

SYNTHESIS OF SOME NOVEL PERYLENE DI IMIDES AND EVALUATION OF THEIR ANTI CANCER ACTIVITY

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ABSTRACT

Objective: The main objective of the study is to synthesis some novel perylene di imides and to evaluate for anti oxidant activity and anticancer activity.

Methods: Antioxidant assay was carried on to study the reducing activity of the compounds. The cytotoxicity assay was studied to find the best potent compound among the synthesized compound by using the HCT-116, a colon cancer cell line. Synthesized substituted amine derivatives of perylene di imides. From the evaluation study Compound, A shows potent activity when compared with the standard drug 5-Fluorouracil.

Results: The results of the total antioxidant capacity assay of perylene compounds are evaluated by the 1,1-diphenyl-2-picryl hydrazyl (DPPH) method and nitric oxide scavenging method. All the synthesized compounds are evaluated for their antioxidant power. From the results of DPPH and nitric oxide scavenging assay, Compound A, B, C and D showed potent activity when compared with the standard. For further evaluation of cell line studies, based upon the IC₅₀ values, Compound A, B and C were taken for study. The molecular modeling data's are exactly correlated with the *in vitro* studies. We have used 5-Fluorouracil and PIPER as a standard for *in vitro* study and molecular modeling study respectively.

Conclusions: From the results, Compound A will be efficient to inhibit telomerase enzyme and the Compound A will be effective for anti-cancer therapy.

Keywords: Perylene derivatives, QSAR plus, G-Quadruplex ligand database, Docking, Anti oxidant study, Cell line study.

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INTRODUCTION

G-Quartets are square planar arrangements of four guanine bases, which can form extraordinarily stable stacks when present in nucleic acid sequences [1,2]. Guanine-rich nucleic acids are well known for their ability to adopt non-Watson-Crick hydrogen-bonded structures [3,4]. These structures are well-known as G-Quadruplexes and share the common feature of stacked guanine tetrads as basic motif [5]. G-Quadruplexes are four-stranded guanine-rich DNA structure found at the ends of the eukaryotic telomeres. Human telomeric DNA is usually 4–14-kilo bases long and is comprised of TTAGGG tandem repeats. Up-regulated telomerase activity in cancer cells maintains the length of telomeres after cell division, conferring immortality.

The role of G-Quadruplex and the telomerase activity has been studied since the early 1990s [6]. G-Quadruplex stabilization leads to inhibition of the telomerase activity, which induce the apoptosis. The compounds like perylene diimide (PDI), naphthalene diimide, oxazole, are the possible intercalators acting on the G-Quartet structure and stabilize it. For example, cationic porphyrin, quindoline and berberine, and tri-substituted are O-alloxazines have been demonstrated to interfere with the oncogenic transcription *in vitro*. Quarfloxin, developed by cylene pharmaceuticals, entered clinical trials due to its ability to interact with G-Quadruplexes *in vivo* [7-10].

QSAR study has been carried out by V Life MDS Software-QSAR Plus Module to predict and compare the biological activity of standard and newly designed compounds. QSAR has been done by developing variable regression methods. The compounds are divided into training and test set compounds by manual selection, random and sphere exclusion methods. The models are developed and based on regression values;

we selected eight equations to design new compounds of perylene di imides. Out of 497 compounds, 59 compounds possess better biological activity when compared with the standard compounds.

These 59 selected compounds from QSAR study has been chosen for docking study. Docking has been done by G-Quadruplex Ligand Database (G4LDB). This is an online database which was having in built tools and performed by open babel 2.3.0 to predict the binding affinity with the targets. The targets (1LIH, 1NZM, 3CE5, 3SC8 and 2HRI) for the docking are selected based upon

Compound 11



the literature survey and the selected compounds are docked. The results are visualized by Discovery studio Visualizer 4.1 Visualizer. From the results, 9 compounds are selected and the results of these compounds are visualized using Discovery Studio 4.1 Visualizer. To narrow down the results, 3 best compounds are selected and the

compounds are Compound 11, Compound 20 and Compound 48. The hydrogen bond interactions and binding free energy levels, pK_i values are compared with the standard PIPER compound.

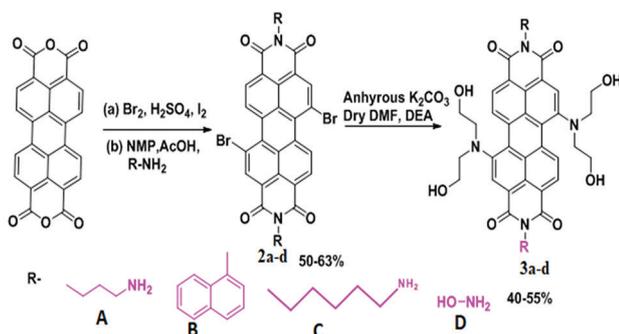
From the results of G4LDB Database, the 9 compounds are docked with the specific protein (PDB ID: 4B18), a telomerase protein. Docking has been done by Auto Dock 4.2. Finally, from the AutoDock results, Compound 11 shows good binding energy when compared with the standard PIPER compound. The study states as from the G4LDB Database 2 compounds possess good binding affinity, and from the Auto Dock Compound 11 possess good binding energy. As in our previous study from the *in silico* studies we studied structure activity relationship for the compounds and predicted biological activity, and by docking studies concluded the scaffold for the synthesis [11-13]. Molecular Docking Analysis was done by the output file of the docking that was generated after the study. The binding energy, Inhibition Constant, and the number of hydrogen bonds were considered for the analysis. Compound 11, 20 and 48 showed potent binding interactions with the protein 4B18 and compared with the standard PIPER.

The best compounds are compared with the standard PIPER and the binding energy values are showing more potent than the standard PIPER. The binding energy values for the compound 11, 20, 48 and PIPER are -6.23 , -6.42 , -6.59 and -5.59 kcal/mol respectively. From the scaffold, we selected Compound 11 as a scaffold moiety and sketched a scheme for the synthesis of Perylene di imides targeting telomerase for anti-cancer activity.

METHODS

Starting materials are all commercially available reagents and solvents used as received except for statements. All solvents were purified using standard procedures. Reactions were monitored by thin-layer chromatography on pre-coated silica gel plates and visualized using ultraviolet irradiation (254 nm). Column chromatography was performed on silica gel (100–200 Mesh).

