ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH



## A REVIEW ON USE OF PLANT EXTRACTS FOR GOLD AND SILVER NANOPARTICLE SYNTHESIS AND THEIR POTENTIAL ACTIVITIES AGAINST FOOD PATHOGENS

## ELIZABATH ANTONY, SHYLAJA GUNASEKARAN, MYTHILI SATHIAVELU, SATHIAVELU ARUNACHALAM\*

Department of Biomedical Sciences, School of Bio-Sciences and Technology, VIT University, Vellore - 632 014, Tamil Nadu, India. Email: asathiavelu@vit.ac.in

#### Received: 01 March 2016, Revised and Accepted: 10 March 2016

## ABSTRACT

Metal nanoparticle synthesis using plant-mediated method is a highly explored field of nanotechnology. It has many advantages such as less time consumption and non-effective to human beings. Gold and silver nanoparticles (AuNPs and AgNPs) have been chosen for this study because of its wide range of applications and importance in different fields. This review explains different plant sources for the synthesis of AuNPs and AgNPs, and their antimicrobial activity against food pathogens which will provide information to future studies.

Keywords: Green synthesis, Gold nanoparticle, Silver nanoparticle, Antioxidant activity, Antimicrobial activity, Food pathogens.

## INTRODUCTION

Owing to the smaller particle size, various shapes, and increased surface area, nanoparticles exhibit different properties than their parent materials and are found to have many interesting applications particularly in biomedical science [1]. Due to their unique properties, it is applicable in medical, electronic, material sciences, pharmaceuticals, agriculture, catalyst, drug delivery, etc. Among all noble metal nanoparticles, silver and gold nanoparticles (AgNPs and AuNPs) are of great importance in the field of nanotechnology [2]. Nanoparticles are synthesized by physical, chemical, and biological or green methods. Various chemical and physical methods are proved to be quite expensive and potentially hazardous to the environments which involve the use of toxic and perilous chemicals that are responsible for various biological risks. This may be the reason for choosing biosynthesis of nanoparticles via green route that does not employ toxic chemicals and proved to be eco-friendly [3]. AgNPs are the most prototypical target of green methods [4-7] since it has many applications. AuNPs are also of considerable interest, although to a much lesser degree than AgNPs. Both the AgNPs and AuNPs have effect against animal pathogens [8]. AgNPs are used as antimicrobial agents in commercial medical and consumer products [9]. Gold, in both nanoparticle and ionic forms, has been studied for antibiotic activities. Food pathogens are highly disastrous to the food industry as they cause foodborne diseases and death. So, this review summarizes various plant extracts used for the green synthesis of AuNPs and AgNPs, their potential activity against food pathogens and other pharmacological activities.

# GREEN SYNTHESIS OF AUNPS AND AgNPS AND THEIR ANTIMICROBIAL ACTIVITY

Nanoparticles can be synthesized using plant extracts. Various plant parts such as root, stem, bark, leaf, and flowers can be used for the same. Some of the plant extracts used for the synthesis of nanoparticles and their activity against food pathogens were discussed and are as follows.

### AgNP synthesis using cauliflower extract

AgNPs using cauliflower extract was done by Sridhara *et al.* which showed the formation of AgNPs of average width 42-83 nm. The antibacterial effect of AgNPs was studied against two food pathogens: *Escherichia coli* and *Staphylococcus aureus* [10].

## AgNP synthesis using papaya fruit extract

AgNP synthesis using leaf extract of papaya fruit extract was reported by Jain *et al.*, where the antimicrobial activity of the extract was checked.

The characterization analysis, such as X-ray diffraction (XRD) and scanning electron microscope (SEM) analysis, showed the formation of AgNPs of size 15 nm. The resultant AgNPs showed potent activity against the common food borne pathogen *E. coli* [11].

## AgNP synthesis using Trianthema decandra root extract

AgNP synthesis using *T. decandra* root extract was performed, and its antibacterial activity was studied. The resultant AgNPs were found to be of average size of 10 nm by XRD and SEM analysis. The AgNP exhibited high activity against the food pathogen *E. coli* [12].

### AgNP using Moringa oleifera leaf extract

Green biosynthesis of AgNP using *M. oleifera* leaf extract was carried out by Das *et al.* The antimicrobial potential of AgNPs was systematically evaluated against two food pathogens: *E. coli* (ATCC 25922) and *S. aureus* (ATCC 25923). The higher activity was showed against *E. coli* than *S. aureus* [13].

## AgNP synthesis using Garcinia mangostana fruit extract

A recent study on the biosynthesis of AgNPs using *G. mangostana* fruit extract was reported by Rajakannu *et al.* The transmission electron microscope (TEM) analysis showed the formation of AgNPs of size 1-100 nm. The antibacterial, antioxidant activity of AgNP were evaluated. AgNPs were tested for antibacterial activity against two food pathogens such as *E. coli* and *S. aureus.* The biosynthesized AgNPs showed significantly higher antioxidant activity compared to *G. mangostana* fruit extract [14].

### AgNP synthesis using Withania somnifera leaf extract

The bioreduction of AgNP using *W. somnifera* reported by Raut *et al.* revealed the importance of blue light in the reduction of silver ions. TEM analysis showed the formation of nanoparticles of size ranging from 5 to 30 nm. The synthesized nanoparticle showed antimicrobial activity against food pathogens such as *S. aureus, E. coli, Candida albicans, Aspergillus niger,* and *Aspergillus flavus.* The potent activity was found to be against *S. aureus* and *A. niger* [15].

