been used in the laboratory yet in Indonesia and this research investigated to overcoming new resistance *C. quinquefasciatus* larvae to *Bs* 2362 in the laboratory at Department Parasitology of Gadjah Mada University, Yogyakarta in Indonesia.

METHODS

Materials

Bs 2362 water dispersible granule (WDG) and Bti WDG (Institut Pasteur Paris), temephos 1% (Gadjah Mada University), aquadest, plastic cups 200 ml, pasteur pipette, anatomical forceps, buffer formalin 10%, measuring glass 100 ml, beaker glass, Erlemeyer glass 500 ml, analytic scale, plastic cups, 10% sucrose solution, buffer formalin 10%, liquid paraffin, tissue processor (Sakura, Torrance, CA), embedding machine (Sigma-Aldrich, St.Louis), Microm HM355s microtome (Thermo Scientific, Kalamazoo, MI), Fisherbrand Superfrost Plus slides, water bath, hot plate, xylol I, II, III, alcohol 100%, 95%, 80%, 70% Mayer-Hematoxylin solution, Eosin solution, glass object, cover glass, light microscope, mosquito cages, fish food, plastic trays.

Mosquitoes

A laboratory colony of *C. quinquefasciatus* was obtained from the Department of Parasitology, Medicine Faculty, Gadjah Mada University, Yogyakarta, Indonesia. 750 early fourth-instar larvae were used in the bioassays. 25 *C. quinquefasciatus* larvae were put into each 7 treatment groups combination plastic cups, following one single *Bs* 2362 treatment plastic cups, temephos treatment plastic cups as a positive

control, and aquadest treatment plastic cups as a negative control [17]. Seven concentration combinations were used for this research; the formula of replication was used [18]:

7 (r-1) ≥ 6 7r-7 ≥ 6 7r ≥ 6 + 7 7r ≥ 13r ≥ 1.85 ≈ 3

Each sample replication was 3 times. Amount of plastic cups for this study were 30 cups.

Methods

The research was a true experimental laboratory. C. quinquefasciatus early fourth-instar larvae were divided into 10 groups. The concentration treatment Group I was combination of Bs 2362 and Bti (8:2) part a million (ppm), the combination treatment Group II was (5:5) ppm, the combination treatment Group III was (7:3) ppm, the combination treatment Group IV was (6:4) ppm, the combination treatment Group V was (2:8) ppm, the combination treatment Group VI was (3:7) ppm, combination treatment Group VII was (4:6) ppm, treatment Group VIII used temephos 1% as a positive control, treatment Group IX used 100 ml aquadest as a negative control, and the treatment Group X as a single Bs 2362. C. quinquefasciatus fourth-instar larvae were dryed at tissue paper than the C. quinquefasciatus early fourth-instar larvae were put in a tube which contains 10% buffer formalin, thereafter C. quinquefasciatus larvae put in the freezer at - 70°C. The C. quinquefasciatus early fourth-instar larvae were examined for the histological effect of midgut by coloring hematoxylin eosin. The lowest lethal concentration 50% (LC $_{50}$) was a standard optimization ratio of combination Bti and Bs 2362. LC_{50} was analyzed by probit.

RESULTS

The combination treatment Group I Bs 2362 with Bti (8:2) ppm was shown many pores at midgut of C. quinquefasciatus early fourth-instar larvae. The optimization combination was the treatment Group I (8:2) ppm which has given the result the lowest LC_{50} 2.274 ppm. The result of treatment Group I was shown in Fig 1.

The combination of treatment Group V Bs 2362 with Bti (2:8) ppm and combination treatment Group VII (4:6) ppm were also shown pores at C. quinquefasciatus early fourth-instar larvae midgut. The LC_{50} of combination treatment Group V and VII were 2.276 ppm and 2.279 ppm, respectively, the results were almost the same as with treatment Group I. The picture of midgut C. quinquefasciatus early fourth-instar larvae from combination treatment V and VII was shown in Figs 2 and 3.