Synthetic scheme



Procedure

Step 1: Bromination and condensation of PTCDA [14,15]

Preparation of dibromo-N,N-diamine substituted-3,4,9,10 perylene tetra carboxylic Diimides (2a,2b,2c and 2d)

To a solution of PTCDA compound (1.0 g, 20 mmol) was added concentrated sulfuric acid (200 mL) which was then stirred for 24 h at room temperature. The mixture was warmed to $55-60^{\circ}\text{C}$, and iodine (0.12 g, 2.5 mmol) was added. After 5 h, bromine (3.3 mL, 62.5 mmol) was added to the mixture slowly which was then warmed to 80°C and stirred for 8 h. The mixture was quenched with ice water and filtered under reduced pressure to give bromo compound. The crude Bromo Compound (500 mg, 0.9 mmol), amines (R_1, R_2, R_3 and R_4) and acetic acid (272 mg, 4.5 mmol) in *N*-methyl-2-pyrrolidinone (NMP; 15 mL) were stirred at 85°C under N_2 for 12 h. After cooling the mixture to room

temperature and pouring into aqueous HCl (10 vol.-%, 100 mL), the precipitate was separated by suction filtration, washed with deionized water until pH 7, washed and dried under vacuum. The crude product was purified by silica gel column Hexane-ethylacetate 4:1, v/v, $R_f = 0.71$ - Compound 2a, $R_f = 0.61$ - Compound 2b, $R_f = 0.42$ - Compound 2c, $R_f = 0.61$ - Compound 2d) as eluent and Compound 2a, 2b 2c and 2d was obtained after evaporation of the solvent as a red powder.

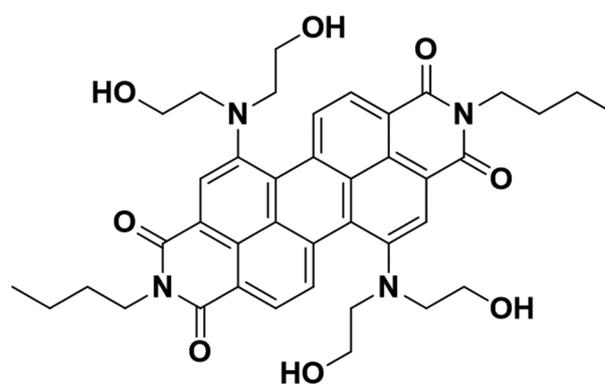
Step 2: Alkylation of substituted and condensed dibromo PTCDA

Preparation of 1,7-Bis(*n*-diethanolamine)-*N,N*-disubstituted amines-3,4,9,10-perylene-tetracarboxylic diimides (3a,3b,3c and 3d)

1,7-dibromo PDI (2a,b,c and d 0.1 mmol) was dissolved into 5 mL of dimethylformamide. To which diethylacetamide (0.5 mmol) and potassium carbonate (K_2CO_3 , 0.5 mmol) was added. The resulted mixture was then allowed reacted at 80°C for 15 h. The reaction mixture was then powered into 15 mL water and the red solid was then re-dissolved in 20 mL dichloromethane (DCM) and washed with 1N hydrochloric acid and then water each for 3 times. Then, DCM layer was dried over Na_2SO_4 . After removal of DCM, the residue was applied to chromatography with CH_2Cl_2 /ethyl acetate (100:0–100:2) as eluents to afford the desired products 3a, 3b, 3c and 3d. The final synthesized compounds are tabulated in Table 1.

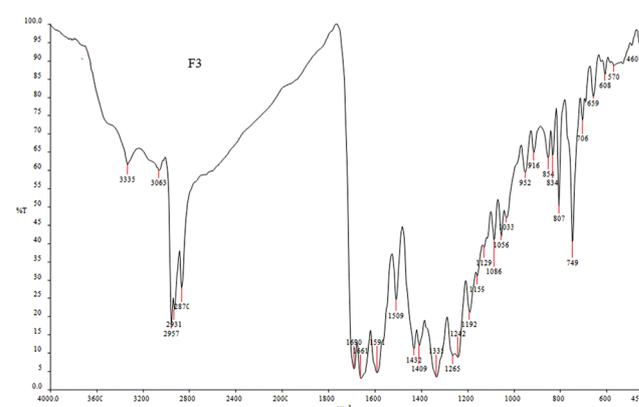
All the compounds are Characterize by IR, ^1H nuclear magnetic resonance (NMR), ^{13}C NMR and high resolution-mass spectrometry HRMS (electrospray ionization [ESI]). The results are as following.

Spectral results-Compound A Step 2:1,7-bis(*n*-diethanolamino)-*N,N*-dibutyl-3,4,9,10-perylene-tetracarboxylic diimides-Compound A



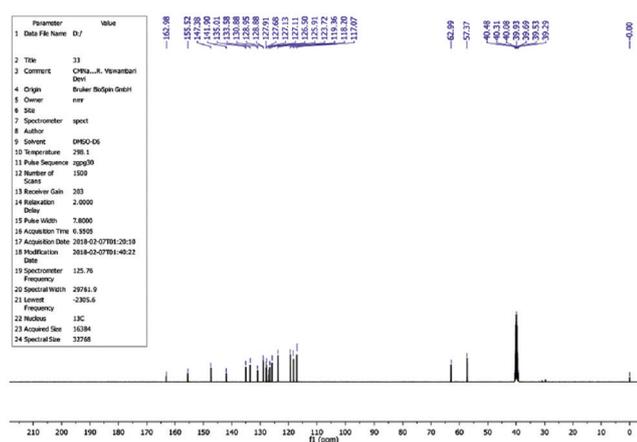
$R_f = 0.64$ (H: EtOAc/1:1);

IR spectra



Result analysis

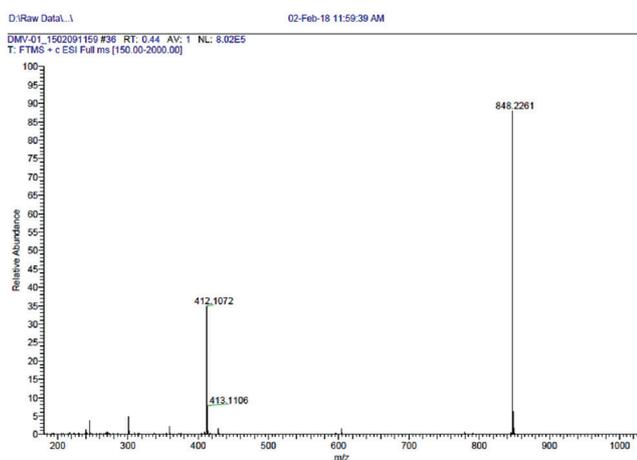
IR (KBr, cm^{-1}) Hydroxyl group (3335 cm^{-1}), Amide carbonyl ($1710, 1690 \text{ cm}^{-1}$), Aromatic CH ($3063, 2957 \text{ cm}^{-1}$), Aliphatic CH (2870 cm^{-1}), C=C (1591 cm^{-1});

