

THE NEPHROTOXICITY OF CONCURRENT USE OF ENALAPRIL AND GENTAMICIN IN RATS

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ABSTRACT

Objective: The present study was aimed to assess the concurrent administration of Enalapril (ENAL) and Gentamicin (GM) in the kidney of rats.

Methods: Sixty male Sprague Dawley rats were divided into 4 main groups (n=15) according to the administered dose. Each main group was further subdivided into three subgroups according to the day of sacrificing (n=5). Group (C) was administered daily with normal saline as control, Group (E) was treated with oral ENAL, Group (G) was treated with 75 mg/kg GM, and Group (EG) was treated with GM and ENAL. The handling of the experiment persisted daily for 15 days, and the investigational examination carried out on days 5, 10, and 15.

Results: The result showed that GM nephrotoxicity augmented with the period of the experimental study, there was rising in the levels of serum creatinine and blood urea nitrogen on the 10th day and persisted in rising significantly during the period on the 15th day of the experiment. Administration of ENAL showed no significant alteration from those of controls. While the concurrent administration of ENAL and GM showed that ENAL gradually increased GM nephrotoxicity, these physiological retrogressions were accompanied with intensive renal histopathological deteriorations.

Conclusion: The present study has revealed that the concurrent administration of ENAL enormously aggravated the functional and histological nephrotoxicity of GM in rats.

Keywords: Nephrotoxicity, Concurrent, Enalapril, Gentamicin.

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INTRODUCTION

Nephrotoxicity can be defined as renal disease or dysfunction that arises as a direct or indirect result of exposure to medicines and industrial or environmental chemicals [1]. Gentamicin (GM) is an aminoglycoside antibiotic derived from a group of common soil microorganisms that are used in the treatment of severe infections [2]. Prolonged therapies with high dosages of GM evolve signs and symptoms of nephrotoxicity [3]. The GM nephrotoxicity display renal inflammatory process and oxidative stress in humans [4] and rats [5,6]. It has been established that GM nephrotoxicity is a dangerous circumstance distinguished by elevated serum creatinine and blood urea nitrogen (BUN) levels and enormous renal tubular necrosis, followed by worsening and renal failure [7].

Angiotensin-converting enzyme inhibitors (ACEI) are medications used primarily for the management of acute and chronic elevated blood pressure, left ventricular dysfunction and heart failure, and chronic kidney disease [8]. One study highlighted the importance of ACEI in the management of hypertension with proteinuria [9]. Enalapril (ENAL) is a long-acting ACEI, used in hypertension and heart failure, and has been shown to lower the death rate in systolic heart failure [10]; and preserve renal function through inhibiting the vasoconstrictive effects of angiotensin II at the efferent arteriole by reducing the concentration of angiotensin II or by blocking its receptor [11]. Experimental studies have shown that ENAL, improved systemic and renal hemodynamics and the intrarenal glomerular dynamics [12].

The effects of ACEI on GM nephrotoxicity are controversial. Captopril has been found to, potentiated GM-induced nephrotoxicity by increasing serum creatinine and urea concentrations and inducing severe necrosis of proximal convoluted tubule (PCT) [13]. However, captopril has been demonstrated to have a beneficial role in protecting GM-induced nephropathy [14,15]. The present study was designed to conclude whether ENAL would alter the pathophysiological changes induced in the kidney by GM.

METHODS

Animals and experimental design

The present study was carried out with the approval of the Institutional Animal Care and Use Committee of the International Islamic University of Malaysia (IACUC, IIUM). Animals of white, male, Sprague Dawley rats of 10–12 weeks old and body weight 250–300 g were used in this investigation. After acclimation to suitable laboratory conditions in animal facilities such as room temperature (24±4°C, 50% - relative air humidity, and 12:12 h light and dark cycles); had free access to laboratory standard pellet diet and water *ad libitum*. 60 rats were distributed into four main groups (n=15) according to the dosage of administered drug (Table 1). Each group was then subdivided into three subgroups according to the sacrificing day (n=5).

Chemicals

Gentamicin (GM) sulfate powder was purchased from (SANOFI) corporation. It was dissolved in distilled water and used as a single intraperitoneal dose of 75 mg/kg/day. ENAL maleate was purchased from Sigma and dissolved in distilled water; the rats treated daily an oral dose of 2 mg/kg using a syringe and metal ball-ended needle.