While the treatment Group X with single Bs 2362 was not shown pores at midgut of C. quinquefasciatus early fourth-instar larvae. The LC $_{\rm 50}$ single Bs 2362 was 32.675 ppm, which biggest than treatment Group I until VII. The histologic picture of single Bs 2362 was shown in Fig. 8.

The results of LC $_{\rm 50}$ of various combination of $\it Bs$ 2362 with $\it Bti$ were shown in Table 1.

DISCUSSION

The combination of *Bs* 2362 with *Bti* Group I (8:2) ppm was the optimization combination with the lowest LC_{50} was 2.274 ppm and shown the most pores at midgut of *C. quinquefasciatus* fourth-instar

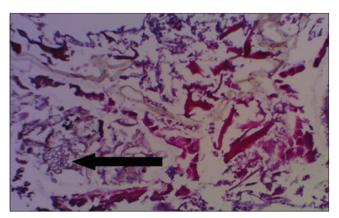


Fig. 1: Midgut pores of *Culex quinquefasciatus* larvae in treatment Group I (black arrow shown pores of midgut)

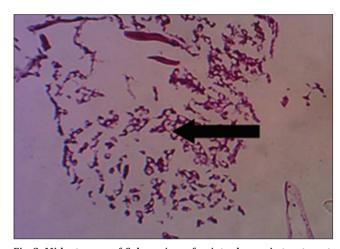


Fig. 2: Midgut pores of *Culex quinquefasciatus* larvae in treatment Group V (black arrow shown pores of midgut)

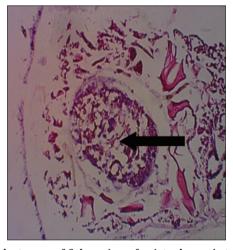


Fig. 3: Midgut pores of *Culex quinquefasciatus* larvae in treatment Group VII (black arrow shown pores of midgut)

larvae at Laboratory Parasitology Gadjah Mada University, Yogyakarta, Indonesia. Laurence Depres, et al. have written at their book about Bti have four toxins Cry4Aa, Cry4Ba, Cry11A, and Cyt (Cytolitc) 1Aa, that the function of Cry4Aa and Cry4Ba were making pores at midgut of epithel mosquitoe larvae [12,14]. Cyt1Aa was strongly affinity of unsaturated fatty acids on epithel mosquitoe larvae midgut; therefore, it is having ability midgut membrane perforation [13]. Bs produced mosquitocidal

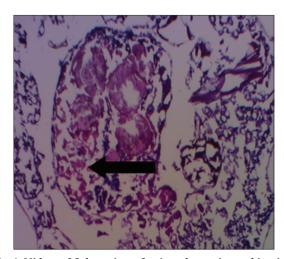


Fig. 4: Midgut of *Culex quinquefasciatus* larvae in combination treatment Group II was (5:5) ppm (black arrow shown pores of midgut)

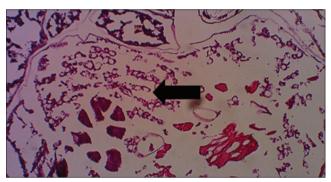


Fig. 5: Midgut of *Culex quinquefasciatus* larvae in combination treatment Group III was (7:3) ppm (black arrow shown pores of midgut)

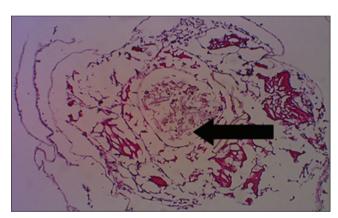


Fig. 6: Midgut of *Culex quinquefasciatus* larvae in combination treatment Group IV was (6:4) ppm (black arrow shown pores of midgut)

toxin (*Mtx*) and binary (*Bin*) toxin, Bin binds with a single receptor of brush border membrane epithel *Culex* larvae by digestive enzyme *Culex* pipiens maltase 1, Bin toxin making pores at receptor of brush border membrane epithel *Culex* larvae [19]. *Mtx* has 3 types of toxin *Mtx* 1, *Mtx*, 2, and *Mtx* 3, while *Mtx* 3 has a role in the formation of pores at midgut *Culex* larvae [20]. The research of Poopathi and Abidha was shown the Bin toxin of *Bs* and multiple toxin of *Bti* after being ingested into mosquito larvae midgut; their effects were shown disruption, separation, and ploughing of columnar epithelial cells midgut, which causing the death of mosquitoe larvae [21]. The study of Wirth *et al.*

