# ASIAN JOURNAL OF PHARMACEUTICAL AND CLINICAL RESEARCH

NNOVARE ACADEMIC SCIENCES Knowledge to Innovation

Vol 9, Issue 5, 2016

Online - 2455-3891 Print - 0974-2441 Review Article

# NITRIC OXIDE MEDIATED NEURODEGENERATION IN PARKINSON'S DISEASE

# VAIBHAV WALIA\*, SANTLAL KANSOTIA

Department of Pharmaceutical Sciences, Division Pharmacology, Maharshi Dayanand University, Rohtak - 124 001, Haryana, India. Email: vaibhav.walia00@gmail.com

Received: 12 March 2016, Revised and Accepted: 21 March 2016

## ABSTRACT

Nitric oxide (NO) is an endogenous molecule which functions as a neurotransmitter, hormone, free radical, etc. NO has been found to regulate the release of neurotransmitters, synaptic transmission, cell death, etc. NO is involved in the pathogenesis of various neuropsychiatric and neurodegenerative disorders. NO plays a key role in cellular apoptosis and neuronal degeneration. Parkinson' disease (PD) is a neurodegenerative disorder characterized by motor dysfunction that can be seen in the patients suffering from PD. The motor dysfunction is due to the progressive degeneration of dopaminergic neurons in mid brain. Dopamine (DA) is highly reactive molecule and is prone to the oxidation very much. The oxidation of DA is accompanied by the production of the reactive oxygen species that activates microglia cells. Upon activation, microglia cells cause the upregulation of inducible NO synthase, the enzyme involved in the production of NO. NO thus plays a key role in the neurodegeneration process implicated in PD. Thus, the aim of the present manuscript is to describe the possible role of NO in PD.

Keywords: Dopamine, Neuromelanin, Nitric oxide, Parkinson.

© 2016 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4. 0/) DOI: http://dx.doi.org/10.22159/ajpcr.2016.v9i5.11667

## INTRODUCTION

Parkinson's disease (PD) is a neurodegenerative disorder characterized by the degeneration of dopaminergic neurons in substantia nigra (SN) region of the brain. This degeneration of dopaminergic neurons in nigro striatal region contributes to the motor dysfunction in the patients of PD [1]. Therefore, the patients suffering from PD exhibits the symptoms such as rigidity, tremor, and bradykinesia. [2]. Excessive oxidation of proteins, and lipids, mitochondrial defects, low glutathione (GSH) levels, excessive dopamine (DA) o-quinone synthesis, and excessive levels of 5-cysteinyldopamine in the cerebrospinal fluid has been implicated in the patients of PD [3,4]. The death of dopaminergic neurons in the brain results in the release of neuromelanin, a pigment composed of a complex polymer of 5, 6-dihydroxyindole, and possibly 5-cysteinyldopamine, on a glycoprotein matrix [5]. Normally, neuromelanin is neuroprotective because it is a potent antioxidant, mops up toxic catecholamine o-quinones, and chelates large amounts of toxic heavy metals [6]. However, in excess, it becomes neurotoxic, largely by the disruption of cells [7]. Neuromelanin is known to activate the microglia cells that contain inducible nitric oxide synthase (iNOS), which when expressed produces greater quantity of NO. Thus, NO plays a key role in the pathogenesis of PD [8].

NO is one of the simplest biologically active molecules of unique chemical nature which plays a very important role in neurotransmission, acts in many tissues to regulate a wide range of physiological, pathological, and cellular processes [9]. NO plays a key role in several process such as the regulation of cell death, neurotransmission, and immune defense [10]. NO functions as a hormone, reactive oxygen species (ROS), neurotransmitter, constitutive mediator, inducible mediator, cytoprotective, and cytotoxic molecule [11]. Abnormal regulation or control of NO synthesis by impaired expression of NOS is capable of affecting a number of important biological processes and has been implicated in a variety of diseases [12]. NO diffuse rapidly across cell membranes and exerts its biological effects through the reaction of NO with a number of targets such as cysteine residues, hem groups, and iron and zinc clusters [13]. NO also enhances catecholamine release and to inhibit reuptake, possibly by reversing the transporter [14]. The above facts suggested the role of NO in PD [15].

