

## EVALUATION OF ANTIBACTERIAL ACTIVITY OF ESSENTIAL OIL OF *CINNAMOMUM ZEYLANICUM*, *EUGENIA CARYOPHYLLATA*, AND *ROSMARINUS OFFICINALIS* AGAINST *STREPTOCOCCUS ORALIS*

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### ABSTRACT

**Objective:** *Streptococcus oralis* plays an important role in the biofilm formation of dental plaque and the occurrence of periodontal disease. The present study was conducted to evaluate in vitro antibacterial activity of three essential oils, namely, *Cinnamomum zeylanicum*, *Eugenia caryophyllata*, and *Rosmarinus officinalis* against *S. oralis*.

**Methods:** The antibacterial activity of essential oils was investigated by diffusion method using sterile discs (or aromagrams). The minimum inhibitory concentration (MIC) of essential oils showing important antibacterial activity was measured using the broth dilution method.

**Results:** Evaluation of the antibacterial activity of three essential oils as determined by the aromagram technique showed that the essential oil of *R. officinalis* had no effect on *S. oralis*, while the latter was extremely sensitive to the other two essential oils, but with a higher efficiency of the essential oil of *C. zeylanicum* (42 mm diameter) than *E. caryophyllata* (20 mm diameter). Similarly, the MIC and minimum bactericidal concentration (MBC) were higher for the essential oil of *C. zeylanicum* than the essential oil of *E. caryophyllata*. The MBC/MIC ratio is of the order of 2. The essential oils studied therefore appear to exert bactericidal activity against *S. oralis*.

**Conclusion:** The findings suggest that essential oils of *C. zeylanicum* and *E. caryophyllata* may be used as an alternative to synthetic antibiotics.

**Keywords:** Essential oil, *Cinnamomum zeylanicum*, *Eugenia caryophyllata*, *Rosmarinus officinalis*, Antimicrobial activity, *Streptococcus oralis*.

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### INTRODUCTION

*Streptococcus oralis* is a Gram-positive coccus-shaped bacterium, considered a commensal bacterium which belongs to the mitis group. It is one of the first bacteria which begin to form the biofilm of dental plaque [1]. The research shows that this bacterium can interact with *Porphyromonas gingivalis*, which is considered one of the main causes of periodontal disease, the most common disease affecting the human oral cavity. In addition, it is an opportunistic bacterium that affects immunodeficient individuals and those with hematologic malignancies. In these individuals, it can create complications such as bacterial endocarditis, respiratory distress syndrome in adults, and streptococcal shock [2].

It is proven that advances in science and medicine have led to the development of many drugs of interest today. However, their use is not always rational and their long-term prescription has led to the so-called iatrogenic diseases, responsible for many adverse effects, even death. Thus, the misuse of antibiotics against various infections results in decreased efficiency due to increased resistance of bacteria [3-5]. This antibiotic resistance phenomenon is general and concerns all bacterial species including those of the oral cavity [6].

Furthermore, the other antibacterial agents used in the prevention and treatment of oral diseases, such as cetylpyridinium chloride, chlorhexidine, amine fluoride, or products containing such agents, are not devoid of toxicity [7], and side effects as in the case of ethanol (commonly used in mouthwashes) was observed in oral cancer [8].

Therefore, the search for alternative products continues, and natural phytochemicals isolated from plants used in traditional medicine are considered good alternatives to synthetic chemicals. Natural substances such as cinnamon bark oil and clove oil (cinnamaldehyde and eugenol) showed activity against oral bacteria [9].

In similar research of natural antimicrobial substances, we are interested in evaluating the antibacterial effect of the essential oils of *Cinnamomum zeylanicum*, *Eugenia caryophyllata*, and *Rosmarinus officinalis* against *S. oralis* which is a pathogen found in an unusual way and predominantly following a lack of hygiene in the oral cavity, in diabetic patients with periodontal disease.

### METHODS

#### Essential oils

The three essential oils tested in the present study were provided by the Subnârôme Laboratory, Department of Food Science and Nutrition at the Institute of Agronomy and Veterinary Hassan II in Rabat, Morocco. They were extracted by hydrodistillation and stored at 4°C before use. The chemical composition of these essential oils was analyzed by a gas chromatograph. The percentage composition of these oils is shown in Table 1.

