

**FORMULATION AND EVALUATION OF GASTRORETENTIVE FLOATING BIOADHESIVE TABLETS OF HYDROCHLOROTHIAZIDE**

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**ABSTRACT**

**Objective:** Floating bioadhesive tablets of hydrochlorothiazide were developed to prolong gastric residence time leading to an increase in drug bioavailability where here a combination of floating and bioadhesion mechanism is combined.

**Methods:** Tablets are prepared by direct compression technique using polymers xanthan gum, carbopol 974 P, HPMC K15M, HPMC K100M, magnesium Stearate USP-NF (Avicel PH 102), microcrystalline cellulose Ph 102, Talc, and sodium bicarbonate.

**Results:** Tablets were evaluated for their physical characteristics, namely, hardness, thickness, friability and weight variation, drug content, and floating properties. The best formulation subjected for kinetic treatment, i.e., zero order, first order, peppas, Higuchi, and Hixon-crowel. The R values are 0.9366, 0.9364, 0.9680, 0.9974, and 0.9283, respectively.

**Conclusion:** Optimized formulae F4 containing polymers HPMC K4M and CARBOPOL 974 P showed more bioadhesion with a controlled release over 12 hrs. Therefore, formulation F4 identified as a successful formulation for the development of floating bioadhesive tablets.

**Keywords:** Hydrochlorothiazide, Floating tablets, Gastroretentive drug delivery system.

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**INTRODUCTION**

Historically, oral drug administration has been the predominant route for drug delivery. During the past two decades, numerous oral delivery systems have been developed to act as drug reservoirs from which the active substance can be released over a defined period at a predetermined and controlled rate. From a pharmacokinetic point of view, the ideal sustained and controlled release dosage form should be comparable with an intravenous infusion, which supplies continuously the amount of drug needed to maintain constant plasma levels once the steady state is reached [1].

Although some important applications, including oral administration of peptide and protein drugs, can be used to prepare colonic drug delivery systems, targeting drugs to the colon by the oral route. More often, the drug absorption is unsatisfactory and highly variable among and between individuals, despite excellent *in vitro* release patterns. The reasons for this are essentially physiological and usually affected by the GI transit of the form, especially its gastric residence time, which appears to be one of the major causes of the overall transit time variability [2].

Hydrochlorothiazide is a diuretic of the benzothiadiazine class, and it was a very good choice drug in the management of mild to moderate hypertension. It inhibits sodium reabsorption in distal tubules causing increased excretion of sodium and water as well as potassium and hydrogen ions. Hydrochlorothiazide (HCTZ) is a poor water soluble drug having plasma half-life of 6-8 hrs and oral bioavailability is 70%. Its absorption window lies in the stomach therefore as long as it retains in the gastric environment an enhanced bioavailability can be seen. The present study was undertaken with the objective to develop an optimized fluid bed dryer containing HCTZ as a model drug to improve absorption and its oral bioavailability. In the current study, the effect of polymer (HPMC K4M, K15M, and Carbopol 974 P), polymer concentration and viscosity on drug release behavior and the buoyancy properties of prepared formulations were evaluated.

**METHODS**

HCTZ obtained as a gift sample from hetero laboratories Hyderabad. HPMC K15M, HPMC K100M, and carbopol 974 P were obtained from Signet Chemical Corporation, Mumbai, Avicel pH 102, Talc, Conc. Hydrochloric acid, Aerosil, sodium Bicarbonate, magnesium stearate obtained from S.D. Fine Chemicals, Mumbai.

**Preparation of compression HCTZ floating tablets by direct [3-5]**

All the ingredients were accurately weighed as per formula and dispensed in clean polythene covers. HCTZ were sifted through sieve No. 60. HPMC K15M, K100M, carbopol 974 P, xanthan gum, magnesium stearate USP-NF (Avicel PH 102), microcrystalline cellulose Ph 102, Talc, and sodium bicarbonate passed through sieve No. 40. After sifting all the above ingredients were transferred into a big polythene cover and mixed for 10 minutes. Then tablets are compressed in compression machine at specified pressure with 9 mm "B" tooling round punch according to the formulae given in Table 1.