# NOS AND THEIR FUNCTIONS

NO is a highly reactive, short-lived molecule produced from a group of enzymes known as NOS [16]. NO is produced from enzymes NOS which exists in three isoforms, from the amino acid L-arginine in the presence of many cofactors NADPH using flavin adenine dinucleotide (FAD), flavin mononucleotide (FMN), heme, thiol and tetrahydrobiopterin, and oxygen [17]. There are three types of NOS - endothelial NOS (eNOS), iNOS and neuronal NOS (nNOS) [18]. In each NOS, there are three distinct domains: Reductase domain, calmodulin (CaM)-binding domain, and oxygenase domain [19]. (1) Reductase domain contains the FAD and FMN moieties, and it transfer electrons from NADPH to oxygenase domain [20]. (2) CaM binding domain detects the changes in intracellular calcium levels, and therefore, the binding of CaM is required for the activity of all NOS isoforms [21]. (3) Oxygenase domain catalyzes the conversion of arginine into citrulline and NO [22]. This domain contains the binding sites for tetrahydrobiopterin, hem (heme), and arginine. iNOS is found in microglia cells and the expression of this isoform has been known to produces the high concentrations of NO [23]. iNOS mediated production of NO is preceded by increased intracellular Ca2+ concentration because is a CaM -dependent isoform [24]. The specific actions of NO on the neurotransmission are attributed primarily by NO produced by nNOS after the activation of N-methyl-d-aspartate (NMDA) receptors [25]. nNOS is linked with NMDA receptors via postsynaptic density protein [26]. Thus, the NMDA receptors activation exposes the enzyme directly to the flux of Ca2+ entering the ion channel. NO produced by the nNOS around the NMDA receptor reflects the activity of glutamate mediated neurotransmission [27]. NO synthesized intracellularly diffuses out through the neuronal membrane to induce the depolarization of the neuronal membrane and evoked the neurotransmitter release [28,29]. e-NOS produces NO results in the activation of soluble guanylyl cyclase, followed by the accumulation of cyclic guanosine monophosphate (cGMP), in vascular smooth muscle cells, which further results in the relaxation and vasodilatation [30]. The production of cGMP leads to the activation of cGMP-dependent protein kinases possibly to increase the expression of anti-apoptotic proteins [31].

## NO AND CELLULAR APOPTOSIS

NO is a potential bioregulator of apoptosis [32]. NO may prevent or induce apoptosis and can also increase or decrease the Bcl-2 levels. NO may act as an antiapoptotic signal by suppression of mitochondrial cytochrome c release, ceramide generation, and caspase activation [33]. NO donors elevate Bcl-2 expression and prevent apoptotic cell death. NO can inhibit apoptosis in some cells, whereas it promotes apoptosis in others cells [34]. Antiapoptotic signaling has been generally correlated with low or physiological NO levels [35]. NO shortage - induced decrease in CREB activity that interferes with the cGMP-dependent protein kinase, the most important intracellular signaling pathway activated by NO [36]. CREB and its associated proteins act as survival factors for human melanoma cells [37]. High physiological concentrations of NO-induced apoptosis in cells by the formation of transition metal complexes [32]. Thus, the activation of iNOS facilitates cell death [38]. NO also results in the activation of neutral SMase and increase the ceramide generation, main cell death mechanisms [39]. The mechanism of death action for ceramide may also involve, at least in part, a p53-dependent suppression of Bcl-2 expression [40]. It has also been showed that p53 promotes permeabilization of the outer mitochondrial membrane by forming complexes with the protective Bcl-xL and Bcl-2 proteins [41]. Moreover, the tumor suppressor p53 may also transactivate the expression of pro-apoptotic genes such as bax and cyclin-dependent kinase inhibitor p21 [42].

#### NO AND NEURODEGENATION

NO plays a key role in the neurodegeneration. It has been suggested that the levels of the GSH reduces in the patients of various neurodegenerative disorders PD and AD [43]. GSH levels were found to be lower in SN of PD. GSH is low molecular weight cellular non-enzymatic antioxidant which is synthesized in two steps: The enzyme g-glutamyl-cysteine synthetase (g-GCS) that catalyzes the synthesis of g-glutamyl-cysteine from glutamate and cysteine; and GSH synthetase (GS), which catalyzes the formation of GSH from glycine and g-glutamylcysteine [44]. It is present as a reduced form and two oxidized species: GSH disulfide and GSH mixed disulfide with protein thiols [45]. GSH acts as a cofactor of GSH peroxidase and participates in the detoxification of lipid and organic peroxides [46]. GSH modulates several proteins, including receptors, molecules involved in signaling and nuclear transcription factors [47]. It has been reported that in neurodegenerative diseases, the activity of both g-GCS and GS progressively decreases through the downregulation of gene expression and GSH decrease could be a consequence of the formation of protein mixed disulfides [48]. NO reacts with oxygen, superoxide anion (0,-), and reducing agents to give products such as nitroxyl, oxides  $(NO_2, N_2O_4)$  and  $N_2O_3$ , peroxynitrite  $(ONOO^-)$ , and S-nitrosothiols (RSNO) that produces the toxic effects and nitrosative stress [49]. In anaerobic conditions, direct reaction between NO and biological thiols can occur by a very slow oxidation reaction that yields thiol disulfide and nitroxyl anion [50]. The presence of oxygen is essential for the formation of S-nitrosoglutathione (GSNO) and protein RSNO [51]. NO toxicity in neuronal cells, indicate the disruption of cellular buffering mediated by GSH and the reaction of GSH with NO makes the neuronal cells more susceptible to damage [52]. The formation of GSNO is plausible, because it has a longer lifetime than NO and protein RSNO and is considered as a pool of NO that can be released when or where it is required for signaling [53]. Upon the depletion of GSH and upon inhibition of GSH synthesis, endogenous NO is the primary factor affecting cell proliferation and viability through the NO-cGMP pathway and NO-mediated DNA damage and protein oxidation [54]. This suggests that GSH could be an essential buffer of NO under physiological conditions and its imbalance is linked with the harmful effects of NO [55]. GSH decrease causes protein nitration, S-nitrosylation, and DNA strand breaks [56]. Such alterations results in the inhibition of cytochrome-c oxidase activity and microtubule network disassembly, which are considered hallmarks of NO toxicity [57]. Also it has been suggested that the treatment with physiological amounts of NO donors, an increase in GSH levels has been also reported [58]. Therefore, NO exerts neurodegenerative and neuroprotective effects in the concentration dependent manner.