#### Bacterial strains and culture conditions

The tested bacterial strain was a Gram-positive bacterium: *S. oralis*, which was isolated from the oral cavity of diabetic patients with gingivitis. Bacterial strains were grown in blood agar medium and incubated at 37°C in a CO<sub>2</sub> incubator for 24 hrs. After incubation, the strains were identified by the API gallery.

The purity of the strain was verified by continuous cultures in blood agar medium.

#### Disc diffusion method

Antimicrobial activity was investigated by the disc diffusion method as already described [10]. The bacterial suspension was adjusted to a bacterial cell density of 1.0 × 10<sup>8</sup> CFU/mL (or 0.5 McFarland turbidity units). A sterile swab immersed in this bacterial suspension was used to inoculate the

entire surface of sheep blood agar; 5  $\mu$ L of each essential oil was applied on a sterilized disc made from Whatman filter paper of 6 mm diameter [11], aseptically placed on the inoculated plates. Then, plates were incubated for 15 minutes at room temperature. Only one disc was tested per plate. After 24 hrs of incubation at 37°C in a CO<sub>2</sub> incubator, the inhibition zones were measured in millimeters. All experiments were done in triplicate. The average inhibition diameter was calculated to classify the essential oils as follows: *S. oralis* is not sensitive for a diameter <8 mm, moderately sensitive (+) for diameter of 8-14 mm, sensitive (++) for diameter of 14-20mm, and very sensitive (+++) for a diameter >20 mm [10,12].

#### Determination of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)

The broth macrodilution method was employed to determine the MIC [13]. Serial dilutions of essential oil ranging from 20 to 0.15  $\mu$ L/mL were prepared in test tubes containing Luria–Bertani broth with 0.15% agar. Each tube was inoculated with a bacterial suspension adjusted to 10<sup>6</sup> CFU/mL. Controls containing medium with either microorganisms or the essential oil alone were included. The tubes were then incubated at 37°C for 24 hrs. MIC values were defined as the lowest concentrations of essential oil at which the absence of growth was recorded. To determine the MBC, 10  $\mu$ L from tubes in which bacterial growth was not observed was spread on Mueller–Hinton agar and incubated at 37°C for 24 hrs. The MBC was defined as the lowest concentration of essential oil at which the incubated microorganism was completely killed [14]. Each test was performed in triplicate.

## RESULTS AND DISCUSSION

### Essential oil composition

As shown in Table 1, essential oils were chosen according to their chemical composition, in particular to their major components. The major compound of *C. zeylanicum* was cinnamaldehyde. Analysis of *E. caryophyllata* indicated eugenol and *R. officinalis* mainly contained cineole.

### Antibacterial activity of essential oils

Results obtained with the disc diffusion assay regarding the growth of inhibition zones of the tested *S. oralis* strain are shown in Table 2.

Our results showed that essential oils from *C. zeylanicum* and *E. caryophyllata* were the most active of the oils tested against *S. oralis*, with average inhibition zones ranging from 42.0 to 22.0mm (+++), while the essential oil of *R. officinalis* did not show antibacterial activity for this bacterium.

### MIC and MBC value determination

Referring to the large inhibition zones observed with the disc diffusion method for two essential oils (*C. zeylanicum* and *E. caryophyllata*), the MIC values were determined with broth dilution assays (Table 3).

*C. zeylanicum* essential oil, mainly composed of aldehyde, was most efficient against *S. oralis* (0.625  $\mu$ L/mL). The MIC of *E. caryophyllata* containing mainly eugenol was 1.25  $\mu$ L/mL.

Concerning the MBC, in most cases, it was close to the MIC, indicating good bactericidal activity against *S. oralis*, with an MBC-to-MIC ratio of the order of 2 for both essential oils.

Plants have been used by humans since antiquity to handle common infectious diseases. Some of these traditional treatments are always included as part of the usual treatment of various diseases [15,16].

These plants are an important reservoir of potential compounds, which have the advantage of having a big diversity in chemical structure and possessing a very wide range of biological activity [17,18].

Three essential oils were selected for their composition. Indeed, in the literature, it has been reported that essential oils containing mainly aromatic phenols or aldehydes presented major antimicrobial activity against respiratory tract pathogens [19,20].