Biochemical analysis

Blood samples were collected by ventricular puncture, in plastic tubes, centrifuged, and the plasma was stored at -80°C until used for biochemical analysis. Serum creatinine and BUN analysis were performed according to the kinetic Jaffe method.

Histopathological study

Cervical dislocation was performed under light anesthesia at the end of the 5th, 10th, and 15th days of the experimental studies. The kidneys were rapidly excised, fixed in 10% formal saline for 72 h; dehydrated successively in 70%, 90%, 95%, and 100% ethanol, cleared in xylene, and embedded in paraffin wax. Paraffin sections of 4–5 µm thickness

Table 1: Distribution of animals by groups

Groups (15 rats/group)	Sub Groups (5 rats/subgroup)		
C: Normal saline (1 ml/day, i.p.)	C1: Killed on day 5	C2: Killed on day 10	C3: Killed on day 15
E: ENAL (2 mg/kg/day, i.p.)	E1: Killed on day 5	E2: Killed on day 10	E3: Killed on day 15
G: GM (75 mg/kg/day, i.p.)	G1: Killed on day 5	G2: Killed on day 10	G3: Killed on day 15
EG: ENAL (2 mg/kg/day, i.p.)+GM (75 mg/kg/day, i.p.)	EG1: Killed on day 5	EG2: Killed on day 10	EG3: Killed on day 15

ENAL: Enalapril

Table 2: Blood urea nitrogen (mmol/L) and serum creatinine (umol/L) in the studied groups

Groups	Blood urea nitrogen (mmol/L)			Serum creatinine (umol/L)		
	Day 5	Day 10	Day 15	Day 5	Day 10	Day 15
C	7.6±0.6	7.6±0.7	7.9±0.6	26±3	30.9±4.2	41.6±9
E	6.9±1.2	7.9±1.1	8.4±0.5	27.53±3.5	46.8±3.6	68.4±19.9
G	12.65±7.53	13.2±5.7*	16.06±6.7*	59.4±23.8*	92±8.8*	95.2±10.7*
EG	13.3±1.1	86.5±11.2**	97.5±17.5**	53±20.8	116±18.7**	754.2±184.5**

Values are expressed as mean±SEM. P values: *Significantly (p<0.05) different from control group, **Highly significantly (p<0.01) different from Gentamicin group. The statistical test employed was ANOVA, followed by a *post hoc* test (Tukey's HSD), n=5 in each group

were stained with Hematoxylin and Eosin. Evaluation of the kidney sections was performed blindly by two independent histopathologists.

Statistical analysis

Statistical analysis was performed using the SPSS version 20. The values were represented as mean ± SEM. A One-way Analysis of Variance (ANOVA) test was used and followed by a *post hoc* test (Tukey's HSD); significance was set to p<0.05 being considered as significant and p<0.05 as highly significant.

RESULTS

All experimental rats were vital and in good health; no mortality was noted throughout the duration of the investigation; nevertheless, the EG group rats showed irritating behavior.

Biochemical analysis

The current research revealed that the GM treated rats, and GM together with ENAL for (5, 10, and 15) days lead to a significant (p<0.05) elevation in the concentrations of the serum creatinine and BUN when compared to the control rats. ENAL administration to the rats did not induce any remarkable alteration in the quantities of serum creatinine and BUN. Rats injected with GM produced a non-significant (p>0.05) elevation in BUN (12.65±7.53 mmol/L) and a significant (p<0.05) rise in serum creatinine level (59.4±23.8 umol/L) on the 5th day. A significant elevation (p<0.05) in BUN (13.2±5.7 mmol/L) and serum creatinine (92±8.8 umol/L) compared to control were observed on the 10th day. On the 15th day of treatment, more deterioration of renal function was observed; there was a significant increase in BUN (16.06±6.7 mmol/L) as well as a significant increase in serum creatinine (95.2±10.7 umol/L).