### AgNP synthesis using Alternanthera dentae aqueous extract

The green rapid syntheses of spherical shaped AgNPs were observed using *A. dentata* aqueous extract by Kumar *et al.* These AgNPs exhibit antibacterial activity against *Pseudomonas aeruginosa, E. coli, Klebsiella pneumonia,* and *Enterococcus faecal* [16].

## AgNP synthesis using Acorus calamus extract

*A. calamus* was also used for the synthesis of AgNPs to evaluate its antioxidant, antibacterial as well as anticancer effects was reported by Nakkala *et al.* The formation of AgNPs was confirmed by ultraviolet (UV)-visible spectroscopy and their average size was found to be 31.83 nm. The synthesized nanoparticles were found to have remarkable antibacterial activity against three different food borne pathogens such as *Bacillus subtilis, Bacillus cereus,* and *S. aureus* [17].

### AgNP synthesis using Ocimum tenuiflorum leaf extract

Bio inspired synthesis of AgNP using *O. tenuiflorum* leaf extract was carried out by Patil *et al.* TEM and power spectral density analysis showed that the size of AgNPs ranges from 25 to 40 nm. The antibacterial activities of formed AgNPs were checked against some of the food pathogens such as *E. coli* and *B. subtilis*. The maximum activity was found against *B. subtilis* [18].

## AgNP synthesis using tea leaf extract

The tea leaf mediated bio-synthesis of AgNP was reported by Sun *et al.* showed the formation of nanoparticles size ranges from 20 to 90 nm. The antibacterial activity of formed AgNPs was found to be active against *E. coli* [19].

### AgNP synthesis using Sensitivum portulacastrum callus extract

The ability of callus extract of *S. portulacastrum* Linn. to produce AgNPs was reported by Nabikhan *et al.* The resulted AgNPs showed size ranges from 5 to 20 nm. The fourier transform infrared spectroscopy (FTIR) analysis indicated the presence of the protein. The AgNPs were observed to have effective activity against clinical strains of bacterial and fungal food pathogens such as *S. aureus, Listeria monocytogenes,* and *Penicillium italicum* [20].

## AgNP synthesis using Tribulus terrestris fruit extract

The dried fruit body extract of the plant *T. terrestris* Linn. was reported to be used as a capping agent in AgNP synthesis by Gopinath *et al.* AgNPs ranged in size from 16 to 28 nm. The extracts possess antibacterial property against multi-drug resistant food pathogens such as *B. subtilis, E. coli,* and *S. aureus. E. coli* showed a maximum zone of inhibition [21].

### AgNP synthesis using Cocos nucifera inflorescence extract

AgNP was synthesized using inflorescence extracts of the tree *C. nucifera* was reported by Mariselvam *et al.* TEM anaslysis showed the formation of AgNP with the size of 22 nm. It showed significant antimicrobial activity against food pathogens such as *Salmonella paratyphi and B. subtilis.* The maximum activity was found against *S. paratyphi* [22].

## AgNP synthesis using Abutilon indicum extract

Ashokkumar *et al.* reported the formation of AgNP ranges from 7 to 17 nm in size using the extract of *A. indicum*. The AgNPs thus obtained showed highly potent antibacterial activity against some of the most dangerous food pathogens such as *S. aureus*, *B. subtilis*, *Salmonella typhi*, and *E. coli*. The maximum activity was showed against *B. subtilis* and *E. coli* [23].

### AgNP synthesis using Allium cepa extract

Biosynthesis of AgNPs using *A. cepa* was carried out by Saxena *et al.*, and it was found to have the particle size of 33.6 nm. The synthesized nanoparticle showed excellent antimicrobial activity against *E. coli* and *Salmonella typhimurium* [24].

### AgNP synthesis using Lantana camara fruit extract

AgNP synthesis using *L. camara* fruit extract was done by Sivakumar *et al.* The spherical shaped AgNPs having size between 12.55 and 12.99 nm was confirmed by TEM analysis. The resulted AgNPs showed excellent antibacterial activity against some of the common food pathogens such as *B. subtilis* MTCC 1133, *S. aureus* MTCC 96, and *S. typhi* MTCC 733. The maximum activity was found to be 26 mm zone of inhibition for *B. subtilis* MTCC 1133 [25].

### AgNP synthesis using Cymbopogon citratus leaf extract

*C. citratus*, (commonly known as lemon grass) a native aromatic herb of India, was used by Geetha *et al.* for the synthesis of AgNPs of size 32 nm. The formed AgNPs showed a strong antibacterial effect against food pathogens such as *E. coli, Shigella flexneri*, and *S. Sonnei* [26].

### AgNP synthesis using Acalypha indica leaf extract

AgNPs synthesize from the leaf extract of *A. indica* was performed by Krishnaraj *et al.* The TEM analysis confirmed the formation of AgNPs ranging in size from 20 to 30 nm. The antibacterial activity of synthesized AgNPs showed effective inhibitory activity against food pathogens *E. coli* and *Vibrio cholera* [27].

## AgNP synthesis using Boerhaavia diffusa plant extract

*B. diffusa* plant extract was used as a reducing agent by Nakkala *et al.* for the green synthesis of AgNPs. The characterization analysis XRD and TEM revealed an average particle size of 25 nm. These nanoparticles were tested for antibacterial activity against three fish pathogens such as *Pseudomonas fluorescens, Aeromonas hydrophila,* and *Flavobacterium branchiophilum* and showed the highest activity against *Flavobacterium* [28].

### AgNP synthesis using Euphorbia hirta leaf extract

AgNP synthesis using leaf extract of *E. hirta* was performed by Elumalai *et al.* The size of the synthesized AgNPs ranges from 40 to 50 nm, and it shows antimicrobial activity against the potent food pathogen *B. cereus* [29].