**Preparation of calibration curve for hydrochlorothiazide****Standard curve IN 0.1 N HCL**

Accurately weighed 10 mg of drug (hydrochlorothiazide) was first dissolved in 10 mL of methanol in 100 mL of volumetric flask to make a concentration of 1000 µg/mL (primary stock solution). 1 mL of primary stock solution was pipetted out into 10 mL of volumetric flask and volume was adjusted with water to make a concentration of 100 µg/mL (secondary stock solution). From the secondary stock solution, various concentrations such as 1, 2, 3, 4, 5 .... 10 µg/mL were prepared for the calibration curve. A standard curve was plotted by taking absorbance of secondary stock solutions in ultraviolet (UV) double beam spectrophotometer at 272 nm.

**Compatibility studies**

Compatibility with excipients was confirmed by IR studies. The pure drug and its formulations along with excipients were subjected to IR

Table 1: Composition of formulations

Ingredients	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10	F11	F12
HCTZ	25	25	25	25	25	25	25	25	25	25	25	25
HPMC K 15	90	120	150	45	60	75	45	60	75	45	60	75
Carbopol 974 p				45	60	75						
Xanthum gum							45	60	75			
HPMC K-100										45	60	75
NaHCO <sub>3</sub>	50	50	50	50	50	50	50	50	50	50	50	50
Magnesium stearate USP-NF (Avicel PH 102)	3	3	3	3	3	3	3	3	3	3	3	3
Talc	3	3	3	3	3	3	3	3	3	3	3	3
Microcrystalline cellulose Ph 102	Q.S											
Total	300	300	300	300	300	300	300	300	300	300	300	300

HCTZ: Hydrochlorothiazide

studies. In the present study, the potassium bromide disc (pellet) method was employed.

Evaluation of precompression blend [3,6,7]

The powder blend of all formulations was evaluated for bulk density, tapped density, compressibility index, Hausner ratio, and Angle of repose.

#### A. Bulk density

30 g of material was passed through a sieve No. 25 to break up agglomerates and introduced into a dry 100 mL cylinder, without compacting, the powder was carefully leveled without compacting and the unsettled apparent volume,  $V_0$ , was read. The bulk density was calculated, in g/mL, using the formula.

$$(M)/(V_0)$$

Where M = Total weight of the powder blend and  $V_0$  is the bulk volume of the powder blend.

#### B. Tapped density

After carrying out the procedure as given in the measurement of bulk density the cylinder containing the sample was tapped using a mechanically tapped density tester (Electrolab) that provides a fixed drop of  $14 \pm 2$  mm at a nominal rate of 300 drops/minute. The cylinder was tapped 500 times initially followed by an additional tap of 750 times until difference between succeeding measurement was  $< 2\%$  and then tapped volume  $V_f$  was measured to the nearest graduated unit. The tapped density was calculated, in g/mL, using the formula:

$$(M)/(V_f)$$

Where M = Total weight of the powder blend and  $V_f$  is the tapped volume of the powder blend.

#### C. Measures of powder compressibility

The compressibility index and Hausner ratio are measures of the propensity of powder to be compressed. As such, they are measures of the relative importance of inter-particulate interactions. As such, they are measures of the relative importance of inter-particulate interactions. In a free-flowing powder, such interactions are generally less significant, and the bulk and tapped densities will be closer in value. For poorer flowing materials, there are frequently greater interparticle interactions and a greater difference between the bulk and tapped densities will be observed. These differences are reflected in the compressibility index and the Hausner ratio, which are calculated using the following formulae [8-11].

$$\text{Compressibility index} = (V_f - V_0) * 100 / V_f$$

Where  $V_f$  = tapped density;  $V_0$  = Bulk density.

#### D. Hausner ratio

It is the ratio of bulk density to tapped density.

$$V_0/V_f$$

$V_0$  = Bulk density;  $V_f$  = Tapped density.

#### E. Angle of repose

The fixed funnel method was employed to measure the repose angle. A funnel was secured with its tip at a given height, H above a graph paper that was placed on a flat horizontal surface. The blend was carefully pored through the funnel until the apex of the conical pile just touched the tip of the funnel. The radius, R, of the base of the conical pile was measured. The angle of repose,  $\alpha$ , was calculated using the following formula.

$$\alpha = \tan^{-1} H/R$$

#### Determination of physical parameters of floating tablets [3,6,7]

##### Weight variation test

Totally, 20 tablets from each batch were individually weighed in grams on an analytical balance. The average weight and standard deviation were calculated, individual weight of each tablet was also calculated using the same and compared with average weight.