The concomitant administration of GM with ENAL for 5 days showed a non-significant (p>0.05) increase in BUN (13.3±1.1 mmol/L) and serum creatinine level (53±20.8 umol/L) compared to GM treated group. Treatment with both drugs for 10 days induced a high significant (p<0.01) increase in BUN (86.5±11.2 mmol/L) and serum creatinine level (116±18.7 umol/L); further treatment for 15th day also showed a high significant (p<0.01) increase in BUN (97.5±17.5 mmol/L) and serum creatinine level (754.2±184.5 umol/L) compared to GM treated group (Table 2).

Histological observation

Analysis of H and E stained sections of kidneys from the control group (C group) showed normal histological architectures of the renal corpuscles, and renal tubules (Fig. 1a and b). Assessment of the kidneys sections of ENAL group (E group) demonstrated preservation of the renal cortex and medulla without any apparent histological alterations in the glomeruli and the renal tubules during the whole period of the treatment (Fig. 1c and d).

GM treatment (G group) gave rise to noticeable histopathological changes in the kidney tissues; the severity of these changes was increased with the increased period of GM exposure. On the 5th day of the investigation (G1 group), the rats exhibited no apparent histopathological alteration.

The earliest histological deterioration changes in the cortical tissue were observed on the 10th day (G2 group); these changes consist of shrinkage of the glomeruli, expansion of the Bowman's space; and an increase of the mesangial cells. In addition, dilation of numerous PCT and distal convoluted tubule, many cells of the PCT became visible with vacuolated cytoplasm and others had pyknotic nuclei; furthermore, many congested blood vessels were observed (Fig. 2a).

The histological changes in the kidneys of G3 group rats were more severe and extended to the renal papillae after 15 days of treatment. The cortex exhibited further shrinkage of the glomeruli and unorganized expansion of the Bowman's space beside to mesangial cell proliferation, and congestion of the capillaries. Furthermore, numerous PCT was injured showing tubular dilatation with vacuolar and cloudy epithelial cells lining, many PCT appeared particularly deprived of their lining epithelial cells, with only a bare basement membrane still existing; on the other hand, some PCT cells appeared disrupted with vacuolated cytoplasm and damaged brush borders. Shed epithelial cells and their fragments were occasionally detected. Remarkable blood vessels congestion, interstitial edema, and mononuclear cell infiltration were observed surrounding blood vessels and renal corpuscles (Fig. 2b and c). This histological deterioration proceeded and extended into the renal papillae; the tubules exhibited worsening histopathological injuries, accompanied by the incidence of congested blood vessels and hyaline casts (Fig. 2d).

The concomitant administration of ENAL and GM in (EG groups) revealed several histopathological changes in renal corpuscles and the urinary tubules in comparison with GM administered rats; the severity of these changes augmented with the increased duration of drug exposure. After 5 days of treatment (EG1 group); there was a progressive pathological deterioration in the renal tissue in compression with those observed in GM treated group. The renal corpuscles showed shrinking of the glomeruli and expansion of the Bowman's space and increased mesangial cells; the epithelial cells of PCT were necrotic, and cellular debris was observed in the tubular lumen correlated with mononuclear cells infiltration and congested blood vessels (Fig. 3a and b).

After 10 days of treatment, the kidneys of (EG2) group showed intense necrotic epithelial cells of PCT with hydropic alterations; the lumen of the expanded renal tubules was filled with desquamated epithelial cells and hyaline casts. The injured renal tubules were associated

with distinguishable mononuclear cells infiltration and congested blood vessels (Fig. 4a and b). The damage was extended into the renal papilla; showing dilated tubules, some of them were filled with hyaline casts; these changes were associated with congested blood vessels (Fig. 4c).

With the advancement of concomitant treatment with GM and ENAL for 15 days (EG3 group); the renal parenchyma exhibited intensive retrogradation in renal corpuscles and tubular epithelium than those observed on the 10th day (EG2 group). Shrunken glomerular tufts with distinguished hemorrhagic lesions concomitant with numerous mesangial cell were detected (Fig. 5a and b).