## NO AND PD

PD is characterized by the death of dopaminergic neurons in the SN region of brain. The main reason behind the death of the dopaminergic neurons is not known clearly but it is suggested that the oxidative stress plays a key role in the pathogenesis of PD. Brain is susceptible to oxidative stress, due to its high oxygen demands, higher rate of oxidative metabolism, lower level of protective antioxidant system, higher membrane to surface area to cytoplasm volume, abundant neuronal network, etc., [59]. However, SN of midbrain is highly susceptible to oxidative stress because of its large population of dopaminergic neurons that produce abundant quantities of ROS species [60]. The higher level of oxidative stress in SN is marked by increased lipid peroxidation, decreased levels of GSH, increased iron concentrations, mitochondrial dysfunction, and DNA and protein oxidation [61].

The oxidation of DA initiates the neurodegeneration possibly due to the depletion of GSH and oxidation of ascorbate [62]. In SN, the increased metabolism of DA has been found to be associated with increased levels of ROS [63]. Increased production of ROS results in the disruption of mitochondrial enzymes participating in respiration and electron transport chain [64]. Oxidative stress in SN occurs due to the autoxidation of DA into semiquinones, which further generates ROS and ROS makes the dopaminergic neurons vulnerable to death [65]. The loss of DAergic neurons is due to the failure of glucose metabolism due to aberration in mitochondrial respiration which is further responsible for the failure of DA neurotransmission [66]. Oxidation of DA results in the DA quinone, which directly modifies the proteins and causes the irreversible inhibition of complex I activity, lead to impaired energy metabolism and cell death [67]. Mitochondrial-related energy failure disrupts the vesicular storage of DA and leads to increased autooxidation of DA neurotransmitter [68].

DA in SN is deaminated by the action of monoamine oxidase, which results in the production of dihydroxyphenylacetic acid and H<sub>2</sub>O<sub>2</sub> [66]. H<sub>2</sub>O<sub>2</sub> produced is converted into highly destructive hydroxyl radicals by the fenton reaction, with Fe2+ being liberated and these hydroxyl radicals then lead to widespread cellular damage [69]. Oxidation of DA produces DA-quinones, which cyclizes to form aminochrome, and aminochrome form adducts with proteins and is the precursor of neuromelanin that contribute to neurodegeneration by triggering neuroinflammation [70-73]. NM is natural protective and sequesters iron, free radicals, and toxic quinines. In PD, the loss of NM occurs gradually due to massive dopaminergic neuronal death [74,75]. NM can also binds with the heavy metals such as iron and the loss of these neurons in PD is correlated with an abundance of nonheme iron (Fe3+). Moreover, it has been suggested that the infusion of ferric iron into SN produces a dose-dependent reduction in dopaminergic activity. Iron released from neuromelanin increases oxidative stress in mitochondria and disrupts the mitochondrial function. The presence of iron together with a diminished supply of antioxidants leads to an increased generation of hydroxyl radicals through various reactions in the microglia, producing a cascade of destructive events including oxidative stress, lipid peroxidation, and eventually apoptosis. Moreover, the unbound iron initiate a range of cytotoxic and inflammatory effects, such as the activation of redox-sensitive transcriptional factor nuclear factor-kB and cytokine release from activated microglia [65]. The neuromelanin pathway can produce deleterious DA-quinone, which are potent inhibitors of mitochondrial complex I and promotes the  $\alpha$ -synuclein fibrillization [76-79].