### AgNP synthesis using Morinda citrifolia extract

Phytosynthesis of AgNP using *M. citrifolia* Linn. was reported by Sathishkumar *et al.* SEM analysis confirmed the synthesis of nanoparticles of size 10-60 nm in diameter with an average size of 27 nm. The resulted AgNPs showed potent activity against food pathogens such as *B. cereus* and *E. coli. E. coli* showed maximum sensitivity against the formed nanoparticles [30].

## AgNP synthesis using banana peel extract

The recent study reports by Ibrahim on the synthesis of AgNPs using banana peel extract as a reducing and capping agent. Typical nanoparticle characterization techniques showed the formation of the nanoparticle of size 23.7 nm. AgNPs showed effective antibacterial activity against representative food pathogens such as *B. subtilis, E. coli*, and *S. aureus*. Out of these organisms, *E. coli* showed maximum inhibitory zone [31].

### AgNP synthesis using Cassia roxburghii plant extract

Biosynthesis characterization of AgNPs using *C. roxburghii* was carried out by Balashanmugam and Kalaichelvan, which synthesized nanoparticles of size 10-30 nm. The aqueous extract was recorded to have potent antibacterial activity. AgNPs were evaluated for their antibacterial activity against four different pathogenic food borne bacteria *B. subtilis, S. aureus, E. coli,* and *Enterobacter aerogenes. B. subtilis* showed high sensitivity toward AgNPs [32].

## AgNP synthesis using peroxidase from Euphorbia extract

Green synthesis of AgNPs using peroxidase from *Euphorbia* was done by Cicek *et al.* to analyze its antibacterial activity. The synthesized AgNPs were characterized using SEM, XRD, which yield particles size ranges from 7 to 20 nm. Its antibacterial activity was checked against some food pathogens, viz., *Serratia marcescens, Yersinia pseudotuberculosis, S. aureus, S. typhimurium, L. monocytogenes,* and *E. coli. Y. pseudotuberculosis* and *S. typhimurium* showed high activity [33].

## AuNP synthesis using A. calamus rhizome extract

AuNPs were synthesized using herbal *A. calamus* rhizome extract was reported by Ganesan and Gurumallesh Prabu, and it was coating on cotton fabric for checking its antibacterial activity against *S. aureus* and *E. coli*. The resulted AuNP was found to be having a particle size below 100 nm. The extract containing AuNPs coated cotton fabric had higher antibacterial activity than other test samples against *E. coli* [34].

## AgNP synthesis using leaf extract of Coleus aromaticus

*C. aromaticus* leaf extract was used as a capping agent for the synthesis of AgNPs by Vanaja *and* Annadurai. SEM analysis showed the formation of nanoparticles of size 40-50 nm. The antimicrobial activity of thus formed AgNP was checked against a food pathogen *B. subtilis*, which showed excellent result [35].

### AuNP synthesis using Artocarpus heterophyllus fruit extract

One-step synthesis of highly-biocompatible spherical AuNPs of size 20-25 nm using *A. heterophyllus* Lam. (jackfruit) fruit extract was done to check its effect on pathogens by Basavegowda *et al.* AuNP thus synthesized from *A. heterophyllus* Lam. fruit extract was found to be effective against *E. coli* [36].

## AgNP synthesis using *Musa balbisiana*, *A. indica*, and *O. tenuiflorum* plant extracts

The green synthesis of AgNP using three plants, viz., *M. balbisiana* (banana), *A. indica* (neem), and *O. tenuiflorum* (black tulsi) was reported by Banerjee *et al.* SEM analysis confirmed the formation of nanoparticles of size 200 nm, and its antibacterial effect against *Bacillus* and *E. coli* was checked. *Bacillus* showed maximum sensitivity against the nanoparticle [37].

### AuNP synthesis using Nepenthes khasiana leaf extract

Green synthesis of AuNPs from the leaf extract of *N. khasiana* was reported by Bhau *et al.*, and its antimicrobial assay was done. The synthesized AuNPs were characterized by UV-visible spectroscopy, SEM, X-ray diffraction, FTIR, and TEM, which show the formation of the nanoparticle of size 50-80 nm. The antimicrobial property of the AuNPs was checked against food pathogens such as *Bacillus*, *E. coli*, and *A. niger*. *A. niger* showed high sensitivity against AgNPs [38].

### AgNP synthesis using Argemone mexicana leaf extract

Biomediated synthesis of AgNPs was carried out by Singh *et al.* using the leaf extract of *A. mexicana*. SEM analysis showed the average particle size of 30 nm. It showed antifungal activity against *A. flavus*, a food pathogen [39].

## AgNP synthesis using Aloe vera leaf extract

Biosynthesis of AgNPs was done using *A. vera* leaf extract by Medda *et al.*, and its antifungal activity was checked against two food spoiling pathogens *Aspergillus* and *Rhizopus*. The average size of an individual particle was estimated to be 70 nm by SEM analysis. The resultant nanoparticles showed high activity against *Aspergillus sp* [40].

## AgNP synthesis using Crataegus douglasii extract

Plant-mediated green synthesis of AgNP and its antibacterial activity using *C. douglasii* was reported by Ghaffari-Moghaddam and Hadi-Dabanlou SEM analysis confirmed the formation of nanoparticles of size 29.28 nm. Antibacterial activity of resultant nanoparticles was checked against food pathogens *E. coli* and *S. aureus*. The assay showed the potent activity of nanoparticle against both *E. coli* and *S. aureus* [41].

