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SIMULTANEOUS QUANTIFICATION OF PENTAZOCINE AND NALOXONE BY STABILITY INDICATING REVERSE-PHASE-HIGH-PERFORMANCE LIQUID CHROMATOGRAPHIC METHOD

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ABSTRACT

Objective: The objective of the study was to develope a stability indicating high-performance liquid chromatographic (HPLC) method for simultaneous assay of pentazocine and naloxone in bulk and tablets.

Methods: Pentazocine and naloxone were analyzed on Dionex C18 column using $0.1M \text{ K}_2\text{HPO}_4$ buffer (pH 4.0) and methanol (60:40, v/v) as the mobile phase. The concentration of pentazocine and naloxone was quantified by photodiode array detector set at 248 nm. The method was validated in compliance with ICH rules. Pentazocine and naloxone tablet formulation was subjected to forced degradation such as acid, neutral and alkali hydrolysis, oxidation, photo, and thermal degradation.

Results: The method was linear, with R^2 =0.9999 in the concentration range 100–300 µg/ml for pentazocine and R^2 =0.9995 in the concentration range 1–3 µg/ml for naloxone. The level of detection and quantification was 0.097 µg/ml and 0.322 µg/ml for pentazocine and 0.0073 µg/ml and 0.0243 µg/ml for naloxone, respectively. The degraded products are resolved well from pentazocine and naloxone with significantly different retention time values. From validation results, it was proved that the method is selective, precise, robust, and accurate for the estimation of pentazocine and naloxone simultaneously.

Conclusion: The developed stability-indicating HPLC method can be applied for quantitative determination of pentazocine and naloxone in tablets.

Keywords: Pentazocine, Naloxone, Synthetic opioids, Degradation, Reverse-phase high-performance liquid chromatographic.

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INTRODUCTION

Pentazocine, chemically known as (2RS,6RS,11RS)-6, 11-dimethyl-3-(3-methylbut-2-en-1-yl)-1,2,3,4,5,6-hexahydro-2,6-methano-3benzazocin-8-ol (Fig. 1), is an synthetic opioid. It has agonist activity at ƙ and σ opiate receptors and antagonist activity at μ opiate receptor [1-3]. Pentazocine is used as a pain reliever for moderate to severe pain [3-5].

Naloxone, chemically known as (4R,4aS,7aR,12bS)-4a,9-dihydroxy-3-prop-2-enyl-2,4,5,6,7a,13-hexahydro-1H-4,12-methanobenzofuro [3,2-e]isoquinoline-7-one (Fig. 1), is a synthetic *N*-allyl oxymorphone derivative with opiate antagonist activity [6,7]. It has antagonist activity at &, μ , and σ opiate receptors. Naloxone is used in emergency cases to reverse respiratory depression due to overdoses of opioids such as morphine, heroin, and other opioids [8-11].

The fixed-dose combination of pentazocine and naloxone was approved by the FDA [12]. The combination of pentazocine and naloxone is used to reduce pain that is extreme requiring opioid therapy as it may not be tolerated or other pain medications have not worked well enough [13,14]. Pentazocine was quantified in pharmaceutical samples [15-17], human serum [18], human plasma [19-21], human urine [21,22], and whole human blood [22] using visible spectrophotometry [15,16], Thin-layer chromatography [17], high-performance liquid chromatographic (HPLC) [18-20], potentiometry [21], and gas chromatography [22]. Naloxone was quantified in microparticles [23], dosage forms [24], transdermal formulations [25], human plasma [26-28], human urine [28], and human liver microsomes [28] using spectrophotometry [23], HPLC [23-27], and liquid chromatography-mass spectrometry [28].

Untill date, no technique was published regarding the simultaneous estimation of pentazocine and naloxone. The present research therefore

focuses on development of reverse-phase HPLC (RP-HPLC) method followed by validation as per the ICH guidelines.