Neuromelanin, released by dying DA neurons, has been reported to activate microglia cells [80]. Activated microglial cells contribute to an inflammatory reaction seen in PD [81]. Increased density of microglial cells expressing iNOS in SN of PD patients compared with control has been observed in the previous studies [82]. Activation of microglia is associated with an upregulation of iNOS resulting

in increased production of NO, suggesting that reactive nitrogen species plays a critical role in the disease [83]. Activated glial cells expresses the higher level of cytokines such as tumor necrosis factor- $\alpha$ , interleukin--1 $\beta$  and interferon- $\gamma$  as well as iNOS, which have been reported in SN in PD [84]. These inflammatory cytokines, along with factors released from the dying dopaminergic cells, seem to amplify, and sustain the neuroinflammation as well as further consequent immune responses leading to a potentially lethal descent into irreversible destruction of dopaminergic neurons in SN [85]. The cytotoxic mechanism thus involves the upregulation of iNOS. Increased expression of iNOS is known to mediate the increased production of NO, which has been shown to cause neuronal toxicity [86]. Therefore, the toxicity mediated by the increased NO levels may contribute to the death of the dopaminergic neurons. It has been suggested that the NO interact with oxygen-free radicals such as superoxide (O2-) to produce ONOO- and which further contribute to cellular injury, including lipid peroxidation, nitrosylation of some molecules, inactivation of sodium channels, and interactions with metals, which have redox potential such as iron and copper [87]. NO also increase the glutamate release, leading to inappropriately high NMDA receptor activity [88]. ONOO oxidize DA, deplete available reduced GSH/ascorbate, incur a substantial loss of endogenous GSH-peroxidase, and destroy the natural ability of GSH to act as an antioxidant [76,89]. ONOO- is potent oxidizing agent than NO, induces DNA fragmentation and lipid peroxidation. ONOO- also induces a dose-dependent impairment in DA synthesis independent of DA oxidation or cell death [13,90]. Exposure to ONOO- disrupts the catalytic activity of tyrosine hydroxylase, the rate-limiting enzyme involved in the DA synthesis [91,92]. ONOO- produced by high NMDA receptor activity depletes the adenosine triphosphate (ATP) pools by two major mechanisms [93]. It attacks mitochondrial proteins and lead to lowered production of ATP generation. It also nicks DNA, followed by the activation of poly (ADP-ribose) polymerase, leading to the depletion of NAD pool and finally ATP depletion, because NAD/NADH have essential roles in oxidative ATP generation [94]. It is well known that when ATP levels get lowered, NMDA receptors become hypersensitive due to lowering of the plasma membrane potential [95]. Thus, the stimulation of NMDA receptors results in the ATP depletion [96]. Excessive activation of NMDA receptors, results in excessive Ca2+ influx through a receptor's associated ion channel [97]. Increased Ca2+ levels in conjunction with the CaM, trigger the activation of nNOS, and subsequent generation of NO, a damaging free radicals, contributing to cell injury and death [98]. NO inhibits several enzymes including complexes I and IV of the mitochondrial electron transport chain, leading to ROS generation [99]. Therefore, overstimulation of NMDA receptors mediates neuronal damage. The intense hyperstimulation of NMDA receptors leads to necrotic cell death, but milder or chronic overstimulation results in apoptotic or other forms of cell death implicated in many neurological disorders [100]. Therefore, either the  $NO\ or\ NOS\ can\ be\ the\ targets\ for\ the\ treatment\ of\ PD\ and\ the\ modulators$ of NO can be the effective treatment of PD.

# REFERENCES

- Machado A, Herrera AJ, Venero JL, Santiago M, de Pablos RM, Villarán RF, et al. Inflammatory animal model for Parkinson's disease: The intranigral injection of LPS induced the inflammatory process along with the selective degeneration of nigrostriatal dopaminergic neurons. ISRN Neurol 2011;2011:476158.
- Tufekci KU, Genc S, Genc K. The endotoxin-induced neuroinflammation model of Parkinson's disease. Parkinsons Dis 2011;2011:487450.
- Olivieri S, Conti A, Iannaccone S, Cannistraci CV, Campanella A, Barbariga M, et al. Ceruloplasmin oxidation, a feature of Parkinson's disease CSF, inhibits ferroxidase activity and promotes cellular iron retention. J Neurosci 2011;31(50):18568-77.
- Martin HL, Teismann P. Glutathione A review on its role and significance in Parkinson's disease. FASEB J 2009;23(10):3263-72.
- Gaeta A, Hider RC. The crucial role of metal ions in neurodegeneration: The basis for a promising therapeutic strategy. Br J Pharmacol 2005;146(8):1041-59.
- Zecca L, Tampellini D, Gerlach M, Riederer P, Fariello RG, Sulzer D. Substantia nigra neuromelanin: Structure, synthesis, and molecular