##### Thickness test

The thickness in millimeters (mm) was measured individually for 10 pre-weighed tablets by using Vernier Calipers. The average thickness and standard deviation were reported.

##### Hardness test

Tablet hardness was measured using a Monsanto hardness tester. The crushing strength of the 10 tablets with known weight and thickness of each was recorded in  $\text{kg}/\text{cm}^2$  and the average hardness, and the standard deviation was reported.

##### Friability test

Totally, 20 tablets were selected from each batch and weighed. Each group of tablets was rotated at 25 rpm for 4 minutes (100 rotations) in the Roche friabilator. The tablets were then dusted and re-weighed to determine the loss in weight. Friability was then calculated as per weight loss from the original tablets.

##### Determination of drug content

Ten tablets with pre-determined weight from each batch were taken and crushed in a mortar and weight equivalent to one average tablet was taken, transferred to a 250 mL volumetric flask and 0.1N HCL was added. The volume was then made up to the mark with 0.1N HCL. The solution was filtered, and the filtrate was sufficiently diluted, and the

absorbance was recorded against the blank at 272 nm. The drug content of the standard containing the drug powder was also determined. The drug content was determined by the formula [12-14].

$$\text{Drug content} = \frac{\text{Amount in test}}{\text{Amount in standard}} \times 100$$

The tablet passes the requirements if the amount of the active ingredient in each of the 10 tested tablets lies within the range of 85-115% of the stated amount.

#### Ex-vivo bioadhesive strength [12-16]

This evaluation test was conducted for all formulations. There is a gradual increase in bioadhesion strength was observed in from F4 to F6. This is due to the increase in the concentration of mucoadhesive polymer carbopol 974P. Here, the study investigates the mucoadhesive properties of formulations from F4 to F6. The maximum bioadhesion strength was found for formulations F4-FH 6, respectively. Bioadhesion is defined as the attachment of a synthetic or natural macromolecule to mucus and/or an epithelial surface. Bioadhesion strength is depends on molecular weight and swelling behavior of the polymers, contact time with mucus. As the concentration of carbopol increased the bioadhesive strength was also increased, the reason for such findings might be the formation of secondary bioadhesion bonds with mucin and interpenetration of the polymer chains in the interfacial region, while other polymers undergo superficial bioadhesion. Bioadhesion strength values of all the formulations represented in Table 2.

#### In vitro buoyancy studies [16-20]

The *in vitro* buoyancy (n=3) was determined by floating lag times according to the method described by Rosa *et al.* The tablets were placed in a beaker containing 100 mL of 0.1N HCL. The time required for the tablet to rise to the surface and float was taken as floating lag time. Total floating time was also measured and shown in the results section in Table 3 and Fig. 1a-d.

#### In vitro drug release studies [17-24]

The release rate of hydrochlorothiazide floating tablets was determined using USP Type 2 apparatus. The dissolution test was performed in triplicate, using 900 mL of 0.1N HCL, at 37±0.5°C at 50 rpm for 12 hrs. A 5 mL sample was withdrawn from the dissolution apparatus at specified time points, and the samples were replaced with fresh dissolution medium. The samples were filtered through a 0.45 µm membrane filter and diluted if necessary. The absorbance of these solutions was measured at 272 nm using Elico SL -159, UV-visible spectrophotometer. The cumulative drug release was calculated using the equation ( $y = 0.03x + 0.024$ ) generated from Beer Lambert's Calibration curve in the linearity range of 1-10 µg/mL.