¹³CNMR Spectra- 3b (Compound B)

Result analysis

¹³C NMR (100 MHz, CDCl₃) δ 162.98, 155.52, 147.38, 141.90, 135.01, 133.58, 130.88, 128.95, 128.88, 127.91, 127.68, 127.13, 127.11, 126.50, 125.91, 123.72, 119.36, 118.20, 117.07, 62.99, 57.37 ppm;

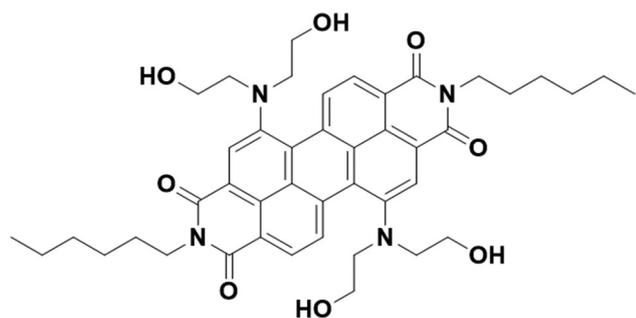
HRMS (ESI) Spectra - 3b (Compound B)



Result analysis

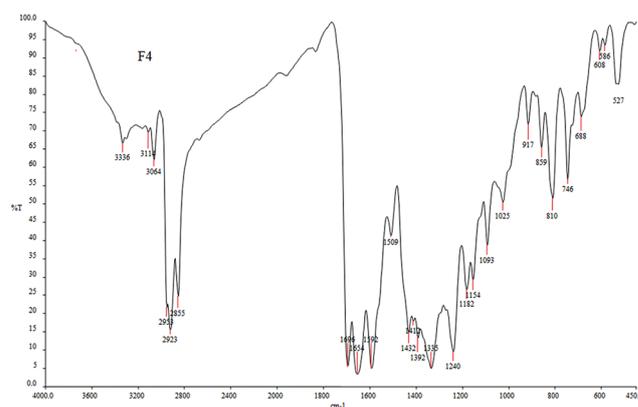
HRMS (ESI): Anal. Calcd. for (C₅₂H₄₀N₄O₈) (M⁺): 848.2846, Found: 848.2261.

Spectral Results=Compound C Step 2: 5,12-bis(bis(2-hydroxyethyl)amino)-2,9dihexylanthra[2,1,9-def:6,5,10-d'e'f']disoquinoline-1,3,8,10(2H,9H)-tetraone - Compound C.



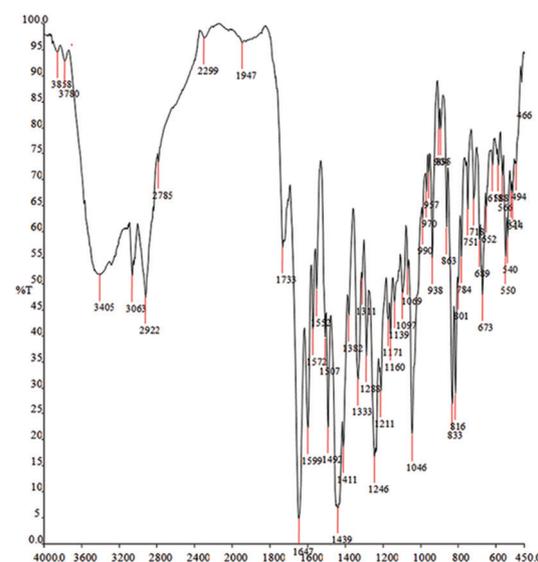
R_f=0.68 (H: EtOAc/1:1);

IR Spectra - 3c



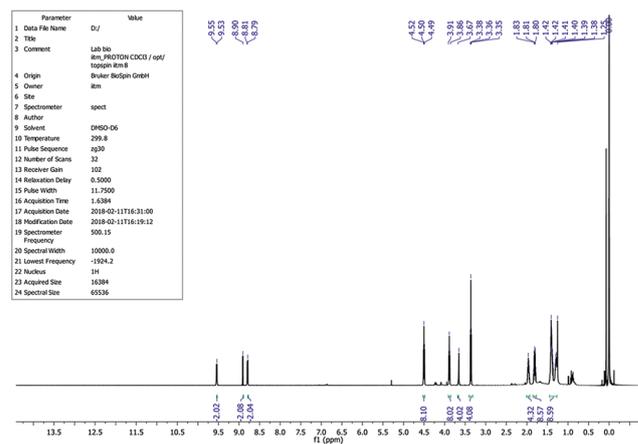
Result analysis

IR (KBr, cm⁻¹); Hydroxyl group (3405 cm⁻¹), Amide carbonyl (1733, 1647 cm⁻¹), Aromatic CH (3057 cm⁻¹), Aliphatic CH (2922 cm⁻¹), C=C (1599 cm⁻¹);

¹H NMR Spectra - 3c (Compound C)

Result analysis

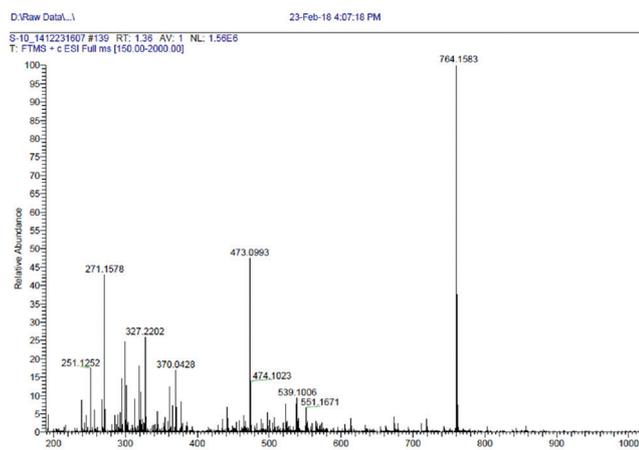
¹H NMR (400 MHz, CDCl₃) δ 9.55 (d, J=10Hz, 2H), 8.90 (s, 2H), 8.81 (d, J=10Hz, 2H), 4.52 (t, J=10Hz, 8H), 3.91 (t, J=5Hz, 8H), 3.67 (s, 4H), 3.38 (t, J=10Hz, 4H), 1.83-1.25 (m, 20H) ppm;