- behaviour. Mol Pathol 2001;54(6):414-8.
- Offen D, Ziv I, Panet H, Wasserman L, Stein R, Melamed E, et al. Dopamine-induced apoptosis is inhibited in PC12 cells expressing Bcl-2. Cell Mol Neurobiol 1997:17:289-304.
- 8. Kavyaa R, Dikshitb M. Role of Nitric oxide/nitric oxide synthase in Parkinson's disease. Ann Neurosci 2005;12(2):???.
- Akyol O, Zoroglu SS, Armutcu F, Sahin S, Gurel A. Nitric oxide as a physiopathological factor in neuropsychiatric disorders. *In Vivo* 2004;18(3):377-90.
- Dröge W. Free radicals in the physiological control of cell function. Physiol Rev 2002;82(1):47-95.
- Gautam P, Jain SK. Functions and significance of nitric oxide in pathophysiological processes. Indian J Biotech 2007;6(3):293-304.
- Valko M, Leibfritz D, Moncola J, Cronin MT, Mazura M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007;39(1):44-84.
- Szabó C, Ischiropoulos H, Radi R. Peroxynitrite: Biochemistry, pathophysiology and development of therapeutics. Nat Rev Drug Discov 2007;6(8):662-80.
- Li X, Rose G, Chiari A, Pan HL, Tobin JR. Eisenach JC 6-NO(2)norepinephrine increases norepinephrine release and inhibits norepinephrine uptake in rat spinal synaptosomes. J Pharmacol Exp Ther 2000:292(3):895-9.
- 15. Pacher P, Beckman JS, Liaudet L. Nitric oxide and peroxynitrite in health and disease 2007;87(1):315-424.
- Blantz RC, Munger K. Role of nitric oxide in inflammatory conditions. Nephron 2002;90(4):373-8.
- Mayer B, Hemmens B. Biosynthesis and action of nitric oxide in mammalian cells. Trends Biochem Sci 1997;22(12):477-81.
- Ito Y, Ohkubo T, Asano Y, Hattori K, Shimazu T, Yamazato M, et al. Nitric oxide production during cerebral ischemia and reperfusion in eNOS - And nNOS-knockout mice. Curr Neurovasc Res 2010;7(1):23-31.
- Venema RC, Ju H, Zou R, Ryan JW, Venema VJ. Subunit interactions of endothelial nitric-oxide synthase. Comparisons to the neuronal and inducible nitric-oxide synthase isoforms. J Biol Chem 1997;272(2):1276-82.
- Alderton WK, Cooper CE, Knowles RG. Nitric oxide synthases: Structure, function and inhibition. Biochem J 2001;357:593-615.
- Balligand JL, Feron O, Dessy C. eNOS activation by physical forces: From short-term regulation of contraction to chronic remodeling of cardiovascular tissues. Physiol Rev 2009;89(2):481-534.
- Yoneyama H, Yamamoto A, Kosaka H. Neuronal nitric oxide synthase generates superoxide from the oxygenase domain. Biochem J 2001;360:247-53.
- 23. Wink DA, Hines HB, Cheng RY, Switzer CH, Flores-Santana W, Vitek MP, *et al.* Nitric oxide and redox mechanisms in the immune response. J Leukoc Biol 2011;89(6):873-91.
- Calabrese V, Boyd-Kimball D, Scapagnini G, Butterfield DA. Nitric oxide and cellular stress response in brain aging and neurodegenerative disorders: The role of vitagenes. *In vivo* 2004;18(3):245-67.
- Liu CY, Xie DP, Liu JZ. Microinjection of glutamate into dorsal motor nucleus of the Vagus excites gallbladder motility through NMDA receptor - Nitric oxide - cGMP pathway. Neurogastroenterol Motil 2004;16(3):347-53.
- Brenman JE, Bredt DS. Synaptic signaling by nitric oxide. Curr Opin Neurobiol 1997;7(3):374-8.
- Garthwaite J, Charles SL, Chess-Williams R. Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. Nature 1988;336(6197):385-8.
- Bronk P, Deák F, Wilson MC, Liu X, Südhof TC, Kavalali ET. Differential effects of SNAP-25 deletion on Ca2 - Dependent and Ca2 - Independent neurotransmission. J Neurophysiol 2007;98(2):794-806.
- Ohkuma S, Katsura M. Nitric oxide and peroxynitrite as factors to stimulate neurotransmitter release in the CNS. Prog Neurobiol 2001;64(1):97-108.
- Friebe A, Koesling D. Regulation of nitric oxide-sensitive guanylyl cyclase. Circ Res 2003;93(2):96-105.
- Ha KS, Kim KM, Kwon YG, Bai SK, Nam WD, Yoo YM, et al. Nitric oxide prevents 6-hydroxydopamine-induced apoptosis in PC12 cells through cGMP-dependent PI3 kinase/Akt activation. FASEB J 2003;17(9):1036-47.
- 32. Choi BM, Pae HO, Jang SI, Kim YM, Chung HT. Nitric oxide as a pro-apoptotic as well as anti-apoptotic modulator. J Biochem Mol Biol 2002;35(1):116-26.
- 33. Chung HT, Pae HO, Choi BM, Billiar TR, Kim YM. Nitric oxide as a bioregulator of apoptosis. Biochem Biophys Res Commun

