

PROFILE OF ANTIBACTERIAL ACTIVITY OF FRACTIONS FROM METHANOL EXTRACTS OF *GARCINIA LATISSIMA* MIQ. FRUIT RINDNENENG SITI SILFI AMBARWATI^{1,2}, AMARILA MALIK^{1*}, BERNA ELYA¹, MUHAMMAD HANAFI³

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ABSTRACT

Objectives: A previous study showed that methanol extracts of *Garcinia latissima* Miq. demonstrated antibacterial activity against *Bacillus subtilis*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. The aim of this study was to obtain active antibacterial fractions from methanol extracts of *G. latissima* Miq.

Methods: Fractionation of extracts was performed on G60 silica gel column chromatography using n-hexane eluent, ethyl acetate, and methanol. Antibacterial tests were done using the paper disc method to determine the zone of inhibition, the microdilution method to determine the minimum inhibitory concentration (MIC), and a bioautographic test.

Results: Fractions A-E, and F had zones of inhibition against *B. subtilis*. Fractions A-E had zones of inhibition against *S. aureus*. Fractions C-E had zones of inhibition against *P. aeruginosa*. The bio-autograph test showed zones of inhibition on several bio-autographic spots, indicating that active compounds were obtained from the fractionation of methanol extract from *G. latissima* Miq. fruit rind. Fraction D's MIC against *B. subtilis*, *S. aureus*, and *P. aeruginosa* was 312.5 ppm. The MIC of Fraction C against *B. subtilis* and of Fraction R against *P. aeruginosa* also was 312.5 ppm.

Conclusion: Fraction D was the most active fraction against the three test bacteria.

Keywords: *Garcinia latissima* Miq., *Bacillus subtilis*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, Zone of inhibition, Minimum inhibitory concentration, Bio-autograph.

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INTRODUCTION

Many diseases are caused by bacteria. 72% of 50 medicinal plants belonging to 26 families have been found to have antibacterial activity [1]. Based on gram staining, bacteria are classified into Gram-negative or Gram-positive bacteria, such as *Pseudomonas aeruginosa* [2]. Previous studies have shown that methanol extract of *Garcinia latissima* Miq. rind has antibacterial activity against *Bacillus subtilis*, *Staphylococcus aureus*, and *P. aeruginosa* [3].

Fractionation is the process of separating compounds based on their polarity. Fractionation can be done using column chromatography. Column chromatography involves liquid-solid adsorption and requires packing columns with adsorbents such as silica gel, wherein the sample is eluted using a solvent. Adsorption will stop when the concentration of a substance in solution and the mass absorbed in the stationary phase reach equilibrium [4]. The use of mixed solvents of different polarity will result in a partition mechanism [5].

Antibacterial tests can be performed by the diffusion method, which is based on the ability of a test substance to diffuse on an agar medium containing the test microbe, and the dilution method, which involves mixing the test substance with the test bacteria in a liquid medium. The bio-autographic test is also easy to use, cost-effective, fast, and able to assess the antibacterial activity of a considerable sample [6].

METHODS

Fractionation of the extract was performed using column chromatography with 300 g silica gel in stationary phase with 25 g extract samples. The motion phases of n-hexane, ethyl acetate, and methanol were used, gradually increasing the polarity. The elution

results were collected in a 100-mL bottle. The concentrate fractions have been tested used thin layer chromatography (TLC) and fractions with the same TLC profiles were combined, resulting in eight fractions.

The inhibition test of the fractions used nutrient agar medium for *B. subtilis* and *S. aureus* and cetrimide for *P. aeruginosa*. The fraction was dissolved in dimethyl sulfoxide (DMSO) at a concentration of 20,000 ppm. 1 mL of bacterial suspension with 106 microbes/mL was placed into a tube containing 4 mL of antibiotic media that was liquefied by heating at 45-60°C. After being shaken with a vortex until homogeneous, solutions were poured into Petri dishes containing 20 mL of solid nutrient media. The Petri dish, with a top layer of seed, was shaken slowly and then allowed to freeze. 6-mm diameter sterile paper discs were placed on the Petri dishes and added with 20 µL of fraction solution (triplo). Then, Petri dishes for *S. aureus* and *B. subtilis* were incubated at 37°C for 24 hrs, while *P. aeruginosa* was incubated at room temperature for 48 hrs. The diameter of the zone of inhibition was observed and measured using calipers [7].