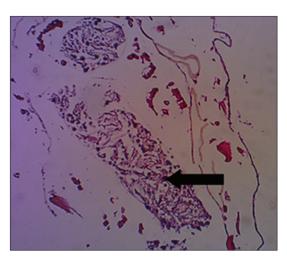


Fig. 7: Midgut of *Culex quinquefasciatus* larvae in combination treatment Group VI was (3:7) ppm (black arrow shown pores of midgut)

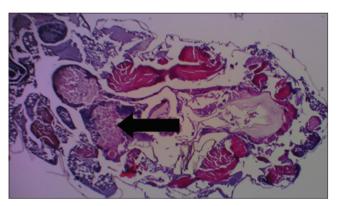


Fig. 8: Midgut of *Culex quinquefasciatus* larvae in treatment Group X single *Bacillus sphaericus* 2362 (black arrow shown pores of midgut)

Table 1: LC₅₀ of seven combination treatment group

Combination Bs 2362 with Bacillus thuringiensis subsp israelensis	LC ₅₀
I (8:2)	2.274
II (5:5)	9.193
III (7:3)	4.146
IV (6:4)	3.191
V (2:8)	2.276
VI (3:7)	3.122
VII (4:6)	2.279
Mean	3.783
Standard deviation	2.483

LC₅₀: Lethal concentration 50%, Bacillus sphaericus

combined the mixture of Bs 2362 with Mtx 2 and Cyt1Aa from Bacillus thuringiensis (8:1:1) was also synergistic with synergy factor at the LC_{50} value 44 for the resistant colonies C. quinquefasciatus to Bs 2362 (Bs-R), which combined have distinct mode of action when Cyt1Aa was lipophilic and lyses cell, while Mtx 2 was making pore at target cells, which the mixture toxins with both mechanism mode of action could suppress resistance to Bs 2362 (Bs-R) [22].

CONCLUSION

The combination of *Bs* 2362 and *Bti* was shown pores at *C. quinquefasciatus* larvae midgut and optimization ratio was shown in Group I with the concentration was (8:2) ppm.

ACKNOWLEDGMENT

First, the authors are thankful to High Education Department Grants. We are also thankful to Parasitology Department of Medicine Faculty of Gadjah Mada University and Maranatha Christian University.

REFERENCES

- Alcantara EP, Aguda RM, Curtiss A, Dean DH, Cohen MB. Bacillus thuringiensis delta-endotoxin binding to brush border membrane vesicles of rice stem borers. Arch Insect Biochem Physiol 2004;55(4):169-77.
- Akhurst RJ, James W, Bird LJ, Beard C. Resistance to the Cry1Ac delta-endotoxin of *Bacillus thuringiensis* in the cotton bollworm, *Helicoverpa armigera* (Lepidoptera: Noctuidae). J Econ Entomol 2003;96(4):1290-9.
- 3. Alzate O, Hemann CF, Osorio C, Hille R, Dean DH. Ser170 of *Bacillus thuringiensis* Cry1Ab delta-endotoxin becomes anchored in a hydrophobic moiety upon insertion of this protein into *Manduca sexta* brush border membranes. BMC Biochem 2009;10:25.
- Barber MJ, Quinn GB. High-level expression in *Escherichia coli* of the soluble, catalytic domain of rat hepatic cytochrome b5 reductase. Protein Expr Purif 1996;8(1):41-7.
- Barton KA, Whiteley HR, Yang NS. Bacillus thuringiensis section sign-endotoxin expressed in transgenic nicotiana tabacum provides resistance to lepidopteran insects. Plant Physiol 1987;85(4):1103-9.
- Wahyono TY, Purwantyastuti, Supali T. Filariasis di Indonesia. Kementerian Kesehatan Republik Indonesia: Pusat Data & Surveilans Epidemiologi; 2010. p. 1-20.
- Romao TP, de Melo Chalegre KD, Key S, Ayres CF, de Oliveira CM, Neto OP, et al. A second independent resistance mechanism to Bacillus sphaericus binary toxin targets its a-glucosidase receptor in Culex quinquefasciatus. FEBS J 2006;273(7):1556-68.
- Mittal PK. Biolarvacides in vector control: Challenges and prospect. J Vector Borne Dis 2003;40:20-32
- Margareth WC. Mosquito resistance to bacterial larvicidal toxins. Open Toxinol J 2010;3:126-40.
- Poopathi S, Brij Kishore T. The challenge of mosquito control strategies: From primodial to molecular approaches. Biotechnol Mol