- 2001;282(5):1075-9.
- von Knethen A, Callsen D, Brüne B. NF-kappaB and AP-1 activation by nitric oxide attenuated apoptotic cell death in RAW 264.7 macrophages. Mol Biol Cell 1999;10(2):361-72.
- 35. Kim YM, Bombeck CA, Billiar TR. Nitric oxide as a bifunctional regulator of apoptosis. Circ Res 1999;84(3):253-6.
- Gudi T, Casteel DE, Vinson C, Boss GR, Pilz RB. NO activation of fos promoter elements requires nuclear translocation of G-kinase I and CREB phosphorylation but is independent of MAP kinase activation. Oncogene 2000;19(54):6324-33.
- Jean D, Harbison M, McConkey DJ, Ronai Z, Bar-Eli M. CREB and its associated proteins act as survival factors for human melanoma cells. J Biol Chem 1998;273(38):24884-90.
- 38. Ferrer P, Asensi M, Priego S, Benlloch M, Mena S, Ortega A, *et al.* Nitric oxide mediates natural polyphenol-induced Bcl-2 downregulation and activation of cell death in metastatic B16 melanoma. J Biol Chem 2007;282(5):2880-90.
- Takeda Y, Tashima M, Takahashi A, Uchiyama T, Okazaki T. Ceramide generation in nitric oxide-induced apoptosis. Activation of magnesiumdependent neutral sphingomyelinase via caspase-3. J Biol Chem 1999;274(15):10654-60.
- Rudin CM, Thompson CB. Apoptosis and disease: Regulation and clinical relevance of programmed cell death. Annu Rev Med 1997;48:267-81.
- Heyne K, Schmitt K, Mueller D, Armbruester V, Mestres P, Roemer K. Resistance of mitochondrial p53 to dominant inhibition. Mol Cancer 2008;7:54.
- Chen X, Ko LJ, Jayaraman L, Prives C. p53 levels, functional domains, and DNA damage determine the extent of the apoptotic response of tumor cells. Genes Dev 1996;10(19):2438-51.
- Rappold PM, Tieu K. Astrocytes and therapeutics for Parkinson's disease. Neurotherapeutics 2010;7(4):413-23.
- Reed MC, Thomas RL, Pavisic J, James J, Ulrich CM, Nijhout HF. A mathematical model of glutathione metabolism. Theor Biol Med Model 2008;5:8.
- Chakravarthi S, Jessop CE, Bulleid NJ. The role of glutathione in disulphide bond formation and endoplasmic-reticulum-generated oxidative stress. EMBO Rep 2006;7(3):271-5.
- Zhu Y, Carvey PM, Ling Z. Altered glutathione homeostasis in animals prenatally exposed to lipopolysaccharide. Neurochem Int 2007;50(4):671-80.
- Sen CK. Redox signaling and the emerging therapeutic potential of thiol antioxidants. Biochem Pharmacol 1998;55(11):1747-58.
- 48. Franco R, Cidlowski JA. Apoptosis and glutathione: Beyond an antioxidant. Cell Death Differ 2009;16(10):1303-14.
- Vanuffelen BE, Van Der Zee J, De Koster BM, Vansteveninck J, Elferink JG. Intracellular but not extracellular conversion of nitroxyl anion into nitric oxide leads to stimulation of human neutrophil migration. Biochem J 1998;330:719-22.
- Zeng H, Spencer NY, Hogg N. Metabolism of S-nitrosoglutathione by endothelial cells. Am J Physiol Heart Circ Physiol 2001;281(1):H432-9.
- Aquilano K, Baldelli S, Cardaci S, Rotilio G, Ciriolo MR. Nitric oxide is the primary mediator of cytotoxicity induced by GSH depletion in neuronal cells. J Cell Sci 2011;124:1043-54.
- Ciriolo MR, De Martino A, Lafavia E, Rossi L, Carrì MT, Rotilio G. Cu, Zn-superoxide dismutase-dependent apoptosis induced by nitric oxide in neuronal cells. J Biol Chem 2000;275(7):5065-72.
- Thomas DD, Ridnour LA, Isenberg JS, Flores-Santana W, Switzer CH, Donzellie S, et al. The chemical biology of nitric oxide. Implications in cellular signaling. Free Radic Biol Med 2008;45(1):18-31.
- 54. Lirk P, Hoffmann G, Rieder J. Inducible nitric oxide synthase Time for reappraisal. Curr Drug Targets Inflamm Allergy 2002;1(1):89-108.
- Aquilano K, Vigilanza P, Rotilio G, Ciriolo MR. Mitochondrial damage due to SOD1 deficiency in SH-SY5Y neuroblastoma cells: A rationale for the redundancy of SOD1. FASEB J 2006;20(10):1683-5.
- Mikkelsen RB, Wardman P. Biological chemistry of reactive oxygen and nitrogen and radiation-induced signal transduction mechanisms. Oncogene 2003;22(37):5734-54.
- Abe K, Aoki M, Kawagoe J, Yoshida T, Hattori A, Kogure K, et al. Ischemic delayed neuronal death. A mitochondrial hypothesis. Stroke 1995;26(8):1478-89.
- Buckley BJ, Whorton AR. Adaptive responses to peroxynitrite: Increased glutathione levels and cystine uptake in vascular cells. Am J Physiol Cell Physiol 2000;279(4):C1168-76.
- 59. Kroemer G, Galluzzi L, Brenner C. Mitochondrial membrane permeabilization in cell death. Physiol Rev 2007;87(1):99-163.
- 60. Vernier P, Moret F, Callier S, Snapyan M, Wersinger C, Sidhu A. The