¹³CNMR Spectra - 3c (Compound C)

Result analysis

^{13}C NMR (100 MHz, CDCl_3) δ 162.65, 144.84, 140.82, 139.11, 137.73, 131.14, 129.25, 128.81, 127.71, 126.15, 125.68, 125.49, 124.22, 123.02, 119.53, 109.54, 61.72, 58.47, 41.58, 39.36, 32.74, 29.74, 27.11, 22.90, 14.40 ppm;

HRMS (ESI) Spectra - 3c (Compound C)

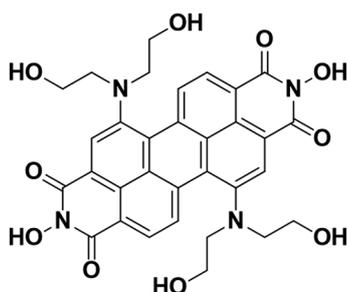


Result analysis

HRMS (ESI): Anal. Calcd. for $(\text{C}_{32}\text{H}_{28}\text{N}_4\text{O}_{10})$ (M+): 628.1805, Found: 628.7116.

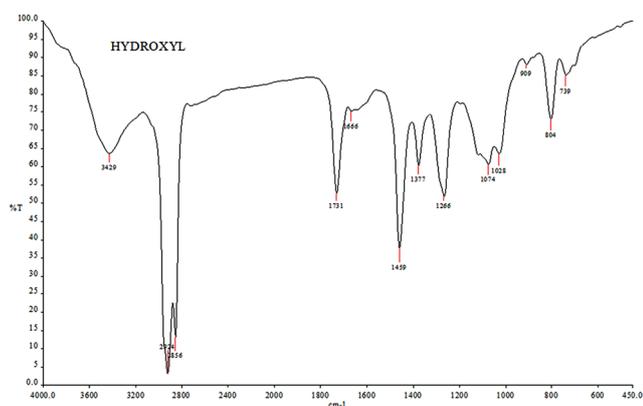
Spectral results

Compound D Step 2: 1,7-Bis(n-diethanolamino)-N,N-dihydroxylamine-3,4,9,10-perylenetetracarboxylic diimides (3d)-Compound D.



$R_f=0.70$ (H: EtOAc/1:1);

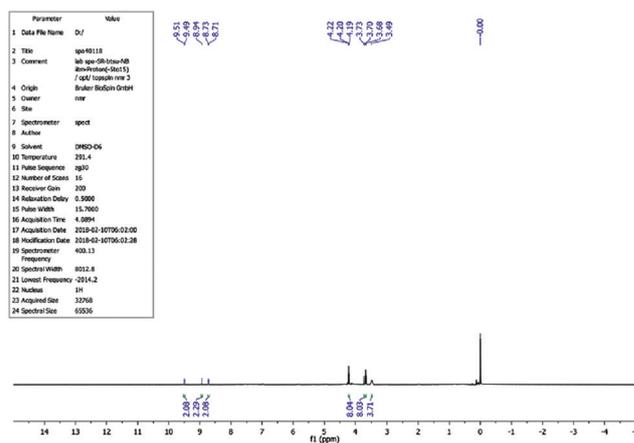
IR Spectra - 3d



Result analysis

IR (KBr, cm^{-1}): Hydroxyl group (3390 cm^{-1}), Amide carbonyl ($1727, 1667\text{ cm}^{-1}$), Aromatic CH ($3057, 2956\text{ cm}^{-1}$), Aliphatic CH (2857 cm^{-1}), C=C (1593 cm^{-1});

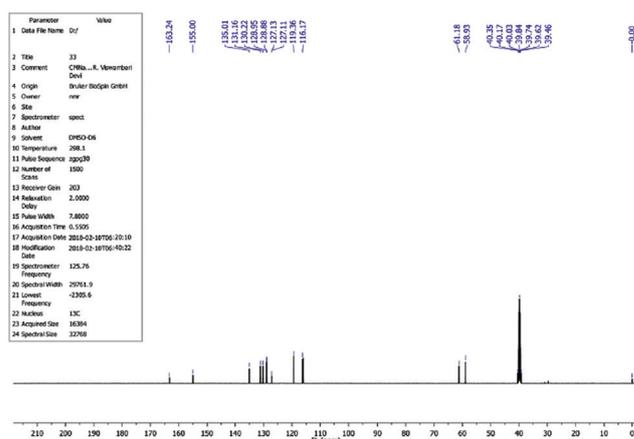
^1H NMR Spectra - 3d (Compound D)



Result analysis

^1H NMR (400 MHz, CDCl_3) δ ppm; δ 9.55 (d, $J=10\text{Hz}$, 2H), 8.90 (s, 2H), 8.81 (d, $J=10\text{Hz}$, 2H), 4.52 (t, $J=10\text{Hz}$, 8H), 3.91 (t, $J=5\text{Hz}$, 8H), 3.67 (s, 4H), 3.38 (t, $J=10\text{Hz}$, 4H).

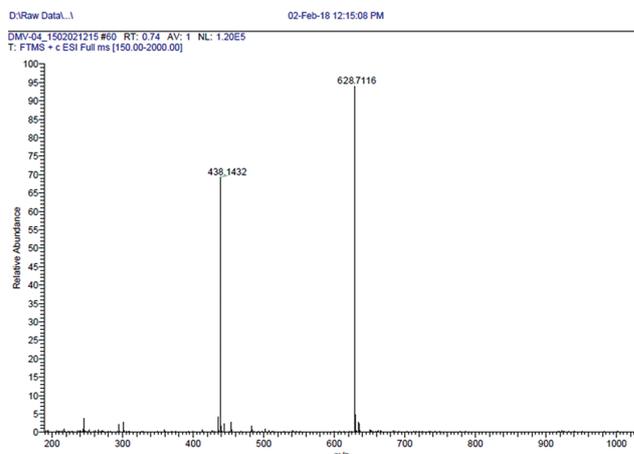
^{13}C NMR Spectra - 3d (Compound D)



Result analysis

^{13}C NMR (100 MHz, CDCl_3) δ 9.51 (d, $J=10\text{Hz}$, 2H), 8.94 (s, 2H), 8.73 (d, $J=10\text{Hz}$, 2), 4.22 (t, $J=10\text{Hz}$, 8H), 3.73 (t, $J=10\text{Hz}$, 8H), 3.49 (s, 4H) ppm;

HRMS (ESI) Spectra - 3d (Compound D)



Result analysis

HRMS (ESI): Anal. Calcd. for $(\text{C}_{32}\text{H}_{28}\text{N}_4\text{O}_{10})$ (M+): 628.1805, Found: 628.7116.