#### Kinetic analysis of dissolution data [13-17]

To analyze the *in vitro* release data various kinetic models were used to describe the release kinetics. The zero order rate equation 1 describes the systems where the drug release rate is independent of its concentration. The first order equation 2 describes the release from a system where release rate is concentration dependent. Higuchi described the release of drugs from insoluble matrix as a square root of time dependent process based on Fickian diffusion equation 3. The Hixson-Crowell cube root law equation 4 describes the release from systems where there is a change in surface area and diameter of particles or tablets [13-17].

$$C = K_0 t \quad (1)$$

Where  $K_0$  is zero-order rate constant expressed in units of concentration/time and  $t$  is the time.

$$\text{Log}C = \text{Log}C_0 - K_1 t/2.303 \quad (2)$$

Table 2: Bioadhesive strength (n=3) of all formulations

Formulation code	Bio adhesion strength (g)	Force of adhesion (N) in dyne
F1	9.4±0.28	0.92
F2	10.1±0.52	0.99
F3	15.6±0.39	1.53
F4	43.6±0.21	4.27
F5	44.2±0.36	4.33
F6	45.4±0.27	4.45
F7	22.5±0.15	2.20
F8	21.4±0.37	2.09
F9	24.2±0.46	2.37
F10	26.6±0.31	2.60
F11	28.2±0.42	2.76
F12	29.6±0.25	2.90

Table 3: In vitro buoyancy studies

Formulation code	Floating lag time (seconds)	Total floating time (hrs)
F1	75	>12
F2	82	>12
F3	76	>12
F4	70	>12
F5	89	>12
F6	84	>12
F7	90	>12
F8	75	>12
F9	84	>12
F10	79	>12
F11	87	>12
F12	86	>12

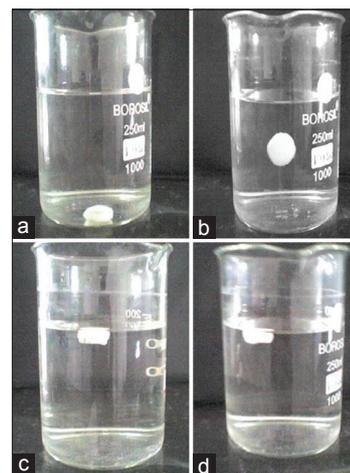


Fig. 1: In vitro buoyancy study of formulation F4. (a) At initial time, (b) after 34 seconds, (c) after 45 seconds, (d) after 12 hrs

Where  $C_0$  is the initial concentration of drug and  $K_1$  is the first order constant.

$$Q = K_H t^{1/2} \quad (3)$$

Where  $K_H$  is the constant reflecting the design variables of the system.

$$Q_0^{1/3} - Q_t^{1/3} = K_{HC} t \quad (4)$$

Where  $Q_t$  is the amount of drug remained in time  $t$ ,  $Q_0$  is the initial amount of the drug in tablet, and  $K_{HC}$  is the rate constant for Hixson-Crowell rate equation. Kinetic results were shown in the Table 4.

**Table 4: Kinetic values obtained from *in vitro* released data of formulation F4**

Kinetic model	Slope	R <sup>2</sup>
Zero-order plot	8.3466	0.9366
First-order plot	-0.0041	0.9364
Higuchi plot	29.935	0.9974
Korsmeyer-Peppas	0.7212	0.9680
Hixon-crowel	0.0298	0.9283

**Standard graph of doxofylline**

The standard graph of doxofylline in 0.1N HCL showed a good linearity with R<sup>2</sup> of 0.999, in the concentration range of 0-32 µg/mL at 272 nm shown in the Fig. 2.

**Properties of the powder blend**

All formulations were evaluated for compressibility index, angle of repose, and Hausner ratio. The results indicated the pre-compressed blend gas good flow shown in Table 5.

**Table 5: Evaluation of precompression parameters**

Formulation code	Evaluation of precompression parameters					
	Bulk density (g/mL)	Tapped density (g/mL)	Porosity (%)	Carr's index (%)	Hausner ratio	Angle of repose (°)
F1	0.489	0.604	0.200	20.11	1.229	35.55
F2	0.488	0.600	0.201	19.19	1.222	33.43
F3	0.479	0.610	0.199	19.99	1.221	33.33
F4	0.482	0.607	0.207	20.12	1.122	39.61
F5	0.488	0.603	0.190	19.07	1.23	37.13
F6	0.481	0.604	0.204	20.36	1.25	32.16
F7	0.488	0.606	0.222	19.99	1.299	33.66
F8	0.545	0.704	0.225	16.25	1.194	29.14
F9	0.504	0.601	0.199	22.64	1.29	30.15
F10	0.500	0.606	0.190	21.20	1.999	28.99
F11	0.501	0.605	0.195	20.65	1.385	35.54
F12	0.535	0.607	0.204	19.91	1.229	32.89