- degeneration of dopamine neurons in Parkinson's disease: Insights from embryology and evolution of the mesostriatocortical system. Ann N Y Acad Sci 2004;1035(1):231-49.
- Facecchia K, Fochesato LA, Ray SD, Stohs SJ, Pandey S. Oxidative toxicity in neurodegenerative diseases: Role of mitochondrial dysfunction and therapeutic Strategies. J Toxicol 2011;2011:683728.
- 62. Serra PA, Sciola L, Delogu MR, Spano A, Monaco G, Miele E, et al. The neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine induces apoptosis in mouse nigrostriatal glia. Relevance to nigral neuronal death and striatal neurochemical changes. J Biol Chem 2002;277(37):34451-61.
- 63. Escames G, López A, García JA, García L, Acuña-Castroviejo D, García JJ, et al. The role of mitochondria in brain aging and the effects of melatonin. Curr Neuropharmacol 2010;8(3):182-93.
- Kirkinezos IG, Moraes CT. Reactive oxygen species and mitochondrial diseases. Semin Cell Dev Biol 2001;12:449-57.
- Mounsey RB, Teismann P. Chelators in the treatment of iron accumulation in Parkinson's disease. Int J Cell Biol 2012;2012;1-12.
- Mazzio EA, Close F, Soliman KF. The biochemical and cellular basis for nutraceutical strategies to attenuate neurodegeneration in Parkinson's disease. Int J Mol Sci 2011;12(1):506-69.
- 67. Suematsu N, Tsutsui H, Wen J, Kang D, Ikeuchi M, Ide T, et al. Oxidative stress mediates tumor necrosis factor-alpha-induced mitochondrial DNA damage and dysfunction in cardiac myocytes. Circulation 2003;107:1418-23.
- Nicholls DG, Budd SL. Mitochondria and neuronal survival. Physiol Rev 2000;80:315-60.
- 69. Whitton PS. Inflammation as a causative factor in the aetiology of Parkinson's disease. Br J Pharmacol 2007;150(8):963-76.
- Ischiropoulos H, Beckman JS. Oxidative stress and nitration in neurodegeneration: Cause, effect, or association? J Clin Invest 2003;111(2):163-9.
- Hastings TG. The role of dopamine oxidation in mitochondrial dysfunction: Implications for Parkinson's disease. J Bioenerg Biomembr 2009;41(6):469-72.
- Norris EH, Giasson BI, Hodara R, Xu S, Trojanowski JQ, Ischiropoulos H, et al. Reversible inhibition of alpha-synuclein fibrillization by dopaminochrome-mediated conformational alterations. J Biol Chem 2005;280(22):21212-9.
- Zecca L, Wilms H, Geick S, Claasen JH, Brandenburg LO, Holzknecht C, et al. Human neuromelanin induces neuroinflammation and neurodegeneration in the rat substantia nigra: Implications for Parkinson's disease. Acta Neuropathol 2008;116(1):47-55.
- García-Molina F, Fenoll LG, Morote JC, García-Ruiz PA, Rodríguez-López JN, García-Cánovas F, et al. Opposite effects of peroxidase in the initial stages of tyrosinase-catalysed melanin biosynthesis. Int J Biochem Cell Biol 2005;37(6):1179-96.
- 75. Double KL, Ben-Shachar D, Youdim MB, Zecca L, Riederer P, Gerlach M. Influence of neuromelanin on oxidative pathways within the human substantia nigra. Neurotoxicol Teratol 2002;24(5):621-8.
- Antunes F, Nunes C, Laranjinha J, Cadenas E. Redox interactions of nitric oxide with dopamine and its derivatives. Toxicology 2005;208(2):207-12.
- 77. Li HT, Lin DH, Luo XY, Zhang F, Ji LN, Du HN, *et al.* Inhibition of alpha-synuclein fibrillization by dopamine analogs via reaction with the amino groups of alpha-synuclein. Implication for dopaminergic neurodegeneration. FEBS J 2005;272(14):3661-72.
- Akagawa M, Ishii Y, Ishii T, Shibata T, Yotsu-Yamashita M, Suyama K, et al. Metal-catalyzed oxidation of protein-bound dopamine. Biochemistry 2006;45(50):15120-8.
- 79. Smythies J, Galzigna L. The oxidative metabolism of catecholamines in the brain: A review. Biochim Biophys Acta 1998;1380(2):159-62.
- Block ML, Hong JS. Microglia and inflammation-mediated neurodegeneration: Multiple triggers with a common mechanism. Prog Neurobiol 2005;76(2):77-98.
- Farooqui T, Farooqui AA. Lipid-mediated oxidative stress and inflammation in the pathogenesis of Parkinson's disease. Parkinsons Dis 2011:2011:247467.
- 82. Choi DY, Liu M, Hunter RL, Cass WA, Pandya JD, Sullivan PG, et al. Striatal neuroinflammation promotes Parkinsonism in rats. PLoS One 2009;4(5):e5482.
- Zhou M, Wang CM, Yang WL, Wang P. Microglial CD14 activated by iNOS contributes to neuroinflammation in cerebral ischemia. Brain Res 2013;1506:105-14.
- 84. Warner DS, Sheng H, Batinic-Haberle I. Oxidants, antioxidants and the ischemic brain. J Exp Biol 2004;207:3221-31.
- 85. del Zoppo G, Ginis I, Hallenbeck JM, Iadecola C, Wang X,