**Table 1: Chemical composition percentage of three essential oils**

Component	Essential oil (%)
<i>C. zeylanicum</i>	
Cinnamaldehyde	74.4
N-acetate	9.91
Eugenylacetate	2.78
Hydrocinnamylacetate	1.14
<i>E. caryophyllata</i>	
Eugenol	79.71
$\beta$ -caryophyllene	15.85
Eugenylacetate	2.70
<i>R. officinalis</i>	
$\alpha$ -pinene	20.62
Camphene	7.00
$\beta$ -pinene	8.89
1,8 cineol	52.77
Camphor	6.64

*C. zeylanicum*: *Cinnamomum zeylanicum*, *E. caryophyllata*: *Eugenia caryophyllata*, *R. officinalis*: *Rosmarinus officinalis*

**Table 2: Inhibition zone diameters obtained with the three essential oils against *Streptococcus oralis***

Essential oil	Diameter in mm	Sensitivity
<i>C. zeylanicum</i>	42 $\pm$ 0.5	VS
<i>E. caryophyllata</i>	22 $\pm$ 0.66	S
<i>R. officinalis</i>	00	R

R: Resistant, S: Sensitive, VS: Very sensitive, *C. zeylanicum*: *Cinnamomum zeylanicum*, *E. caryophyllata*: *Eugenia caryophyllata*, *R. officinalis*: *Rosmarinus officinalis*

**Table 3: MIC and MBC of two selected essential oils against *Streptococcus oralis***

Essential oil	MIC ( $\mu$ L/mL)	MBC ( $\mu$ L/mL)	MBC/MIC
<i>C. zeylanicum</i>	0.625	1.25	2
<i>E. caryophyllata</i>	1.25	2.5	2

MIC: Minimum inhibitory concentration, MBC: Minimum bactericidal concentration, *C. zeylanicum*: *Cinnamomum zeylanicum*, *E. caryophyllata*: *Eugenia caryophyllata*

Chromatographic analysis of the essential oils showed that the major compounds of *C. zeylanicum*, *E. caryophyllata*, and *R. officinalis* were, respectively, cinnamaldehyde (74.4%), eugenol (79.71%), and 1,8 cineole (52.77%); these components could be the active elements of these essential oils. These results are similar to those reported by other authors: Burdock in 1995 and Raynaud in 2006 [21,22].

The study of the antibacterial effect of essential oils by the standardized disc assay method showed that *R. officinalis* had no effect on *S. oralis*, while this bacterium was extremely sensitive to the essential oil of *E. caryophyllata* and that of *C. zeylanicum*.

In addition, we showed that cinnamon presented higher activity against *S. oralis*. This result is consistent with other studies that reported that the essential oil of *C. zeylanicum* containing cinnamaldehyde (an aromatic aldehyde) showed higher activity than that of *E. caryophyllata* [20,23,24].

Moreover, the essential oil of *E. caryophyllata* containing an aromatic phenol (clove containing eugenol) was less active (++) than *C. zeylanicum*. These results could be directly linked to the structures of the major aromatic phenols from clove essential oil. In fact, essential oils containing the aromatic phenols, carvacrol and thymol, were more efficient (+++) against *Streptococci* [23]. These phenolic compounds are deemed to have great antibacterial activity [25-29].

The differences between our results and previous studies could be due to the fact that the composition of essential oils is not strictly defined

but is a complex mixture of organic substances, varying in quality and quantity [30-32].

Indeed, previous studies showed that the essential oil of *C. zeylanicum* was more effective than that of *E. caryophyllata* on the oral microbiota [33].

The antibacterial activity of the oils selected was studied by determining the MIC and MBC. In this study, MIC results were reliable with the inhibition zone diameters observed with the disc diffusion method, *C. zeylanicum* being the more effective essential oil followed by the essential oil of *E. caryophyllata*. The MBC/MIC ratio is of the order of 2. According to Guinoiseau [34], both essential oils studied appear to exert a bactericidal effect against *S. oralis* [35], but investigations such as pharmacokinetic and pharmacodynamic studies are needed to characterize the antibacterial activity *in vivo* and their clinical efficacy [36].