**Table 6: Evaluation of post-compression parameters**

Formulation	Evaluation of post-compression parameters				
	Hardness of tablets* (kg/cm <sup>2</sup> )	Friability of tablets* (%)	Weight variation of tablets* (mg) (%)	Thickness of tablets* (mm)	Drug content (%)
F1	5.51±0.01	0.23±0.05	300±2	5.12±0.02	96.28
F2	4.54±0.06	0.21±0.06	300±2	4.52±0.05	97.23
F3	4.12±0.03	0.20±0.05	300±2	4.23±0.01	99.12
F4	4.35±0.09	0.17±0.04	300±2	6.13±0.03	98.85
F5	5.42±0.02	0.19±0.03	300±2	4.0±0.06	99.54
F6	4.58±0.08	0.20±0.08	300±2	4.4±0.08	99.43
F7	4.31±0.19	0.23±0.05	300±2	4.2±0.01	98.67
F8	4.47±0.11	0.21±0.06	300±2	5.5±0.02	98.97
F9	4.49±0.14	0.20±0.05	300±2	5.3±0.04	98.28
F10	5.4±0.12	0.20±0.05	300±2	5.2±0.03	99.43
F11	4.33±0.21	0.22±0.07	300±2	4.35±0.06	98.12
F12	4.30±0.04	0.21±0.03	300±2	5.7±0.05	99.48

\*Average (n=3)

**Table 7: *In vitro* drug release study of various formulations**

Time	Various formulation of % drug release											
	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10	F11	F12
0	0	0	0	0	0	0	0	0	0	0	0	0
0.5	13.2	10.6	9.6	15.8	12.8	11.7	13.6	11.2	9.7	12.8	11.3	9.9
1	17.3	14.8	11.7	23.1	17.6	15.1	20.7	10.3	10.2	15.6	13.5	12.6
2	20.6	16.4	12.9	31.3	21.9	18.6	28.3	14.7	12.9	19.2	17.8	14.8
3	26.9	22.9	17.3	40.8	31.9	22.7	34.5	20.9	15.7	24.9	20.8	18.7
4	37.4	34.3	23.5	47.3	39.1	29.3	40.7	26.8	21.6	35.8	31.9	28.4
5	42.1	39.1	32.6	55.1	46.9	34.4	51.6	32.8	29.1	40.7	36.3	31.8
6	54.3	44.3	38.4	62.7	51.9	41.8	58.6	37.5	36.5	52.8	41.4	38.9
7	67.5	51.6	43.7	70.4	57.6	54.4	64.4	44.6	41.6	65.2	48.8	47.6
8	73.7	59.9	57.3	76.3	64.4	58.5	69.4	51.8	54.6	72.6	57.3	53.7
9	79.2	67.7	62.9	81.9	70.6	63.4	76.2	59.8	60.4	78.3	64.7	60.8
10	82.5	74.2	69.3	86.5	77.5	69.2	81.7	67.8	67.2	81.6	70.8	69.7
11	87.8	78.5	73.1	90.9	81.6	74.3	87.2	76.5	70.6	86.5	76.3	72.3
12	90.1	81.3	78.1	98.8	87.6	80.8	94.8	83.2	78.9	90.8	84.7	73.7

\*Average (n=3)

**Drug-excipient interaction studies**

*Fourier transform infrared spectroscopic studies (FTIR)*

The FTIR spectra of drug and optimized formulation were recorded in Figs. 3 and 4. The characteristic peaks of the optimized formulation followed the same trajectory as that of the drug alone with minor differences. Thus, there may be no drug-excipient interactions.