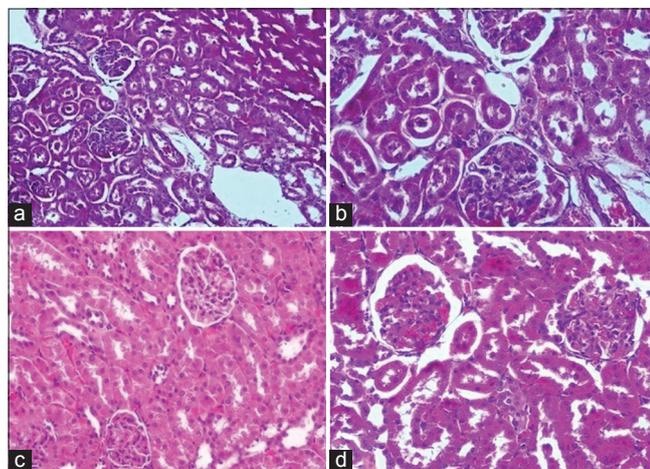


Fig. 1: (a and b) Control rat is showing normal histological structure of the glomerulus and architecture of tubules in the cortical portion. (c and d) Enalapril-treated rat kidney for 10 and 15 days, respectively, showing no pathological changes. H and E. X "A&C 200," "B and D 400"

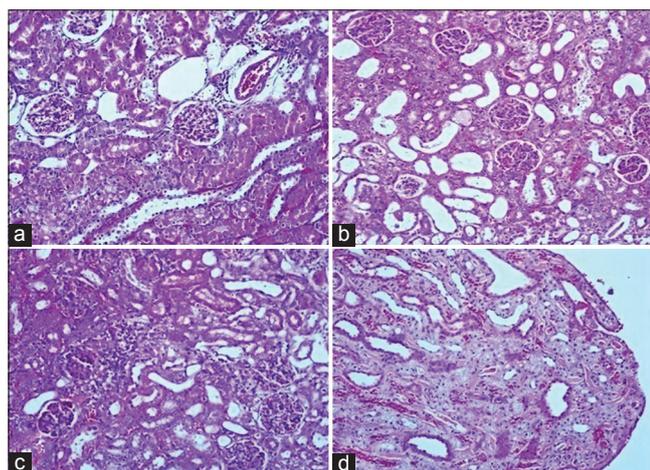


Fig. 2: (a) Gentamicin (GM) treated rat kidney for 10 days (G2) showing renal corpuscles with shrinkage of the glomeruli and widening of the Bowman's space, many PCT with the necrotic epithelial cell, presence congested blood vessels. (b and c) GM treated rat kidney for 15 days (G3) showing some renal corpuscles with shrinkage of the glomeruli, widening of the Bowman's space, capillary congestion and increased mesangial cells, many PCT with tubular dilatation and devoid of epithelial cells and mononuclear inflammatory cell infiltrates, and presence congested blood vessels. Furthermore, GM treated rat kidney for 15 days showing renal papilla with congested blood vessels, Tubular dilatation, hyaline casts (d). H and E. ×200

The lining epithelial cells of the renal tubules revealed extensive histopathological changes characterized swelled and necrotic cells, destructed brush border of dilated PCT; congested capillaries and focal hemorrhage were observed between the deteriorated renal tubules. The tubular lumens were filled with desquamated cells, cell debris and hyaline eosinophilic material, in addition, mononuclear cellular infiltration was observed in the interstitial tissues (Fig. 5a and b). Considerable renal papillary necrosis with congested blood vessels; in addition, hyaline casts, granular eosinophilic debris, and desquamated necrotic tubular epithelial cells accompanied with interstitial mononuclear cell infiltration were found in the rats of this group (Fig. 5c).

DISCUSSION

Aminoglycosides mainly GM has long been known as a significant contributor to a kidney problem, it is the most distinctive sources of drug-induced nephrotoxicity [16]. Previous experimental researches attempted to minimize the toxic impact of GM on kidney by combining it with several medicines and remedies; Vitamin E and N-Acetyl Cysteine significantly restored renal functions in rats treated rats with 80 mg/kg/8 days of GM [17]. Histopathological observation showed that Piperacillin has protective effects against the nephrotoxicity of GM in rabbits treated daily with 100 mg/kg GM for 5 days [18]. Moreover,