MATERIALS AND METHODS

Instrumentation

HPLC analysis of pentazocine and naloxone was performed using Waters HPLC Alliance system (Waters Corporation, USA) equipped with a controller, quaternary pump, degasser, auto sampler, photodiode detector, and column oven. The processing of data was done using empower 2 software.

Materials

Pentazocine (98% purity) and naloxone (98% purity) were procured from Rainbow Pharma Training Lab (Hyderabad, India). The organic solvent (methanol) and chemicals (K_2HPO_4 , NaOH, HCl, H_2O_2 , and H_2PO_4) were of HPLC and analytical grade, respectively. Pentazocine and naloxone combination tablets (Lupin Pharmaceuticals, Inc., Baltimore) labeled to contain 50 mg of pentazocine and 0.5 mg of naloxone was obtained from a local pharmacy store (Hyderabad, India).

Chromatography conditions

The column was a C18 Dionex column (250 mm × 4.6 mm, 5 µm particle size; Thermo Fisher Scientific, US). The column temperature was kept at ambient (25°C). The mobile phase consisted of K_2 HPO₄ buffer (0.1 M, pH 4.0 units, adjusted with H₂PO₄) and methanol (60:40, *v*/*v*). The flow rate and runtime were kept at 1.0 ml/min and 8 min, respectively. Detection wavelength of 248 nm was optimized. The mobile phase was used as a diluent in the preparation of solutions.

Standard solutions

Hundred mg of pentazocine and 1 mg of naloxone were correctly weighed with microbalance and dissolved in 100 ml of diluent in a

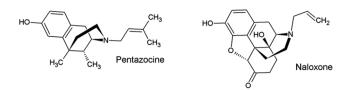


Fig. 1: Structures of drugs selected

100 ml standard flask to get a stock solution (1000 μ g/ml of pentazocine and 10 μ g/ml of naloxone). This mixed stock solution was diluted aptly to get working solution with concentration 200 μ g/ml of pentazocine and 2.0 μ g/ml of naloxone with mobile phase.

Calibration plot

Calibration solutions with five different concentrations (pentazocine – 100, 150, 200, 250, and 300 μ g/ml; and naloxone – 1.0, 1.5, 2.0, 2.5, and 3.0 μ g/ml) were prepared from mixed stock solution by apt dilution with diluent. These solutions were analyzed using the conditions given above (section – "chromatography conditions"). The calibration plot for pentazocine and naloxone was constructed separately by plotting the peak areas against the concentrations. The method's linearity was evaluated by determining the regression coefficient (R²). Pentazocine and naloxone content in unknown samples were computed by referencing them to their respective calibration plots.

Tablet analysis

Pentazocine and naloxone combination tablets were grinded into powder. An appropriate weight of tablet powder equal to 200 mg of pentazocine and 2 mg of naloxone was taken, dissolved in 30 ml of mobile phase and sonicated for 20 min. The volume was then completed to 100 ml with mobile phase (2000 μ g/ml of pentazocine and 20 μ g/ml of naloxone). One ml of prepared solution was diluted with diluent to 10 ml (200 μ g/ml of pentazocine and 2 μ g/ml of naloxone) for analysis by the proposed method. Pentazocine and naloxone contents in the tablets were computed by referencing them to their respective calibration plots.

Stress-induced degradation

The stress conditions used to induce degradation are as follows [29]:

- Neutral hydrolysis
- Base hydrolysis
- Acid hydrolysis
- Oxidation
- Thermal
- Photolytic.

The degradation was performed by adding 10 ml of reagent to 10 ml of tablet solution (2000 µg/ml pentazocine and 20 µg/ml naloxone). Hydrochloric acid (0.1 N), sodium hydroxide (0.1 N), deionized water, and hydrogen peroxide (30%) were used as reagent for acid hydrolysis, base hydrolysis, neutral hydrolysis, and oxidation, respectively. The solutions were sonicated at 25±2°C for 30 min. The samples were transmitted to a volumetric flask (100 ml) and filled with diluent to 100 ml. Following degradation, samples are filtered with a membrane filter of 0.45 µm pore size. Thermal stress was accomplished by exposing tablet powder (200 mg pentazocine and 2 mg naloxone) to 105°C for 6 h. Photolytic stress was accomplished by exposing tablet powder (200 mg pentazocine and 2 mg naloxone) to sunlight for 24 h. The thermal and photo degraded tablet sample solution was prepared as described earlier (section - "tablet analysis") and filtered with a membrane filter of 0.45 µm pore size. The suggested HPLC method was then used to analyze each sample. The peak purity of pentazocine and naloxone in stressed samples was evaluated by the photodiode array detector.