## CONCLUSION

We show interesting antibacterial activity of two essential oils against *S. oralis* isolated from the oral cavity of diabetic patients with gingivitis, particularly *C. zeylanicum* essential oil, but we need further investigations to evaluate the bactericidal properties in practical applications on clinical strains and to assess the potential for therapeutic application. As there is no evidence for the potential clinical use of these essential oils, further research is needed to determine whether they could efficiently substitute antibiotics or, perhaps, be used in combination.

Indeed, this preliminary result may be a basis for launching other studies on the action of these active ingredients on this pathogenic strain *in vivo*.

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## REFERENCES

1. Feur E, Labeyrie C, Boucher J, Eid A, Cabut S, Dib S, et al. Health indicators among high school teenagers, Val-de-Marne, France in 2005: Overweight, dental caries and risk of depression. *Wkly Epidemiol Bull* 2007;4:29-31.
2. Maeda K, Nagata H, Kuboniwa M, Ojima M, Tsukaza O, Naoto M, et al. Identification and characterization of *Porphyromonas gingivalis* customer proteins bind to *Streptococcus oralis* glyceraldehyde-3-dehydrogenase. *Infect Immun* 2013;81(3):753-63.
3. Singer RS, Finch R, Wegener HC, Bywater R, Walters J, Lipsitch M. Antibiotic resistance the interplay between antibiotic use in animals and human beings. *Lancet Infect Dis* 2003;3(1):47-51.
4. Bisht R, Katiyar A, Singh R, Mittal P. Antibiotic resistance – A global issue of concern. *Asian J Pharm Clin Res* 2009;2:34-9.
5. Sharma P, Mack JP, Rojzman A. Ten highly effective essential oils inhibit growth of methicillin resistant *Staphylococcus aureus* and methicillin sensitive *Staphylococcus aureus*. *Int J Pharm Pharm Sci* 2013;5(1):52-4.
6. Bidanlt P, Chandad F, Grenier D. Risks of bacterial resistance associated to systemic antibiotherapy in periodontics. *J Can Dent Assoc* 2007;73(8):721-5.
7. Rodrigues F, Lehmann M, do Amaral VS, Reguly ML, de Andrade HH. Genotoxicity of three mouthwash products, cepacol, periogard, and plax, in the *Drosophila* wing-spot test. *Environ Mol Mutagen* 2007;48(8):644-9.
8. Lachenmeier DW. Safety evaluation of topical applications of ethanol on the skin and inside the oral cavity. *J Occup Med Toxicol* 2008;3:26.
9. Saeki Y, Ito Y, Shibata M, Sato Y, Okuda K, Takazoe I. Antimicrobial action of natural substances on oral bacteria. *Bull Tokyo Dent Coll* 1989;30(3):129-35.
10. El Amri J, El Badaoui K, Zair T, Bouharb H, Chakir S, Alaoui T. Study of the antibacterial activity of the essential oils of *Teucrium capitatum* L and the extract of *Silene vulgaris* on different strains tested. *J Appl Biosci* 2014;82:7481-92.
11. Dulger B, Ugurlu E. Evaluation of antimicrobial activity of some endemic *Scrophulariaceae* members from Turkey. *Pharm Biol* 2005;43(3):275-9.
12. Franchomme P, Jollois R, Penoël D. *L'Aromathérapie Exactement*. Limoges, France: Roger Jollois; 2001.
13. Bouhdid S. Antimicrobial and antioxidant activities of essential oils. Doctoral Thesis. Faculty of Science, University Abdelmalek Essaadi, Tetouan, Morocco; 2009.
14. Smith-Palmer A, Stewart J, Fyfe L. Antimicrobial properties of plant essential oils and essences against five important food-borne pathogens. *Lett Appl Microbiol* 1998;26(2):118-22.
15. Rios JL, Recio MC. Medicinal plants and antimicrobial activity. *J Ethnopharmacol* 2005;100 1 Suppl 2:80-4.
16. Rollinger JM, Haupt S, Stuppner H, Langer T. Combining ethnopharmacology and virtual screening for lead structure discovery: COX-inhibitors as application example. *J Chem Inf Comput Sci* 2004;44(2):480-8.