## AgNP synthesis using Afzelia quanzensis bark extract

*A. quanzensis* bark extract was used by Singh *et al.* for the green synthesis of AgNPs and to study their antibacterial activity. SEM analysis confirmed the formation of nanoparticles of size 10-80 nm. The antimicrobial activity of synthesized nanoparticle was checked against *E. coli* and *S. aureus*. The AgNP was more active against *E. coli* [42].

## AgNP synthesis using Eucalyptus globulus extract

Microwave accelerated green synthesis of AgNPs from *E. globulus* was recently reported by Ali *et al.* The characterization techniques yield AgNP of size ranges from 1.9 to 4.3 nm and 5-25 nm. The antimicrobial activity was checked against food pathogens such as *E. coli* and *S. aureus. S. aureus* showed maximum sensitivity [43].

## AgNP synthesis using olive leaf extract

Green synthesis of AgNPs using olive leaf extract and its antibacterial activity was reported by Khalil *et al.* SEM analysis showed the formation of nanoparticles of size 20-25 nm. The antimicrobial activity of resultant nanoparticles was checked against food pathogens *E. coli* and *S. aureus.* The antimicrobial assay proved potent activity against both the pathogens [44].

## AgNP synthesis using Origanum vulgare extract

*O. vulgare* mediated AgNP synthesis was reported by Sankar *et al.* SEM analysis showed particles of size 63-85 nm. The resultant particles showed antibacterial activity against *E. coli, S. dysenteriae, S. typhi,* and *Shigella sonnei* [45].

## AgNP synthesis using Pulicaria glutinosa extract

Green synthesis of AgNPs mediated by *P glutinosa* extract was done by Khan *et al.* and its antibacterial activity was also checked. TEM analysis confirmed the synthesis of nanoparticles of an average size ranges from 40 to 60 nm. Antibacterial activity showed excellent results against food borne pathogens *E. coli* and *S. aureus* [46].

## AgNP synthesis using tuber extract

Tuber extract mediated biosynthesis of AgNPs is a recent nanotech innovative work done by Sivakumar *et al.* The prepared materials were characterized by UV-visible spectroscopy, XRD, and FTIR. TEM analysis showed the formation of AgNPs of size ranges from 4.77 to 5.6 nm. Green synthesized AgNPs exhibited strong antioxidant and effective antibacterial activity against food pathogens such as *B. subtilis, E. coli*, *S. typhi*, and *S. aureus*. Maximum activity was found against *E. coli* [47].

### AgNP synthesis using Rhinacanthus nasutus leaf extract

Leaf extract of *R. nasutus* was used by Pasupuleti *et al.* for the green routed AgNP synthesis. Characterization technique like TEM confirmed the formation of AgNPs of size <22 nm. The *in vitro* antimicrobial activity of the AgNPs synthesized using *R. nasutus* leaf extract was investigated against some common food pathogens such as *B. subtilis, S. aureus, E. coli, A. niger*, and *A. flavus*. The nanoparticle showed maximum activity against gram positive *S. aureus* and two fungi *A. niger* and *A. flavus* [48].

## AuNP synthesis using Pistacia integerrima gall extract

*P. integerrima* gall extract was used for green synthesis of AuNPs and its biological activities were analyzed by Islam *et al.* SEM analysis showed the formation of nanoparticles of size ranges from 20 to 200 nm. The synthesized AuNPs showed antibacterial and antifungal activities against some of food pathogens such as *B. subtilis, S. aureus, A. niger,* and *A. flavus* [49].

## AgNP synthesis using Petroselinum crispum leaf extract

Plant-mediated synthesis of AgNPs using *P. crispum* (parsley) leaf extract was reported by Roy *et al.* Spectral analysis and antibacterial activity was also checked. TEM analysis showed the formation of nanoparticles with the size 30-32 nm. The antibacterial analysis was done against food pathogens such as *E. coli* and *S. aureus.* AgNPs showed maximum activity against *E. coli* [50].

## AgNP synthesis using Dioscorea bulbifera tuber extract

Synthesis of AgNPs using *D. bulbifera* tuber extract was reported by Ghosh *et al.*, and its antimicrobial activity was also reported. TEM analysis confirmed the formation of nanoparticles of size ranges from 8 to 20 nm. The resulting AgNPs were found to possess potent antibacterial activity against pathogenic food borne organisms such as *E. coli, B. subtilis, S. typhi*, and *S. aureus. E. coli* showed maximum sensitivity against AgNPs [51].

### AuNP synthesis using Ananas comosus fruit extract

Plant-mediated synthesis of AuNP using fruit extract of *A. comosus* plant confirmed the formation of nanoparticles by Basavegowda *et al.* The

resulted particles were of size 20 nm. Its activity was checked against some of the common food pathogens such as *A. niger, A. flavus,* and *E. coli. E. coli* showed positive result [52].

## AuNP synthesis using Zingiber officinale root extract

Green synthesis of AuNPs using *Z. officinale* root extract was done by Velmurugan *et al.*, which resulted the formation of nanoparticles of size 10-20 nm. The resultant nanoparticles were used to check antimicrobial activity against some of the food pathogens such as *Staphylococcus* spp., *Listeria* spp., and *Bacillus* spp. *Staphylococcus* and *Listeria* spp. showed maximum activity [53].

## AgNP synthesis using Mangosteen leaf extract

Biosynthesis of AgNPs using *Mangosteen* leaf extract and evaluation of their antimicrobial activity was carried out by Veerasamy *et al.* Characterization analysis, SEM showed the formation of nanoparticles of size 35 nm. Antimicrobial activity was checked against food borne pathogens such as *E. coli* and *S. aureus. S. aureus* showed positive result for the antimicrobial assay [54].