In vitro studies

Anti oxidant assay: Free radical scavenging ability by the use of a stable 1,1-diphenyl-2-picryl hydrazyl (DPPH) radical (DPPH)

The effect of the perylene di imides on DPPH radical was estimated according to the procedure described by Von Gadow *et al.* 2 mL of 6×10^{-5} M methanolic solution of DPPH was added to 50 μ L of a methanolic solution (10 mg/mL) of the sample. Absorbance measurements commenced immediately [16,17]. The decrease of absorbance at 515 nm was continuously recorded in a spectrophotometer for 16 min at room temperature all determinations were performed in triplicate.

Nitric oxide scavenging activity

Nitric oxide generated from sodium nitroprusside in aqueous solution at physiological pH was measured by Griess reaction [18,19]. The reaction mixture (3 mL) containing sodium nitroprusside (10 mm) in phosphate buffer saline and the test extract (10, 25, 50 and 100 μ g mL) was incubated at 25°C for 150 min, after incubation 1.5 mL of the reaction mixture was removed and 1.5 mL of the Griess reagent (1% sulphanilamide, 2% orthophosphoric acid and 0.1% naphthylethylene diamine hydrochloride) was added. The absorbance of the chromophore formed was read at 546 nm.

In vitro cell cytotoxicity estimation by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay

Cell line

The Human colon cancer cell line (HCT 116) was obtained from National Centre for Cell Science, Pune and grown in Eagles Minimum Essential Medium containing 10% fetal bovine serum [20,21]. The cells suspended in the medium by a gentle passage with the pipette and the cells homogenized. Seeding of cells One mL of the homogenized cell suspension was added to each well of a 24 well culture plate along with different concentration of butylamine, naphthalamine, hexylamine and hydroxylamine perylene derivatives (i.e., 0-250 μ g/mL) and incubated at 37°C in a humidified CO₂ incubator with 5% CO₂. After 48 h incubation, the cells were observed under an inverted tissue culture microscope. With 80% confluence of cells, cytotoxicity assay was carried out.

MTT assay

Cytotoxicity assay

The assay was carried out using MTT. MTT is cleaved by mitochondrial Succinate dehydrogenase and reductase of viable cells, yielding a measurable purple product formazan [22,23]. This formazan production is directly proportional to the viable cell number and inversely proportional to the degree of cytotoxicity. After 48 h incubation, the wells were added with MTT and left for 3 h at room temperature. All wells have removed the content using the pipette and 100 μ L safety data sheets in dimethyl sulfoxide was added to dissolve the formazan crystals, absorbances were read in Multiskan™ FC Microplate Photometer at 570 nm (Mosman 1983).

- The percentage cell viability was then calculated with respect to control as follows:
% Cell viability = $[A]_{\text{Test}}/[A]_{\text{control}} \times 100$
- The % cell inhibition was determined using the following formula.
% Cell Inhibition = $100 - \text{Abs (sample)}/\text{Abs (control)} \times 100$.

RESULTS

Molecular modeling study helps to open future de nova modeling of the new compound to treat cancer. Based on the *in silico* studies, we have chosen compound 11 as the scaffold moiety and to synthesis the perylene derivatives from the Scaffold. From the literature survey scheme for the synthesis have been drawn and bromination of perylene di imides has been done condensed with amines. Antioxidant assay and Anticancer activity showed Compound A has significant activity when compared with the standard Drug 5-Fluorouracil.

DISCUSSIONS

Anti oxidant Assay showed a maximum activity of 44.2% and 28.93% respectively at 1000 μ g/mL, whereas an ascorbic acid at the same concentration exhibited 97.40% inhibition respectively. Four different amines exhibited considerable DPPH free radical scavenging activity as indicated by their IC₅₀ values and this has been shown in (Table 2). IC₅₀ Indicate the potency of scavenging activity. Standard ascorbic acid found to have an IC₅₀ of 0.052 mg/mL. In comparison to standard ascorbic acid, butyl amine naphthylamine, hexyl and hydroxyl amine derivatives of perylene compounds showed of 5.311, 1.246, 2.611 and 1.753 respectively. Butyl and Hydroxyl derivatives are seen to have the least free radical scavenging activity. Nitric oxide scavenging assay has done and perylene compounds has potent nitric oxide scavenging activity of 42.31%, 37.18% & 26.92 % respectively at 1000 μ g/mL, where as Quercetin at the same concentration exhibited 85.96% inhibition respectively. These amines exhibited considerable Nitric Oxide scavenging activity as indicated by IC₅₀ value 1.462, 1.429 and 1.765 mg/mL for compound A, B and C, whereas Hydroxyl amine perylene derivative has showed the least nitric oxide scavenging activity of IC₅₀ value 4.147 mg/mL (Table 3). The scavenging of NO by the extracts was increased in dose dependent manner. A significant decrease in the NO radical due to the scavenging ability of the compounds and these are compared with Quercetin and the IC₅₀ was found to be 0.518 mg/mL.