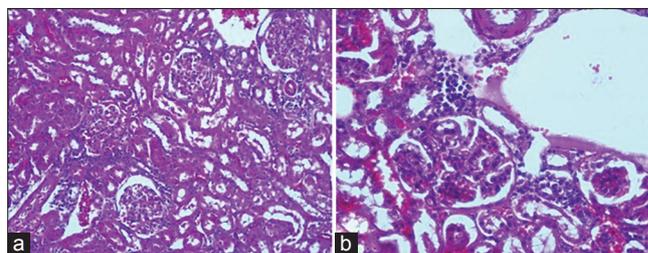


Fig. 3: (a and b) Gentamicin and Enalapril treated rat kidney (GE1) for 5 days showing renal corpuscles with shrinkage of the glomeruli, increased mesangial cells and widening of the Bowman's space, many PCT with necrotic epithelial cell and cast in the tubules, mononuclear infiltration, presence congested blood vessels. H and E. X "A 200," "B 400"

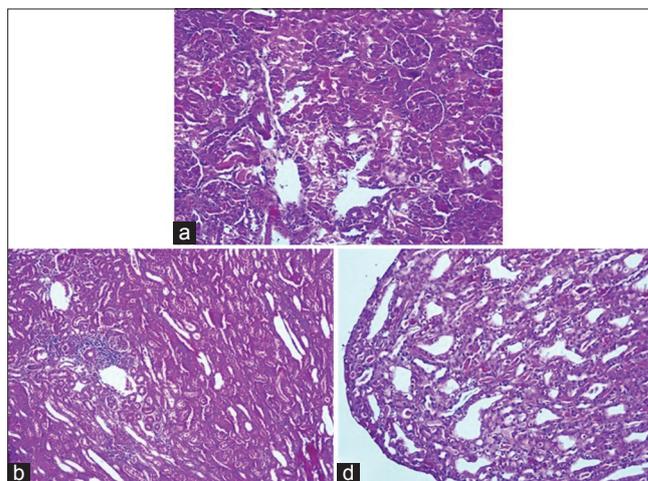


Fig. 4: (a and b) Gentamicin and Enalapril treated rat kidney (GE2) for 10 days showing renal corpuscles with more mesangial cells many PCT with necrotic epithelial cell and hydronic changes and cast in the tubules, mononuclear infiltration presence congested blood vessels. (c) Renal papilla of (GE2) with congested blood vessels, tubular dilatation, and hyaline casts. H and E. ×200

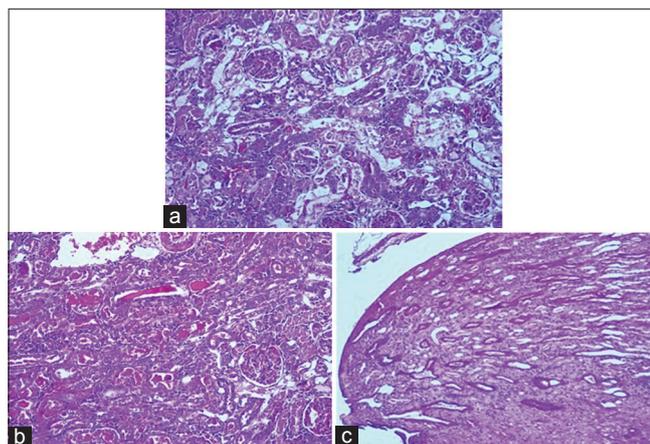


Fig. 5: (a and b) Gentamicin and Enalapril treated rat kidney (G3) for 10 days showing renal corpuscles with shrunken glomeruli, increased mesangial cells and congested capillaries, severe necrotic PCT and hydropic change cast in the tubules. (c) Necrotic renal papilla of (G3) with congested blood vessels, tubular dilatation, hyaline casts and mononuclear infiltration. H and E. $\times 200$