17. Anbukumaran A, Ambikapathy V, Panneerselvam A. Preliminary phytochemical and antimicrobial activity of leaves of *Azima tetraacantha* Lam. *World J Pharm Life Sci* 2016;2(2):127-32.
18. Sbayou H, Talbi H, Talha I, Amghar S, Hilali A. Evaluation of the genotoxicity of essential oil from *Origanum compactum* benth. in human lymphocytes. *Asian J Pharm Clin Res* 2016;9(2):274-6.
19. Inouye S, Takizawa T, Yamaguchi H. Antibacterial activity of essential oils and their major constituents against respiratory tract pathogens by gaseous contact. *J Antimicrob Chemother* 2001;47(5):565-73.
20. Inouye S, Yamaguchi H, Takizawa T. Screening of the antibacterial effects of a variety of essential oils on respiratory tract pathogens, using a modified dilution assay method. *J Infect Chemother* 2001;7(4):251-4.
21. Burdock GA, editor. *Fenaroli's Handbook of Flavor Ingredients*. 3<sup>rd</sup> ed., Vol. I. Boca Raton, Florida: CRC Press; 1995. p. 134.
22. Raynaud J. *Prescription and Advice in Aromatherapy*. Paris: Lavoisier Technique et Documentation; 2006. p. 86-214.
23. Fabio A, Cermelli C, Fabio G, Nicoletti P, Quaglio P. Screening of the antibacterial effects of a variety of essential oils on microorganisms responsible for respiratory infections. *Phytother Res* 2007;21(4):374-7.
24. Bouhdid S, Abrini J, Baudoux D, Manresa A, Zhiri A. Essential oils of oregano compact and cinnamon: Antibacterial potency and mechanism of action. *J Pharm Clin* 2012;31(3):141-8.
25. Ultee A, Gorris LG, Smid EJ. Bactericidal activity of carvacrol towards the food-borne pathogen *Bacillus cereus*. *J Appl Microbiol* 1998;85(2):211-8.
26. Ultee A, Kets EP, Smid EJ. Mechanisms of action of carvacrol on the food-borne pathogen *Bacillus cereus*. *Appl Environ Microbiol* 1999;65(10):4606-10.
27. Ettayebi K, El Yamani J, Rossi-Hassani BD. Synergistic effects of nisin and thymol on antimicrobial activities in *Listeria monocytogenes* and *Bacillus subtilis*. *FEMS Microbiol Lett* 1999;183:191-5.
28. Ultee A, Slump RA, Steging G, Smid EJ. Antimicrobial activity of carvacrol toward *Bacillus cereus* on rice. *J Food Prot* 2000;63(5):620-4.
29. Sbayou H, Boumazza A, Hilali A, Amghar S. Chemical composition and antibacterial and antioxidant activities of *Thymus satureioides* coss. essential oil. *Int J Pharm Pharm Sci* 2016;8(10):183-7.
30. Bruneton J. *Pharmacognosie Phytochimie, Plantes Médicinales*. 4<sup>th</sup> ed. Paris: Lavoisier Technique et Documentation; 2009. p. 483.
31. Dorman HJ, Deans SG. Antimicrobial agents from plants: Antibacterial activity of plant volatile oils. *J Appl Microbiol* 2000;88(2):308-16.
32. Delaquis PJ, Stanich K, Girard B. Antimicrobial activity of individual and mixed fractions of dill, cilantro, coriander and eucalyptus essential oils. *Int J Food Microbiol* 2002;74 1 Suppl 2:101-9.
33. Gupta C, Kumari A, Garg AP, Catanzaro R, Marotta F. Comparative study of cinnamon oil and clove oil on some oral microbiota. *Acta Biomed* 2011;82(3):197-9.
34. Guinoiseau E. *Antibacterial Molecules Derived from Essential Oils: Separation, Identification and Mode of Action*. France: Life Sciences, University of Corsica; 2010. p. 65-70.
35. Mayaud L, Carricajo A, Zhiri A, Aubert G. Comparison of bacteriostatic and bactericidal activity of 13 essential oils against strains with varying sensitivity to antibiotics. *Lett Appl Microbiol* 2008;47:167-73.
36. Pankey GA, Sabath LD. Clinical relevance of bacteriostatic versus bactericidal mechanisms of action in the treatment of gram-positive bacterial infections. *Clin Infect Dis* 2004;38(6):864-70.