### AuNP synthesis using Diospyros ferrea extract

AuNP synthesis using *D. ferrea* was reported by Ramesh and Armash. SEM analysis confirmed the formation of nanoparticle of average size 70-90 nm. Its activity was checked against the food pathogen *B. cereus,* and it showed the positive result [55].

## AuNP synthesis using Mentha piperita extract

*M. piperita* extract was used by Mubarak Ali *et al.* to synthesis spherical shaped AuNPs with size around 150 nm and showed antimicrobial activities against the two common food pathogens *S. aureus* and *E. coli* [8].

### AgNP synthesis using aqueous bark extract of Cinnamon zeylanicum

The aqueous bark extract of *C. zeylanicum* was used as a green agent by Sathishkumar *et al.* to synthesis AgNPs and its bactericidal activity was also checked. TEM and XRD analysis confirmed the formation of nanoparticles of size 31-40 nm. Its antibacterial activity was checked against the most common food pathogen *E. coli* [56].

## AgNP synthesis using Piper longum fruit extract

*P. longum* fruit extract was used by Reddy *et al.* to synthesis AgNPs. Characterization analysis, such as FTIR and SEM, was done and the analysis confirmed the formation of nanoparticles of size 40-70 nm. The formed AgNP showed potent activity against food pathogens such as *B. cereus*, *B. subtilis*, and *S. aureus* [57].

### AuNP synthesis using Abelmoschus esculentus seed extract

The aqueous seed extract of *A. esculentus* was used as a capping agent by Jayaseelan *et al.* to synthesize AuNPs of size 45-75 nm and its antifungal activities were tested against *Puccinia graminis tritici, A. flavus, A. niger,* and *C. albicans.* Resultant AuNPs showed maximum activity against *C. albicans* [58].

#### AgNP synthesis using *Cajanus cajan* leaf extract

AgNP synthesis using *C. cajan* leaf extract was reported by Nagati *et al.* TEM analysis confirmed the formation of nanoparticle of size 5-60 nm. Antibacterial activity of resultant nanoparticles was checked against *E. coli* and *S. aureus. S. aureus* showed highly sensitive against nanoparticles [59].

### AgNP synthesis using Dolichos biflorus seed extract

Phytofabrication of AgNPs using *D. biflorus* Linn. (Horse Gram) seed extract, and its bactericidal and antioxidant activities were recently reported by Vijayalakshmi *et al.* The AgNPs exhibited antibacterial efficacy against different human pathogens [60].

## APPLICATIONS OF AgNPs AND AuNPs

Nanotechnology applications are highly useful for biological molecules because of their unique properties. It is a growing field of material science and biological science [61]. AgNPs have broad applications in diverse areas such as integrated circuits [62], sensors [63], biolabeling filters, antimicrobial deodorant fibers [64], cell electrodes [65], and low cost paper batteries [66]. It also has important role in health industry, food industry, textile coatings, and environmental applications [67]. The use of AuNPs has found application in analytical methods such as colorimetric techniques for the determination of heavy metal ions in aqueous solutions [68]. The catalytic activity of AuNPs makes them use in water gas shift reactions [69-71]. They are also used in the field of sensors [72,73]. AuNPs are used in the development of biosensors and also for DNA labeling [74,75]. Spherical AuNPs are used to develop functional electrical coatings [76,77].

## CONCLUSION

Plants or their extracts can be used in the synthesis of AuNPs and AgNPs as a green route. Such nanoparticles produced using plants have been used in various applications for human benefit. Elucidation of the mechanism of plant-mediated synthesis of nanoparticles is a very promising area of research. Therefore, the information compiled in this review might be useful in elucidating the mechanism of nanoparticle synthesis using plants as well as opening way for exploring other plants for this purpose.

### REFERENCES

- Iravani S. Green synthesis of metal nanoparticles using plants. Green Chem 2011;13(10):2638-50.
- Ahmad A, Mukherjee P, Senapati S, Mandal D, Khan MI, Kumar R, et al. Extracellular biosynthesis of silver nanoparticles using the fungus Fusarium oxysporum. Colloids Surf B Biointerfaces 2003;28:313-8.
- Reddy GA, Joy JM, Mitra T, Shabnam S, Shilpa T. Nano silver A review. Int J Adv Pharm 2012;1(2):9-15.
- Hu B. Microwave assisted rapid facile green synthesis of uniform silver nanoparticles: Self-assembly into multi-layered films and their optical properties. J Phys Chem C 2008;112(30):11169-74.
- Sankar S, Valli Nachiyar C. Microbial synthesis and characterization of silver nanoparticles using endophytic bacterium *Bacillus cereus*: A novel source in benign synthesis. Glob J Med Res 2012;12:43-9.
- Darroudi M. Green synthesis of colloidal silver nanoparticle by sonochemical method. Adv Mater Lett 2012;66:117-20.
- Subba Rao Y, Kotakadi VS, Prasad TN, Reddy AV, Sai Gopal DV. Green synthesis and spectral characterization of silver nanoparticles from Lakshmi *Tulasi* (*Ocimum sanctum*) leaf extract. Spectrochim Acta A Mol Biomol Spectrosc 2013;103:156-9.
- MubarakAli D, Thajuddin N, Jeganathan K, Gunasekaran M. Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. Colloids Surf B Biointerfaces 2011;85(2):360-5.
- Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. Biotechnol Adv 2009;27(1):76-83.
- Sridhara V, Pratimai K, Krishnamurthy G, Sreekanth B. Vegetable assisted synthesis of silver nanoparticles and its antibacterial activity against two human pathogens. Asian J Pharm Clin Res 2013;6(2):53-7.
- Jain D, Daima HK, Kachwaha S, Kothari SL. Synthesis of plant mediated silver nanoparticle using papaya fruit extract and evaluation of their microbial activities. Digest J Nanomater Biostruct 2009;4(3):557-63.
- Geethalakshmi R, Sarada DV. Synthesis of plant-mediated silver nanoparticles using *Trianthema decandra* extract and evaluation of their anti-microbial activities. Int J Eng Sci Technol 2010;2(5):970-5.
- Das S, Kumar U, Parida K, Bindhani BK. Green biosynthesis of silver nanoparticles using *Moringa oleifera* L. leaf. Int J Nanotechnol Appl 2013;3(2):51-62.
- Rajakannu S, Shankar S, Perumal S, Subramanian S, Dhakshinamoorthy PG. Biosynthesis of silver nanoparticles using *Garcinia mangostana* fruit extract and their antibacterial, antioxidant activity. Int J Curr Microbiol Appl Sci 2015;4(1):944-52.
- Raut RW, Mendhulkar VD, Kashid SB. Photosensitized synthesis of silver nanoparticles using *Withania somnifera* leaf powder and silver nitrate. J Photochem Photobiol B 2014;132:45-55.
- Kumar DA, Palanichamy V, Roopan SM. Green synthesis of silver nanoparticles using *Alternanthera* dentata leaf extract at room temperature and their antimicrobial activity. Spectrochim Acta A Mol Biomol Spectrosc 2014;127:168-71.