In-vitro cell cytotoxicity analysis by MTT assay

The study was undertaken to determine the cell cytotoxicity of the synthesized compounds by MTT assay. HCT-116 cell line is a colon cancer cell line was taken and also the study helps to screen the best compound which is successfully able to kill the cancer cells [24,25]. Therefore, a cell viability graph was also plotted to understand the result. The study was designed with a single cell line (HCT-116) and different ligands in different concentrations. Therefore eight concentrations (0.1 μ M, 1 μ M, 10 μ M, 50 μ M and 100 μ M) for 24 h and 48 h, were fixed to understand the inhibition of the cell viability in a dose-response manner. 5-Fluorouracil was taken as a standard drug for this study, by injection into a vein it is used for colon cancer. The three prototypes of perylene amine derivatives are treated with Normal Vero cells to study the cell viability %. A viability assay is an assay to determine the ability of organs, cells or tissues to maintain or recover viability. The results of the three prototypes along with the standard are tabulated in Table 4. Among the three prototypes, only Compound A and Compound B showed immaculate activity in dose-response manner. Compound A successfully able to kill cancer cell and also showed 32.37 μ M IC₅₀ value. Compound A also showed potent cell population inhibition in 50 μ M and 100 μ M. The percentage of cell inhibition at the highest concentration was found to be 94.53%. The standard compound 5-Fluorouracil showed 26.923 μ M IC₅₀, which is proximately closer to the synthesized compound (Compound A). The concentration of drug-treated with HCY-116 colon cancer cells are plotted for three compounds (A, B and C) are compared with a standard drug. Compound B also successfully showed potent activity towards the cell line (HCT-116, colon cancer cell line). The IC₅₀ value of the compound was found to be 35.67 μ M. The percentage of cell inhibition at the highest concentration was found to be 91.79%. The standard compound 5-Fluorouracil showed 26.93 μ M IC₅₀, which is proximately nearer to the synthesized compound (Compound B). Regression analysis was also done. R² value of Compound A, Compound B and 5-Fluorouracil (Standard) were found to be 0.797, 0.790, and 0.809 respectively. Compound C doesn't have much potent towards the HCT-116 colon cancer cell line when compared with Compound A, B and with the standard Drug 5-Fluorouracil. The consolidated flowchart of inhibition of drug with respect to the concentration was depicted in Table 5 and Fig. 1. Colon cancer Cell lines treated with different concentrations of the compounds A, B, C and Standard drug (5-Fluorouracil) are represented in Fig. 2.

CONCLUSION

The study was undertaken to find out, a single novel molecule which can be used for multiple targets for different types of cancers. The

Table 1: Structure of novel perylene di-imides-final target molecules

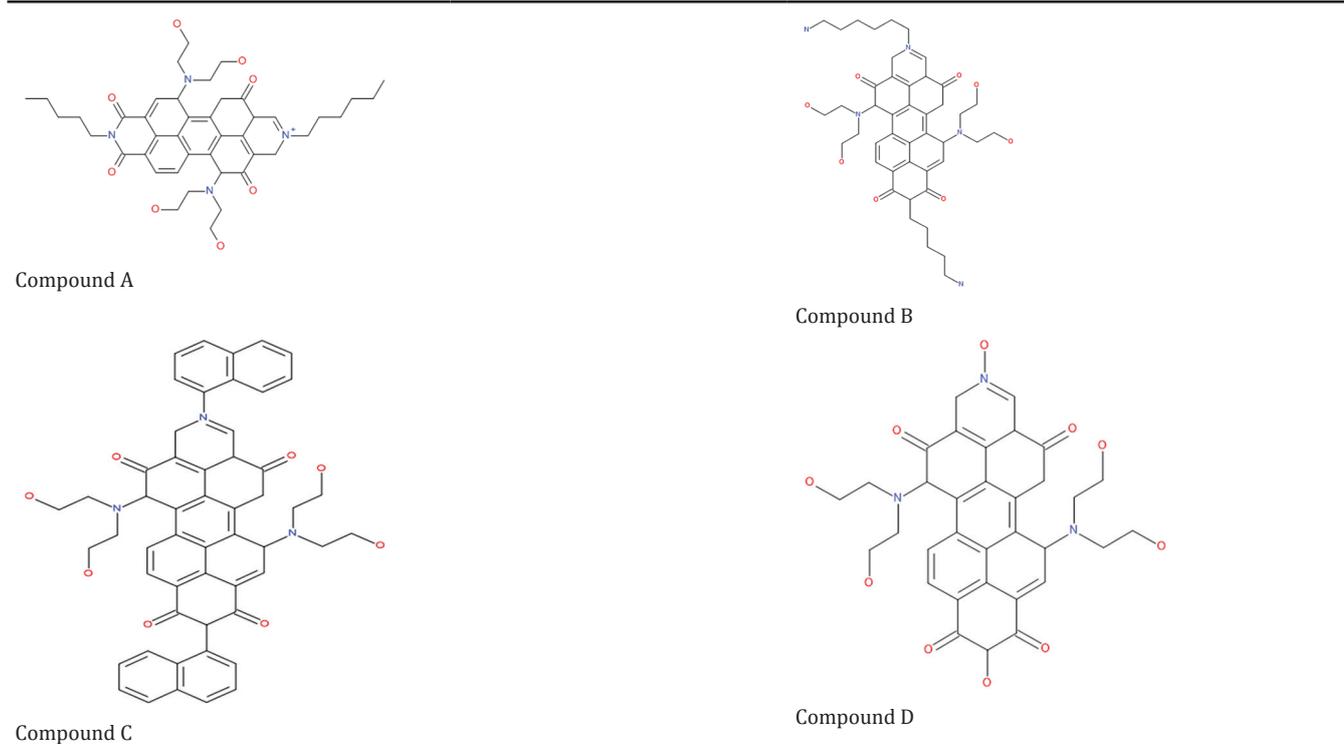


Table 2: DPPH free radical scavenging assay

Compound	Control	200	400	600	800	1000
A						
OD (%)	0.629 (0)	0.56 (10.97)	0.554 (11.92)	0.546 (13.2)	0.533 (15.26)	0.522 (17.01)
B						
OD (%)	0.629 (0)	0.56 (29.09)	0.554 (38.79)	0.546 (39.9)	0.533 (42.77)	0.522 (44.2)
C						
OD (%)	0.629 (0)	0.56 (0.16)	0.554 (8.43)	0.546 (15.58)	0.533 (21.14)	0.522 (24.96)
D						
OD (%)	0.629 (0)	0.56 (16.22)	0.554 (16.69)	0.546 (19.55)	0.533 (20.67)	0.522 (28.93)
Std-Ascorbic Acid						
OD (%)	0.169 (0)	0.561 (76.85)	0.582 (79.73)	0.626 (85.75)	0.658 (90.14)	0.711 (97.40)