- Biol Rev 2006;1(51-65):1538-2273.
- 11. Wirth MC, Walton WE, Federici BA. Evolution of resistance to the *Bacillus sphaericus* bin toxin is phenotypically masked by combination with the mosquitocidal proteins of *Bacillus thuringiensis* subspecies *Israeliensis*. Environ Microbiol 2010;12(5):1154-60.
- Bravo A, Gill SS, Soberón M. Mode of action of *Bacillus thuringiensis* cry and cyt toxins and their potential for insect control. Toxicon 2007;49(4):423-35.
- 13. Ben-Dov E. *Bacillus thuringiensis* subsp. *Israelensis* and its dipteran-specific toxins. Toxins (Basel) 2014;6(4):1222-43.
- Laurence D, Christophe L, Roger F. Using the bio-insecticide *Bacillus thuringiensis Israelensis* in mosquito control. In: Stoytcheva M, editor. Pesticides in the Modern World Pests Control and Pesticides Exposure and Toxicity Assessment. China, Shangha: InTech; 2011. p. 93-126.
- Wirth MC, Park HW, Walton WE, Federici BA. Cyt1A of *Bacillus thuringiensis* delays evolution of resistance to Cry11A in the mosquito Culex quinquefasciatus. Appl Environ Microbiol 2005;71(1):185-9.
- Wirth MC, Federici BA, Walton WE. Cyt1A from Bacillus thuringiensis synergizes activity of Bacillus sphaericus against Aedes aegypti (Diptera: Culicidae). Appl Environ Microbiol 2000;66 (3):1093-7.
- WHOPES. Guidelines for Laboratory and Field Testing of Mosquito Larvicides. Geneva, Switzerland: World Health Organization; 2005. p. 1-41.
- Gomez KA, dan Gomez AA, editor. Statistical Procedures for Agricultural Research. 2nd ed. New York; John Willey & Son; 1984.
- 19. Opota O, Gauthier NC, Doye A, Berry C, Gounon P, Lemichez E, et al. Bacillus sphaericus binary toxin elicits host cell autophagy as a response to intoxication. PLoS One 2011;6 (2):e14682.
- Vanlalhruaia SK, Gurusubramanian G. Bacillus sphaericus in the biological control of mosquito vector complex. Sci Vision 2011;11(2):61-71.
- Poopathi S, Abidha S. Mosquitocidal bacterial toxins (*Bacillus sphaericus* and *Bacillus thuringiensis serovar israelensis*): Mode of action, cytopathological effects and mechanism of resistance. J Physiol Pathophysiol 2010;1(3):22-38.
- Wirth MC, Yang Y, Walton WE, Federici BA, Berry C. Mtx toxins synergize *Bacillus sphaericus* and Cry11Aa against susceptible and insecticide-resistant *Culex quinquefasciatus* larvae. Appl Environ Microbiol 2007;73(19):6066-71.