Melatonin also reduced GM-induced nephrotoxicity in rats [19]. In contrast, other studies reported that the simultaneous administration of furosemide and GM might intensify GM-induced nephrotoxicity by reducing the volume of distribution of GM and increase the renal tubular absorption of GM and accelerate its accumulation in renal tissues [20]. The present investigation demonstrated that treatment with GM-induced physiological deterioration and morphological renal lesions; there was a gradual increase of BUN and creatinine levels starting from the 5th day toward the end of the experimentation on the 15th day; this rising was commensurate with the duration of treatment; it is concordant with the increment of BUN demonstrated in rats administered with GM and amikacin [21]. Treatment of rats with 50 mg/kg/day of GM for 10 days [22], and for 15 days [4], indices significantly higher BUN and serum creatinine concentrations associated with renal injury in the glomeruli and tubular epithelial cells necrosis scores that proportionally increased with increasing doses of the antibiotic. In addition, short-term treatment with GM for 3 days or less has a very little prevalence of nephrotoxicity [23]. Conversely, longer duration of GM treatment "more than 2-week" is associated with worsening of renal function and resulted in a significantly greater decrease in glomerular filtration rate [24]. A gradual deterioration in kidney function was also observed in rats treated with 40 mg/kg GM [25]. However, other studies have demonstrated that rats treated with 70 mg/kg GM for 11 days did not reveal an increase in the level of the BUN, despite they develop a decline in creatinine clearance [26]. These contradicting outcomes outputs may be ascribed to the dosage of GM, species, and gender of the used experimental animals and to their usability toward nephrotoxicity through the inclination of GM toward the kidneys and to its accessibility to accumulate GM in the kidney cortical tissues.

In the present study, the gradual increase of BUN and creatinine levels throughout the period of this experimental study "5th to the 15th day" was associated with abnormal morphology of the renal tissue in the rats treated with GM. The incidence of the degenerative changes in the renal cortex was observed on the 10th day of the treatment and continued throughout the rest period of the experiment progressing to the renal papillae on 15th day; this observation is consistent with previous investigations that observed severe degenerative changes in renal glomeruli and necrosis of the PCT on the 10th day and increased throughout the whole period of the study [27]. The mechanisms and consequences of nephrotoxicity induced by GM have not been established so far; the generation of reactive oxygen species, causing a deficiency in intrinsic antioxidant enzymes was suggested as a

mechanism in the development of GM nephrotoxicity [28]. Induction of lipid peroxidation and mitochondrial hydrogen peroxide production were observed in the cortical renal tissue of rats treated with GM that leads to cellular degradation by denaturation of protein, peroxidation of lipids, and DNA destruction [29,30].

The present investigation has shown that ENAL gradually augmented the retrogression of renal physiological activities stimulated by GM. The intense degenerative histological changes observed on the 5th day and became severe and increased with the duration of drug exposure and extended to the renal papillae. There are controversial results regarding the outcome of the concomitant use of ACEI with GM; previous investigations using ACEI with GM indicated that ACEI might reduce or rise GM nephrotoxicity. Improvement in the functional deteriorations and renal damage induced by GM was observed after the administration of captopril that leads to suppression of progression of GM nephrotoxicity; this amelioration was related to the captopril potentiality to augmented bradykinin deliberation that leads to "vasodilator" which motivates prostaglandin production, ensuring an enlargement of the intrarenal blood vessels, and leading to an amelioration in the glomerular hemodynamic and preservation of the kidney from nephrotoxicity [15, 31]. On the other hand, other studies have shown that ACEI increase GM nephrotoxicity, the concurrent treatment with perindopril and GM-induced a greater renal impairment than after the administration of GM alone [32], captopril has been found to increase GM nephrotoxicity and progressively decreased the creatinine clearance in rats [33], captopril also aggravated the detrimental consequence of GM on the renal function in rats depleted of potassium [13]. Captopril was also accompanied by a reduction of creatinine clearance and a decrease in excretion of cyclic GMP in hypertensive rats [34]. Furthermore, Patzer [35] detected that throughout ACEI commencement, renal dysfunction could take place due to a decline in renal perfusion pressure and subsequent reduction in glomerular filtration; this is due to vasodilation of the renal efferent arteriole, which minimizes the kidney's capacity to recompense for low perfusion conditions. Further structural, metabolic, functional, and clinical studies are needed to elucidate the mechanisms of the elevation effect of ENAL on the manifestations of GM nephrotoxicity.

CONCLUSION

The present study has revealed that the simultaneous administration of ENAL enormously aggravated the functional and histological nephrotoxicity of GM in rats.

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CONFLICTS OF INTEREST

The authors have no conflicts of interest to declare.

AUTHOR'S CONTRIBUTION

The authors declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

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