- Nakkala JR, Mata R, Gupta AK, Sadras SR. Biological activities of green silver nanoparticles synthesized with *Acorous calamus* rhizome extract. Eur J Med Chem 2014;85:784-94.
- Patil RS, Kokate MR, Kolekar SS. Bioinspired synthesis of highly stabilized silver nanoparticles using *Ocimum tenuiflorum* leaf extract and their antibacterial activity. Spectrochim Acta A Mol Biomol Spectrosc 2012;91:234-8.
- Sun Q, Cai X, Li J, Zheng M, Chen Z, Chang-Ping Y. Green synthesis of silver nanoparticles using tea leaf extract and evaluation of their stability and antibacterial activity. Colloids Surf A Physicochem Eng Aspect 2014;444:226-31.
- Nabikhan A, Kandasamy K, Raj A, Alikunhi NM. Synthesis of antimicrobial silver nanoparticles by callus and leaf extracts from saltmarsh plant, *Sesuvium portulacastrum* L. Colloids Surf B Biointerfaces 2010;79(2):488-93.
- Gopinath V, MubarakAli D, Priyadarshini S, Priyadharsshini NM, Thajuddin N, Velusamy P. Biosynthesis of silver nanoparticles from *Tribulus terrestris* and its antimicrobial activity: A novel biological approach. Colloids Surf B Biointerfaces 2012;96:69-74.
- 22. Mariselvam R, Ranjitsingh AJ, Usha Raja Nanthini A, Kalirajan K, Padmalatha C, Mosae Selvakumar P. Green synthesis of silver nanoparticles from the extract of the inflorescence of *Cocos nucifera* (Family: *Arecaceae*) for enhanced antibacterial activity. Spectrochim Acta A Mol Biomol Spectrosc 2014;129:537-41.
- Ashokkumar S, Ravi S, Kathiravan V, Velmurugan S. Synthesis of silver nanoparticles using A. Indicum leaf extract and their antibacterial activity. Spectrochim Acta A Mol Biomol Spectrosc 2015;134:34-9.
- Saxena A, Tripathi RM, Singh RP. Biological synthesis of silver nanoaprticles by Onion (*Allium cepa*) extract and their antibacterial activity. Digest J Nanomat Biostruct 2010;5(2):427-32.
- Sivakumar P, Nethradevi C, Renganathan S. Synthesis of silver nanoparticles using *Lantana camara* fruit extract and its effect on pathogens. Asian J Pharm Clin Res 2012;5(3):97-101.
- Geetha N, Geetha TS, Manonmani P, Thiyagarajan M. Green synthesis of silver nanoparticles using *Cymbopogan Citratus* (Dc) Stapf. Extract and its antibacterial activity. Aust J Basic Appl Sci 2014;8(3):324-31.
- Krishnaraj C, Jagan EG, Rajasekar S, Selvakumar P, Kalaichelvan PT, Mohan N. Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. Colloids Surf B Biointerfaces 2010;76:50-6.
- Nakkala JR, Mata R, Gupta SR, Sadras SR. Green synthesis and characterization of silver nanoparticles using *Boerhaavia diffusa*. Ind Crop Prod 2014;76:562-6.
- Elumalai EK, Prasad TN, Hemachandran J, Therasa SV, Thirumalai T, David E. Extracellular synthesis of silver nanoparticles using leaves of *Euphorbia hirta* and their antibacterial activities. J Pharm Sci Res 2010;2(9):549-54.
- Sathishkumar G, Gobinatha C, Karpagam K, Hemamalini V, Premkumar K, Sivaramakrishnan S. Phyto-synthesis of silver nanoscale particles using *Morinda citrifolia* L. and its inhibitory activity against human pathogens. Colloids Surf B Biointefaces 2012;95:235-40.
- Ibrahim HM. Green synthesis and characterization of silver nanoparticles using banana peel extract and their antimicrobial activity against representative microorganisms. J Rad Res Appl Sci 2015;8:265-75.
- Balashanmugam P, Kalaichelvan PT. Biosynthesis characterization of silver nanoparticles using *Cassia roxburghii* DC. Aqueous extract, and coated on cotton cloth for effective antibacterial activity. Int J Nanomedicine 2015;10 Suppl 1:87-97.
- Cicek S, Gungor AA, Ahmet A, Nadaroglu H. Biochemical evaluation and green synthesis of nano silver using peroxidase from *Euphorbia* (*Euphorbia amygdaloides*) and its antibacterial activity. J Chem 2015;2015:Article ID: 486948, 7.
- Ganesan RM, Gurumallesh Prabu H. Synthesis of gold nanoparticles using herbal *Acorus calamus* rhizome extract and coating on cotton fabric for antibacterial and UV blocking applications. Arabian J Chem 2014;85:784-94.
- Vanaja M, Annadurai G. *Coleus aromaticus* leaf extract mediated synthesis of silver nanoparticles and its bactericidal activity. J Appl Nanosci 2013;3(3):217-23.
- Basavegowda N, Kumar GD, Tyliszczak B, Wzorek Z, Sobczak-Kupiec Z. One-step synthesis of highly-biocompatible spherical gold nanoparticles using *Artocarpus heterophyllus Lam*. (jackfruit) fruit extract and its effect on pathogens. Ann Agric Environ Med 2015;22(1):84-9.
- 37. Banerjee P, Satapathy M, Mukhopahayay A, Das P. Leaf extract mediated green synthesis of silver nanoparticles from widely available