DPPH: 1,1-Diphenyl-2-picryl hydrazyl

Table 3: Nitric oxide scavenging assay

Compound	Control	200	400	600	800	1000
A						
OD (%)	0.390 (0)	0.282 (27.69)	0.26 (33.33)	0.251 (35.64)	0.242 (37.95)	0.225 (42.31)
B						
OD (%)	0.390 (0.00)	0.34 (12.82)	0.292 (25.13)	0.28 (28.21)	0.266 (31.79)	0.245 (37.18)
C						
OD (%)	0.390 (0)	0.385 (1.28)	0.355 (8.97)	0.334 (14.36)	0.315 (19.23)	0.285 (26.92)
D						
OD (%)	0.390 (0)	0.389 (0.26)	0.382 (2.05)	0.371 (4.87)	0.361 (7.44)	0.35 (10.26)
Std-Quercetin						
OD (%)	0.584 (0)	0.464 (20.55)	0.324 (44.52)	0.236 (59.59)	0.157 (73.12)	0.082 (85.96)

perylene di-imides were used as a G-Quadruplex stabilizer, which can be stabilized and inactivate telomerase protection towards telomere, which promotes apoptosis. QSAR studies have been carried out using standard perylene compounds targeting telomerase by QSAR PLUS Module- Vlife Software. Around 491 compounds have been drawn using accelrys draw and studies. From the results, 59 compounds are selected for docking using G4LDB, patch Dock Server, and AutoDock by using selective G-Quadruplex targets (1NZM, 3CE5, 1L1H and 4B18).

9 compounds are selected from the three docking results based upon the binding free energy, *pki*, complementary shape analysis. These 9 compounds are compared with the standard drug PIPER and the interactions are visualized by Discovery studio Visualizer 4.1. Among the 9 compounds, 2 compounds (Compound 11 and 20) possess best interactions with the receptor and may have potent inhibitory effect against the telomerase enzyme for anti-cancer activity. From the 2 compounds Compound, 11 was chosen as a scaffold for synthesis. The

synthesis has been designed as bromination of Perylene Di imides at position 1st and 7th position and then condensed with amines, finally alkylation with Diethanolamine to remove the bromine as HBr. We have synthesized four PDI prototypes and characterized by FT-IR, ¹HNMR, ESI-Mass and also purified by column chromatography with

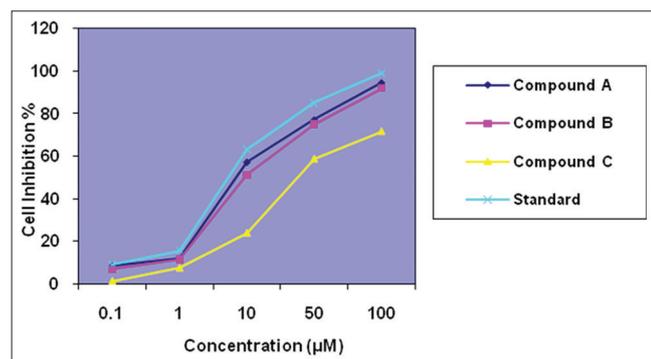
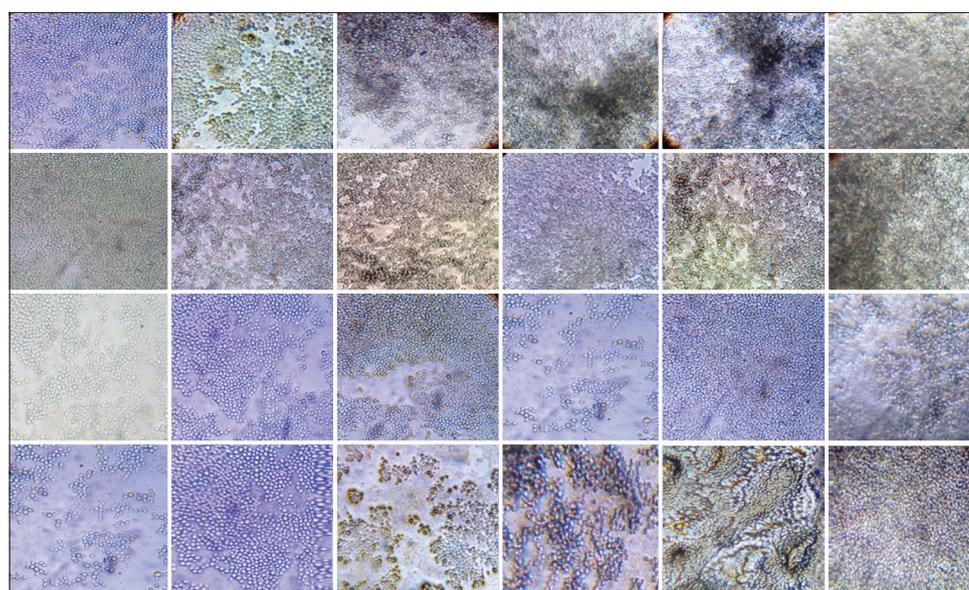


Fig. 1: Consolidated table of concentration of drugs versus cell inhibition % - HCT-116 colon cancer cells

a combination of solvents. We have taken a colon cancer cell line i.e. HCT-116 to screen our three ligands (Compound A, B and C) by MTT assay for understanding the cell cytotoxicity towards the cancer cells. Ligands were chosen based on the elaborate molecular modeling study to prove our hypothesis. The molecular modeling data's are exactly correlated with the *in vitro* studies. We have used 5-Fluorouracil and PIPER as a standard for *in vitro* study and molecular modeling study respectively. *In vitro* study revealed that Compound A and Compound B were remarkably able to kill the cancer cells as per the hypothesis being made earlier. Compound A and Compound B showed significant activity with respect to 5-Fluorouracil. Molecular modeling study helps to open future de novo modeling of a new compound to treat cancer. Other compounds were failed to show promising cytotoxicity towards cell line. The compounds will be exposed to other human cancer cell lines to understand the molecular mechanism of the ligands for the future prototypes.

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Figs. 2: Compound A, B, C and standard compound (5-fluorouracil) treated with HCT-116 cell line in different concentrations (i) 0.1 µM (ii) 1 µM (iii) 10 µM (iv) 50 µM (v) 100 µM (vi) Control

Table 4: Consolidated table of concentration of drugs versus cell viability % - normal Vero cells

S. No	Concentration (µM)	Compound A	Compound B	Compound C	Standard Drug
1	0.1	95.13	98.63	96.86	98.11
2	1	90.09	94.87	89.90	94.82
3	10	88.61	91.56	88.22	87.34
4	50	83.46	84.95	82.15	82.30
5	100	80.50	82.21	78.68	78.21

Table 5: Consolidated table of concentration of drugs versus cell inhibition % - HCT-116 colon cancer cells

S. No	Concentration (µM)	Compound A	Compound B	Compound C	Standard Drug
1	0.1	8.50	6.95	1.30	9.05
2	1	11.94	11.58	7.53	15.43
3	10	57.29	51.16	23.90	63.17
4	50	77.13	74.95	58.78	84.98
5	100	94.53	91.79	71.69	98.97
6.	IC50	32.37 µM	35.67 µM	57.46 µM	26.93 µM

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AUTHORS CONTRIBUTION

The authors are equally contributed to the research work and preparing the manuscript.