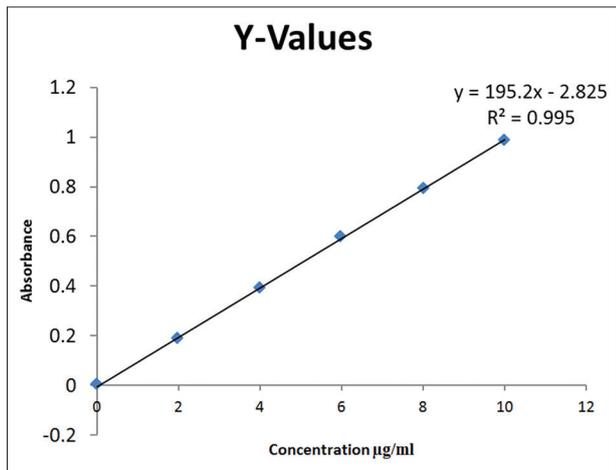


Fig. 2: Standard calibration curve of hydrochlorothiazide at  $\lambda_{max}$  272 nm

**Evaluation of the prepared tablets for physical parameters**

All formulations were tested for physical parameter such as hardness, thickness, weight variation, friability, and drug content. All estimated parameters were found to be within the limits shown in Table 6.

**In vitro dissolution studies**

Based on the dissolution release studies keeping in view of bioadhesion strength formulation F4-F6 were considered to be in good floating as well as bioadhesion property based on similarity factor with innovator drug dissolution profile formulae F4 were considered as optimized formulae dissolution release pattern observed in Table 7 and dissolution profiles seen in Fig. 5.

Table 8: Comparison of drug profile of optimized batch with innovator

Batch No.	Similarity factor (F2)	Difference factor (F1)
F1	73.8	26.6
F2	73.9	26.1
F3	73.9	26.1
F4	73.71	25.3
F5	73.73	25.5
F6	78.64	24.5
F7	79.66	24.6
F8	84.29	16.81
F9	76.22	24.7
F10	75.6	25.1