The TLC bio-autographic test was performed using bilayer media [8]. Bacteria were inoculated on suitable media and then poured over solid agar medium. The eluted and dried TLC plates were attached to the agar layer and left for about 1 hr to allow the diffusion of the compound. After the TLC plate was removed, *S. aureus* and *B. subtilis* plates were incubated at 37°C for 24 hrs; meanwhile, *P. aeruginosa* was incubated at room temperature for 48 hrs. The zone of inhibition indicated the presence of active compounds functioning as antibacterial agents [9].

The minimum inhibitory concentrations (MICs) of fractions were determined using the microdilution method. Fractions were dissolved

in DMSO and diluted using bacterial media or broth. Each 50- μ L fraction solution was added with 50 μ L of inoculum containing bacteria at a concentration of 10^6 CFU/mL. From each well, the following fraction concentrations were obtained: 5,000 ppm (test), 2,500 ppm, 1,250 ppm, 625 ppm, 312,5 ppm, 156,25 ppm, and 78,125 ppm (triplo test). Microplates were incubated at 37°C for 24 hrs. *Pseudomonas aeruginosa* was incubated at room temperature for 48 hrs. After incubation, 0.6 mg/mL of MTT solution was added to a 10- μ L sterile Aqua dest [9] and then incubated for 20 minutes. Observations were performed visually by looking for the presence or the absence of discoloration. The presence of bacterial growth is characterized by a change in color from light yellow to purple or pink. The lowest concentration of non-discolored wells is the MIC [10].

RESULTS AND DISCUSSION

Fractionation results are presented in Table 1. Fraction F was the largest fraction, followed by Fraction E.

The first antibacterial test was the zone of inhibition test for the six resulting fractions of *G. latissima* Miq rind extract. The results are shown in Table 2.

The zone of inhibition test showed that all fractions were active against *B. subtilis* and *S. aureus*, except Fraction F, which did not show activity against *S. aureus*. Fractions C-E were active against *P. aeruginosa*. Although the concentration of fractions used for all tests was similar (20,000 ppm), the number of antibacterial agents diffusing on the agar surface was unknown. This is one deficiency of the zone of inhibition method, so this method is not appropriate for establishing MICs. A bio-autographic test was performed on the active fractions from the zone of inhibition test. The test results are shown in Figs. 1-3. The red circles indicated the presence of an inhibition area of bacterial growth resulting from the fraction eluted during TLC. The motion phases used in TLC were n-hexane:ethyl acetate (8:1) for Fraction A, dichloromethane:methanol

(8:1) for Fraction B, dichloromethane:methanol (8:1) for Fraction C, n-hexane:chloroform:ethyl acetate (1:1:2) for Fraction D, dichloromethane:methanol (6:1) for Fraction E, and dichloromethane:methanol (6:1) for Fraction F.

The presence of zones of inhibition indicated active compounds in the tested fractions. From the bio-autographic test results, semi-polar compounds were able to create a zone of inhibitions.

Antibacterial tests were also performed using the microdilution method for calculating the MIC. The results are shown in Table 3.

The results provided in Table 3 showed that the Fraction D of methanol extract from the fruit rind of *G. latissima* Miq had moderate activity against *B. subtilis*, *S. aureus*, and *P. aeruginosa*, with MIC values of 312.5 ppm. Fraction C against *B. subtilis* and Fraction E against *P. aeruginosa* had similar MIC values, which were 312.5 ppm. Fractions B and E had weak activity against *B. subtilis*, with MIC values of 625 ppm. Fractions A-C, and E also had weak activity against *S. aureus*, with MIC values of 625 ppm. Fractions B and C, and F had weak activity against *P. aeruginosa* with MIC values of 625 ppm. Fraction A was not active against *B. subtilis* and *P. aeruginosa* because the MIC values were 1.250 ppm. Fraction F was also not active against *B. subtilis* and *S. aureus* because the MIC values were 1.250 ppm.