CONFLICTS OF INTERESTS

The authors declare no conflict of interest.

REFERENCES

- Schwab M. Encyclopedia of Cancer. 2nd ed. New York: Springer; 2008. p. 310-2.
- Antonio MD, Rodriguez R, Balasubramanian S. Experimental approaches to identify cellular G-Quadruplex structures and functions. *Methods* 2012;57:84-92.
- Sen D, Gilbert W. Formation of parallel four-stranded complexes by guanine-rich motifs in DNA and its implications for meiosis. *Nature* 1988;334:364-6.
- Davis JT. G-quartets 40 years later: From 5'-GMP to molecular biology and supramolecular chemistry. *Chem. Int Ed Engl* 2004;43:668-98.
- Brya TM, Baumann P. G-Quadruplexes: From guanine gels to chemotherapeutics. *Mol Biotechnol* 2011;49:198-208.
- Patel DJ, Phan AT, Kuryavyi V. Human telomere, oncogenic promoter and 5'-UTRG-quadruplexes: Diverse higher order DNA and RNA targets for cancer therapeutics. *Nucleic Acids Res* 2007;35:7429-55.
- Grand CL, Han H, Muñoz RM, Weitman S, Von Hoff DD, Hurley LH, et al. The cationic porphyrin ttmpy4 down-regulates c-MYC and human telomerase reverse transcriptase expression and inhibits tumor growth *in vivo*. *Mol Cancer* 2002;1:565-73.
- Lu YJ, Ou TM, Tan JH, Hou JQ, Shao WY, Peng D, et al. 5-N-methylated quindoline derivatives as telomeric g-quadruplex stabilizing ligands: Effects of 5-N positive charge on quadruplex binding affinity and cell proliferation. *J Med Chem* 2008;51:6381-92.
- Bejugam M, Sewitz S, Shirude PS, Rodriguez R, Shahid R, Balasubramanian S. Trisubstituted isoalloxazines as a new class of g-quadruplex binding ligands: Small molecule regulation of c-kit oncogene expression. *J Am Chem Soc* 2007;129:12926-7.
- Duan W, Rangan A, Vankayalapati H, Kim MY, Zeng Q, Sun D, et al. Design and synthesis of fluoroquinophenoxazines that interact with human telomeric g-quadruplexes and their biological effects. *Mol Cancer Ther* 2001;1:103-20.
- Hemalatha CN, Aanandhi MV. G-Quadruplex ligands as stabilizer targeting telomerase as anti-cancer agents. *Asian J Pharm Clin Res* 2017;10:50-3.
- Hemalatha CN, Aanandhi MV. 3D QSAR and docking study of perylene- di imides analogues as potent apoptosis inducer and efficacious anticancer agent. *Indian Drugs* 2017;54:15-27.
- Hemalatha CN, Aanandhi MV. Application of 3D QSAR and docking studies in optimization of perylene di imides as anti cancer agent. *Indian J Pharm Educ Res* 2018;52:46-54.
- Sissi C, Lucatello L, Krapcho AP, Maloney DJ, Boxer MB, Camarasa MV, et al. Tri-, tetra- and heptacyclic perylene analogues as new potential antineoplastic agents based on DNA telomerase inhibition. *Bioorg Med Chem* 2007;15:555-62.
- Taner K, Belgin SI, Yasemin GI, Funda Y. Evaluation of perylenediimide derivatives for potential therapeutic benefits on cancer chemotherapy. *Chem Biol Drug Des* 2012;80:675-81.
- Von Gadow A, Joubert E, Hansmann CF. Comparison of antioxidant activity of aspalathin with that of other plant phenols of Rooibos tea (*Aspalathon linearis*), α -tocopherol, BHT and BHA. *J Agric Food Chem* 1997;45:632-8.
- Yen GC, Duh PD. Scavenging effect of methanolic extracts of peanut hulls on free-radical and active oxygen species. *J Agric Food Chem* 1994;42:629-32.
- Marcocci I, Marguire JJ, Droy-Lefaiz MT, Packer L. The nitric oxide scavenging properties Ginkgo biloba extract. *Biochem Biophys Res Commun* 1994;201:748-55.
- Shahriar M, Hossain I, Sharmin FA, Akhter S, Haque A, Bhuiyan MA. *In vitro* antioxidant and free radical scavenging activity of *Withania somnifera* Root. *IOSR J Pharm* 2013;3:38-47.
- Abondanza TS, Oliveria CR, Barbosa CM. Bcl-2 expression and apoptosis induction in human HL-60 leukaemic cells treated with a novel organotellurium compound RT-04. *Food Chem Toxicol* 2008;46:2540-45.
- Mosmann T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J Immunol Methods* 1983;65:55-63.
- Lakshmi S, Dhanaya GS, Joy B, Padmaja G, Remani P. Inhibitory effect of an extract of *Curcuma zedoaria* on human cervical carcinoma cells. *Med Chem Res* 2008;17:335-44.
- Rossetti L, Franceschin M, Schirripa S, Bianco A, Ortaggi G, Savino M. Selective Interactions of Perylene derivatives having different side chains with inter- and intramolecular G-Quadruplex DNA Structures. A correlation with telomerase inhibition. Luigi Rossetti, Marco Franceschin. *Bioorg Med Chem Lett* 2005;15:413-20.
- Sarvesh H, Vimal K. *In vitro* cytotoxicity effect On MCF-7 cell line of co-encapsulated artesunate and curcumin liposome. *Int J Pharm Pharm Sci* 2017;9:123-8.
- Bashari MH, Hidayat S, Ruswandi YA, Putri T, Qomarilla N, Dwiwina RG. The N-Hexane fraction of *Myrmecodia* pendans inhibits cell survival and proliferation in colon cancer cell line. *Int J Pharm Pharm Sci* 2018;10:108-12.