Indian plants: Synthesis, characterization, antimicrobial property and toxicity analysis. Biores Bioproc A Springer Open J 2014;1:3.

- Bhau BS, Ghosh S, Puri S, Borah B, Sarmah DK, Khan R. Green synthesis of gold nanoparticles from the leaf extract of *Nepenthes khasiana* and antimicrobial assay. Adv Mater Lett 2015;6(1):55-8.
- Singh A, Jain D, Upadhyay MK, Khandelwala N, Verma HN. Green synthesis of silver nanoparticles using *Argemone Mexicana* leaf extract and evaluation of their antimicrobial activities. Digest J Nanomat Biostruct 2010;5(2):483-9.
- Medda S, Hajra A, Deb U, Bose P, Mondal NK. Biosynthesis of silver nanoparticles from *Aloe vera* leaf extract and antifungal activity against *Rhizopus* sp and *Aspergillus* sp. Appl Nanosci 2015;5:875-80.
- Ghaffari-Moghaddam M, Hadi-Dabanlou R. Plant mediated green synthesis and antibacterial activity of silver nanoparticles using *Cratageous douglasii* fruit extract. J Ind Eng Chem 2014;20:739-44.
- 42. Singh K, Panghal M, Kadyan S, Chaudhary U, Yadav JP. Green silver nanoparticles of *Phyllanthus amarus*: As an antibacterial agent against multi drug resistant clinical isolates of *Pseudomonas aeruginosa*. J Nanobiotechnology 2014;12:40.
- 43. Ali K, Ahmed B, Dwivedi S, Saquib Q, Al-Khedhairy AA, Musarrat J. Microwave accelerated green synthesis of stable silver nanoparticles with *Eucalyptus globulus* leaf extract and their antibacterial and antibiofilm activity on clinical isolates. PLoS One 2015;10(7):e0131178.
- 44. Khalil MM, Ismail EH, Khaled Z. Green synthesis of silver nanoparticles using olive leaf extract and its antibacterial activity. Arabian J Chem 2014;7(6):1131-9.
- Sankar R, Karthik A, Prabu A, Karthik S, Shivashangari KS, Ravikumar V. *Origanum vulgare* mediated biosynthesis of silver nanoparticles for its antibacterial and anticancer activity. Colloids Surf B Biointerfaces 2013;108:80-4.
- 46. Khan M, Khan ST, Khan M, Adil SF, Musarrat J, Al-Khedhairy AA, et al. Antibacterial properties of silver nanoparticles synthesized using *Pulicaria glutinosa* plant extract as a green bioreductant. Int J Nanomedicine 2014;9:3551-65.
- Sivakumar T, Gajalakshmi D, Subramanian VK, Palanisamy K. Tuber extract mediated biosynthesis of silver nanoparticles and its antioxidant, antibacterial activity. J Biol Sci 2015;15(2):68-77.
- Pasupuleti VR, Prasad TN, Shiekh RA, Balam SK, Narasimhulu G, Reddy CS, *et al.* Biogenic silver nanoparticles using *Rhinacanthus nasutus* leaf extract: Synthesis, spectral analysis, and antimicrobial studies. Int J Nanomed 2013;8(1):3355-65.
- 49. Islam NU, Jalil K, Shahid M, Muhammad N, Rauf A. *Pistacia integerrima* gall extract mediated green synthesis of gold nanoparticles and their biological activities. Arabian J Chem 2015; Available from: http://www.dx.doi.org/10.1016/j.arabjc.2015.02.014.
- Roy K, Sarkar CK, Ghosh CK. Plant mediated synthesis of silver nanoparticles using Parsely (*Petroselinum crispum*) leaf extract: Spectral analysis of the particles and antibacterial study. Appl Nanosci 2014;5:945-51.
- Ghosh S, Patil S, Ahire M, Kitture R, Kale S, Pardesi K, et al. Synthesis of silver nanoparticles using *Dioscorea bulbifera* tuber extract and evaluation of its synergistic potential in combination with antimicrobial agents. Int J Nanomed 2012;7:483-96.
- 52. Basavegowda N, Sobczak-Kupiec A, Malina D, Yathirajan HS, Keerthi VR, Chandrashekar N, *et al.* Plant mediated synthesis of gold nanoparticles using fruit extracts of *Ananas comosus(L)* (Pineapple) and evaluation of biological activities. Adv Mater Lett 2013;4(5):332-7.
- Velmurugan P, Anbalagan K, Manosathyadevan M, Lee KJ, Cho M, Lee SM, *et al.* Green synthesis of silver and gold nanoparticles using *Zingiber officinale* root extract and antibacterial activity of silver nanoparticles against food pathogens. Bioproc Biosys Eng 2014;37:1935-43.
- Veerasamy R, Xin TZ, Gunasagaran S, Xiang TF, Yang EF, Jeyakumar N, et al. Biosynthesis of silver nanoparticles using *Mangosteen* leaf extract and evaluation of their antimicrobial activities. J Saudi Chem Soc 2011;15:113-20.
- Ramesh V, Armash A. Green synthesis of gold nanoparticles against pathogens and cancer cells. Int J Pharm Res 2015;5(10):1757-75.
- 56. Sathishkumar M, Sneha K, Won SW, Cho CW, Kim S, Yun YS. Cinnamon zeylanicum bark extract and powder mediated green synthesis of nano-crystalline silver particles and its bactericidal activity. Colloids Surf B Biointerfaces 2009;73(2):332-8.
- Reddy NJ, Nagoor Vali D, Rani M, Rani SS. Evaluation of antioxidant, antibacterial and cytotoxic effects of green synthesized silver nanoparticles by *Piper longum* fruit. Mater Sci Eng C Mater Biol Appl 2014;34:115-22.
- 58. Jayaseelan C, Ramkumar, Abdul A, Perumal P. Green synthesis of gold