- Feuerstein GZ. Inflammation and stroke: Putative role for cytokines, adhesion molecules and iNOS in brain response to ischemia. Brain Pathol 2000;10(1):95-112.
- Fattah DA, Khalifa N. Inflammatory biomarkers in patients with Parkinson's disease: Correlations with clinical severity and levodopa therapy. Egypt J Neurol Psychiat Neurosurg 2011;48(2):111-6.
- 87. Medeiros R, Prediger RD, Passos GF, Pandolfo P, Duarte FS, Franco JL, et al. Connecting TNF-alpha signaling pathways to iNOS expression in a mouse model of Alzheimer's disease: Relevance for the behavioral and synaptic deficits induced by amyloid beta protein. J Neurosci 2007;27(20):5394-404.
- 88. Li DP, Chen SR. Nitric oxide stimulates glutamatergic synaptic inputs to baroreceptor neurons through potentiation of Cav2.2-mediated Ca(2) currents. Neurosci Lett 2014;567:57-62.
- 89. Blum D, Torch S, Lambeng N, Nissou M, Benabid AL, Sadoul R, et al. Molecular pathways involved in the neurotoxicity of 6-OHDA, dopamine and MPTP: Contribution to the apoptotic theory in Parkinson's disease. Prog Neurobiol 2001;65(2):135-72.
- Carr AC, McCall MR, Frei B. Oxidation of LDL by myeloperoxidase and reactive nitrogen species: Reaction pathways and antioxidant protection. Arterioscler Thromb Vasc Biol 2000;20(7):1716-23.
- 91. Park S, Geddes TJ, Javitch JA, Kuhn DM. Dopamine prevents nitration of tyrosine hydroxylase by peroxynitrite and nitrogen dioxide: Is nitrotyrosine formation an early step in dopamine neuronal damage? J Biol Chem 2003;278(31):28736-42.
- 92. Ara J, Przedborski S, Naini AB, Jackson-Lewis V, Trifiletti RR, Horwitz J, et al. Inactivation of tyrosine hydroxylase by nitration

- following exposure to peroxynitrite and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). Proc Natl Acad Sci U S A 1998;95(13):7659-63.
- Gow AJ, Farkouh CR, Munson DA, Posencheg MA, Ischiropoulos H. Biological significance of nitric oxide-mediated protein modifications. Am J Physiol Lung Cell Mol Physiol 2004;287(2):L262-8.
- Pall ML. NMDA sensitization and stimulation by peroxynitrite, nitric oxide, and organic solvents as the mechanism of chemical sensitivity in multiple chemical sensitivity. FASEB J 2002;16(11):1407-17.
- Duncan AJ, Heales SJ. Nitric oxide and neurological disorders. Mol Aspects Med 2005;26(1-2):67-96.
- Leist M, Single B, Castoldi AF, Kühnle S, Nicotera P. Intracellular adenosine triphosphate (ATP) concentration: A switch in the decision between apoptosis and necrosis. J Exp Med 1997;185(8):1481-6.
- Lipton SA. Failures and successes of NMDA receptor antagonists: Molecular basis for the use of open-channel blockers like memantine in the treatment of acute and chronic neurologic insults. NeuroRx 2004;1(1):101-10.
- Nakamura T, Lipton SA. Preventing Ca2 Mediated nitrosative stress in neurodegenerative diseases: Possible pharmacological strategies. Cell Calcium 2010;47(2):190-7.
- Dias V, Junn E, Mouradian MM. The role of oxidative stress in Parkinson's disease. J Parkinsons Dis 2013;3(4):461-91.
- 100.Verche VL, Ikiz B, Jacquier A, Przedborski S, Re DB. Glutamate pathway implication in amyotrophic lateral sclerosis: What is the signal in the noise? J Recept Ligand Channel Res 2011;4:1-22.