These values were in accordance with the MIC reference values for antimicrobial activity. If <100 ppm, the antimicrobial activity is strong. If the MIC value is between 100 and 500 ppm, antimicrobial activity is moderate. An MIC value between 500 and 1,000 ppm is weak, while more than 1,000 ppm indicates inactivity in terms of antimicrobial activity [11]. The antimicrobial activity of these tested fractions was compared with standard drugs, including erythromycin for *B. subtilis* and gentamicin for *P. aeruginosa*. The results of this study indicated that one fraction from the methanol extract of *G. latissima* Miq. rind that exhibited moderate activity in inhibiting the growth of *B. subtilis*, *S. aureus*, and *P. aeruginosa* bacteria.

CONCLUSION

Fraction D was the most active fraction of the three tested bacteria. The MIC test of fractions from ethyl acetate of *G. latissima* Miq. rind can be further explored through isolating active fractions, which could play an important role in the discovery of new medicines. Further studies are needed, including a toxicity test, to purify active compounds from *G. latissima* fractions with potential use as antibacterial agents in the pharmaceutical industry.

Table 1: Fractionation results from methanol extracts of *G. latissima* Miq. fruit rind

Fraction	Bottle number	Fraction weight (g)	% Fraction
A	1-42	0.1825	13.1492
B	43-70	0.0812	0.5850
C	71-77	0.2085	1.5022
D	78-84	0.4414	3.1903
E	85-112	3.6210	26.0894
F	113-148	9.3446	67.328
Total weight of fraction		13.8792	

G. latissima: *Garcinia latissima*

Table 2: Zone of inhibition results from 20,000 ppm methanol fractions of *G. latissima* rind

Fraction	Diameter of zone of inhibition (mm) of bacteria		
	<i>B. subtilis</i>	<i>P. aeruginosa</i>	<i>S. aureus</i>
A	6.747 \pm 0.237	0.000	7.700 \pm 0.400
B	7.110 \pm 0.608	0.000	7.867 \pm 0.379
C	8.140 \pm 0.078	6.167 \pm 0.058	6.800 \pm 0.200
D	8.243 \pm 0.645	6.933 \pm 0.058	6.633 \pm 0.379
E	7.903 \pm 0.291	6.167 \pm 0.058	6.367 \pm 0.208
F	7.017 \pm 0.770	0.000	0.000
Antibiotic standard	29.175 \pm 0.983	14.250 \pm 0.693	24.855 \pm 0.021
Negative control	0	0	0

B. subtilis: *Bacillus subtilis*, *P. aeruginosa*: *Pseudomonas aeruginosa*, *S. aureus*: *Staphylococcus aureus*

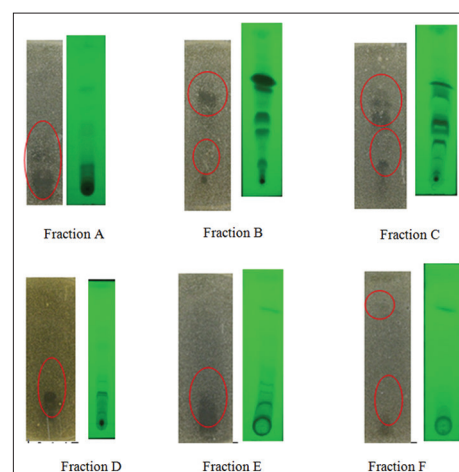


Fig. 1: Bio-autographic results of active fractions from *Garcinia latissima* Miq. rind methanol extract against *Bacillus subtilis*