nanoparticles using seed aqueous extract of *Abelmoschus esculentus* and its antifungal activity. Ind Crop Prod 2013;45:423-9.

- Nagati V, Koyyati R, Donda MR, Alwala J, Kundle KR, Padigya PR. Green Synthesis and characterization of silver nanoparticles from *Cajanus cajan* leaf extract and its antibacterial activity. Int J Nanomater Biostruct 2012;2(3):39-43.
- 60. Vijayalakshmi M, Rameshkumar G, Rajagopal T, Thangapandian V, Ponmanickam P. Phytofabrication of silver nanoparticles using Horse gram (*Dolichos biflorus L.*) seed extract and assessment of its bactericidal and antioxidant activities. Thai J Pharm Sci 2015;39(4):149-55.
- Kaviya S, Santhanalakshmi J, Viswanathan B, Muthumary J, Srinivasan K. Biosynthesis of silver nanoparticles using citrus sinensis peel extract and its antibacterial activity. Spectrochim Acta A Mol Biomol Spectrosc 2011;79(3):594-8.
- Kasthuri J, Veerapandian S, Rajendiran N. Biological synthesis of silver and gold nanoparticles using Apiin as reducing agent. Colloids Surf B Biointerfaces 2009;68(1):55-60.
- Kotthaus SG, Hang BH, Schafer H. Study of isotropically conductive bondings filled with aggregates of nano-sited Ag-particles. IEEE Trans Comp Packaging Technol 1997;20:15-20.
- Zhang WW. Research and development for antibacterial materials of silver nanoparticle. New Chem Mater 2003;31:42-4.
- Bhattacharya D, Gupta RK. Nanotechnology and potential of microorganisms. Crit Rev Biotechnol 2005;25(4):199-204.
- Hong KH, Park JL, Sul H, Youk JH, Kang TJ. Preparation of antimicrobial poly (Vinyl alcohol) nanofibers containing silver nanoparticles. J Polym Sci Part B Polym Phys 2006;44:2468-72.
- 67. Cho KH, Park JE, Osaka T, Park SG. The study of antimicrobial activity

and preservative effects of nanosilver ingredient. Electrochim Acta 2005;51:956-60.

- Duran N, Marcato DP, De Souza HI, Alves LO, Espsito E. Antibacterial effect of silver nanoparticles produced by fungal process on textile fabrics and their effluent treatment. J Biomed Nanotechnol 2007;3:203-8.
- 69. Armendariz V, Gardea-Torresdey JL, Jose-Yacaman M, Gonzalez J, Herrera I, Parsons JG. Gold nanoparticles formation by oat and wheat biomasses. In: Proceedings – Waste Research Technology Conference at the Kansas City: Mariott-Country Club Plaza; 2002.
- Andreeva D. Low temperature water gas shift over gold catalysts. Gold Bull 2002;35:82-8.
- Grisel R, Weststrate KJ, Gluhoi A, Nieuwenhuys BE. Catalysis by gold nanoparticles. Gold Bull 2002;35:39-45.
- Hutchings GJ, Haruta M. A golden age of catalysis: A perspective. Appl Catal A 2005;291:2-5.
- Yáñez-Sedeño P, Pingarrón JM. Gold nanoparticle-based electrochemical biosensors. Anal Bioanal Chem 2005;382(4):884-6.
- Liu J, Lu Y. Colorimetric biosensors based on DNA enzyme-assembled gold nanoparticles. J Fluoresc 2004;14:343-54.
- Groning R, Breitkreutz J, Baroth V, Muller RS. Nanoparticles inplant extracts: Factors which influence the formation of nanoparticles in black tea infusions. Pharmazie 2001;56(10):790-2.
- Tang D, Yuan R, Chai Y. Ligand-functionalized core/shell Ag@Au nanoparticles label-free amperometric immun-biosensor. Biotechnol Bioeng 2006;94(5):996-1004.
- Singh A, Chaudhary M, Sastry M. Construction of conductive multilayer films of biogenic triangular gold nanoparticles and their application in chemical vapour sensing. J Nanotechnol 2006;17(9):2399-409.