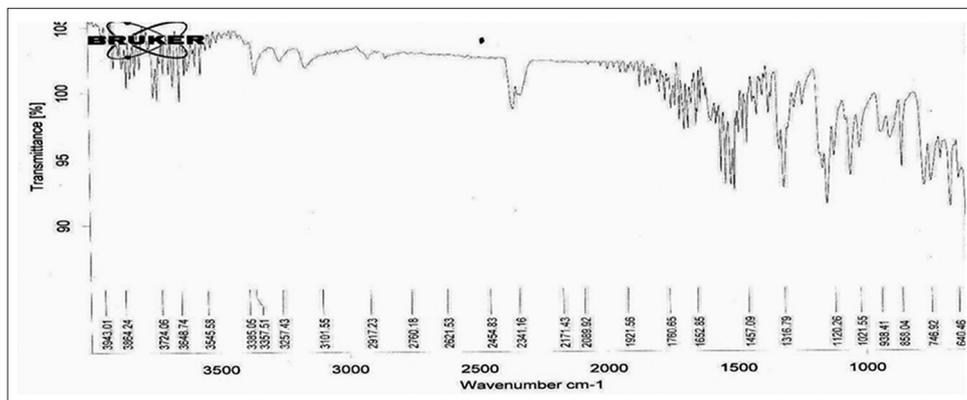


Fig. 3: Fourier transform infrared spectrum of hydrochlorothiazide pure drug

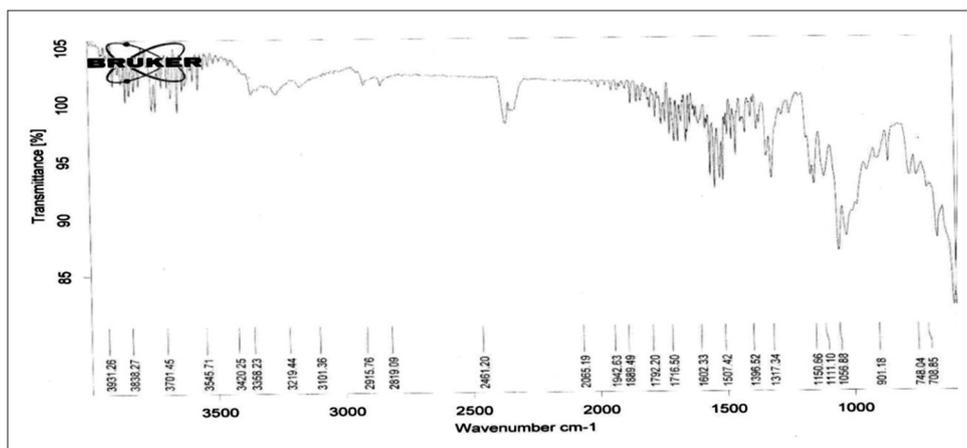


Fig. 4: Fourier transform infrared of hydrochlorothiazide best formulation F4

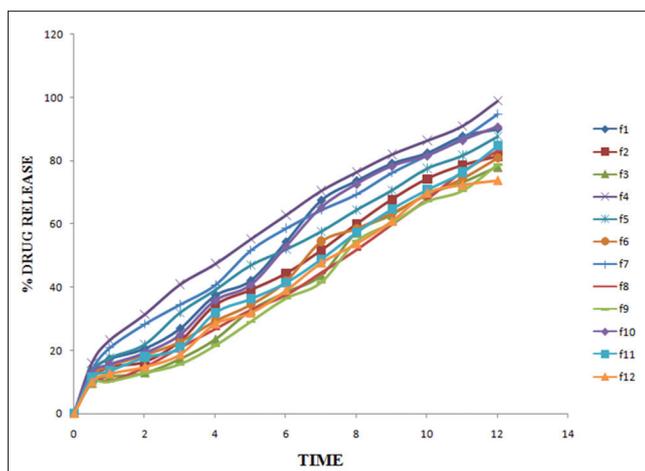


Fig. 5: Dissolution profile graphs

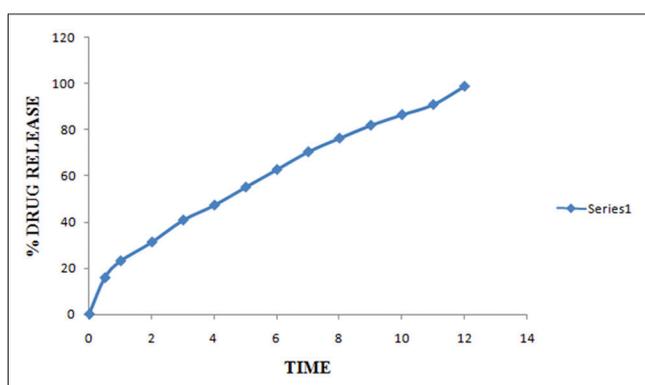


Fig. 6: Best % drug release F4

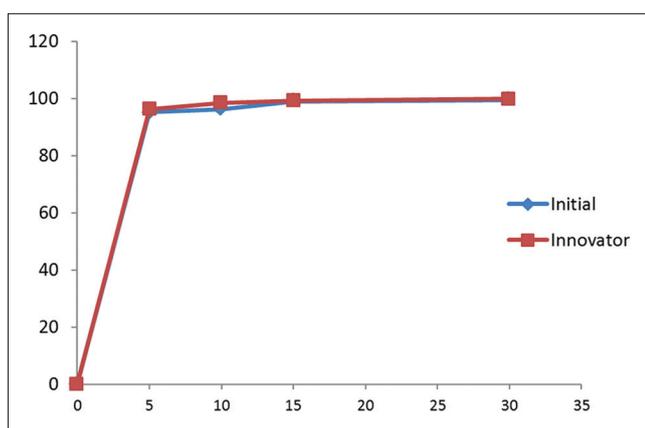


Fig. 7: Graphical representation of dissolution profile of innovator versus optimized batch (F4) initially

#### Data treatment

From Table 8, similarity factor for F4 formulation was found to be more, i.e., 84.29 compared to other formulations and difference factor was found to be less, i.e., 16.81 compared to other formulations. Therefore, F4 formulation said to be comparable with that of innovator product where the similar matching dissolution profiles can be seen in Figs. 6 and 7.

#### CONCLUSION

From the compatibility studies, it was concluded that HPMC K15, K100, xanthan gum, carbopol 974 p, sodium bicarbonate, magnesium stearate, talc, and microcrystalline cellulose were compatible

with hydrochlorothizide and thus suitable for the formulation of hydrochlorothizide floating tablets. *In vitro* buoyancy studies were performed for all the formulations, F1-F12 by using 0.1 N HCL solution at 37°C. All the formulations were floated. The formulation F4-F6 containing of HPMC K 4M and carbopol 974 P showed more Bioadhesive strength than other formulations. *In vitro* dissolution studies were also performed for all formulations. The formulation F4 showed the controlled release for 12 hrs. Thus, F4 was identified as ideal batch based on its results.

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