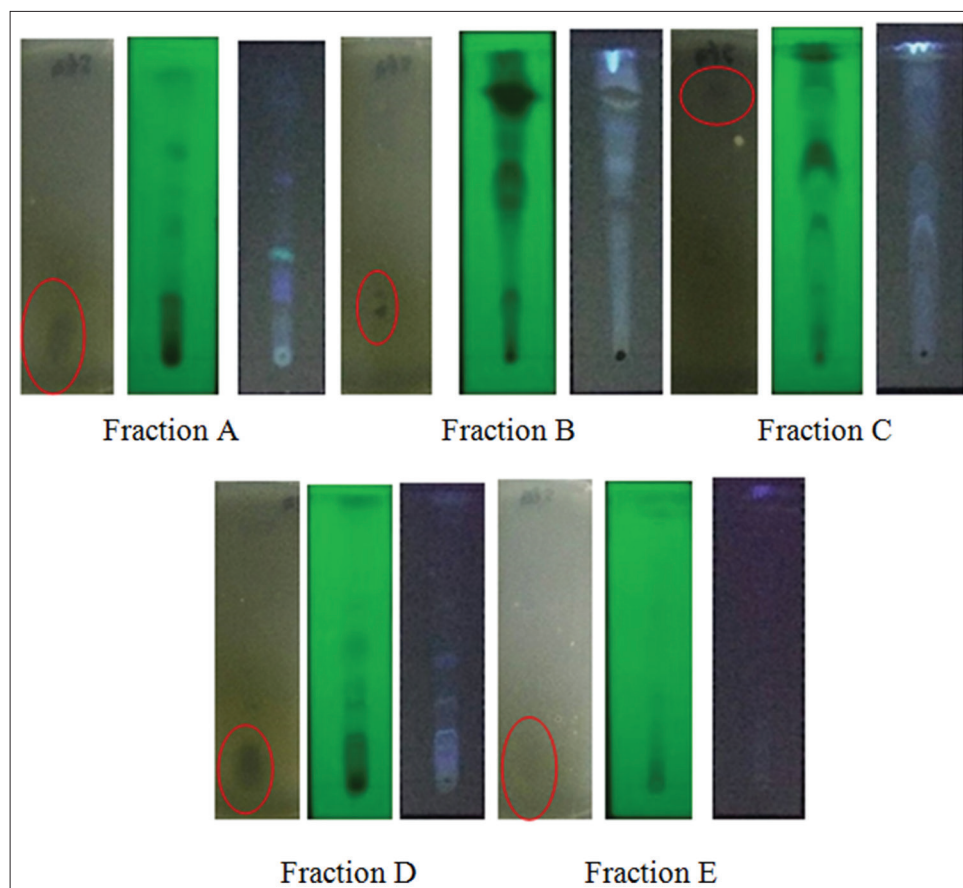


Fig. 2: Bio-autographic results of active fractions from *Garcinia latissima* Miq. rind methanol extract against *Staphylococcus aureus*

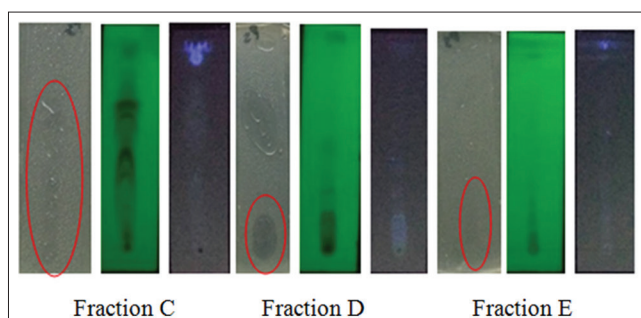


Fig. 3: Bio-autographic results of active fractions from *Garcinia latissima* Miq. rind methanol extract against *Pseudomonas aeruginosa*

Table 3: MIC results of fractions from *G. latissima* Miq. rind methanol

Fraction	<i>B. subtilis</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>
A	1.250	625	1.250
B	625	625	625
C	312.5	625	625
D	312.5	312,5	312.5
E	625	625	312.5
F	1.250	1.250	625
Antibiotic standards	25		0.39

G. latissima: *Garcinia latissima*, MIC: Minimum inhibitory concentration, *B. subtilis*: *Bacillus subtilis*, *P. aeruginosa*: *Pseudomonas aeruginosa*, *S. aureus*: *Staphylococcus aureus*

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