RESULTS AND DISCUSSION

Method development

Different combinations of methanol and buffer (0.1% phosphoric acid buffer, 0.1M Na_2HPO_4 buffer, and 0.1M K_2HPO_4 buffer) with different

pH, as well as different flow rates, were tested. Different C18 stationary phases (Waters, Develosil, Sunniest, and Dionex) with dimension 250 mm×4.6 mm and 5 μ m particle size were tested. Best peak width, peak symmetry, resolution, and sensitivity were obtained with Dionex C18 column, mobile phase mixture with 0.1M K₂HPO₄ buffer (pH 4.0), and methanol in the ratio of 60:40 (*v*/*v*). The optimized flow rate was 1.0 ml/min and detection wavelength was 248 nm. The optimized conditions showed a rapid and good separation of pentazocine and naloxone with retention time 3.714 min–4.761 min, respectively (Fig. 2).

Method validation

Method validation was carried out as per the ICH regulations [30,31].

Selectivity

During selectivity check, diluent blank, placebo, and tablet sample solution were screened and compared with standard solution for interference at retention times of pentazocine and naloxone. Significant interference was not observed at retention times of pentazocine and naloxone in the chromatograms of diluent blank, placebo, and tablet sample solution (Fig. 3). Thus, selectivity was demonstrated.

System suitability

The parameters regarding system suitability were tested by analysis (n=5) of pentazocine and naloxone standard solution at a concentration of 200 μ g/ml and 2 μ g/ml, respectively. The results were found within the acceptance criteria in line with ICH directives (Table 1).

Linearity

Linearity of pentazocine and naloxone was achieved over the range of $100-300 \ \mu g/ml$ and $1-3 \ \mu g/ml$, respectively. Linear regression equation and regression coefficient were calculated and given below:

- Pentazocine: Peak area=76697 x-12761, regression coefficient, R^2 =0.9999
- Naloxone: Peak area=14621 *x*-246.8, regression coefficient, R²=0.9995.

The good linearity of the method for pentazocine and naloxone was demonstrated through regression coefficient values (>0.999).

Limit of detection (LOD) and limit of quantitation (LOQ)

LOD and LOQ were assessed based on the standard deviation (SD) of intercept and slope (m) of the calibration plot. The below equations were employed to compute the LOD and LOQ values.

LOD=SD/m×3.3

LOQ=SD/m×10

The LOD and LOQ were 0.097 μ g/ml and 0.322 μ g/ml for pentazocine and 0.0073 μ g/ml and 0.0243 μ g/ml for naloxone, respectively. Values lesser than 1 μ g/ml confirm that the developed method was sensitive adequately.

Precision

The precision was appraised by analyzing (n=6) pentazocine and naloxone standard solution at a concentration of 200 μ g/ml and 2 μ g/ml, respectively, on the same day [31,32]. The mean peak area along with the relative SD was determined (Table 2). The results were found to be within the acceptance criteria (percent relative standard deviation [RSD] value - <2.0%).

Accuracy

The accuracy was appraised by recovery study through standard addition method [32,33]. The percent recovery studies for pentazocine and naloxone were done by spiking three different amounts of pentazocine and naloxone standard (50, 100, and 150%) to the pre-analyzed tablet sample. The percent recovery of pentazocine and naloxone was determined in three replicates for each level (Tables 3 and 4). The results of percent recovery were found within the acceptance criteria (80–120%).

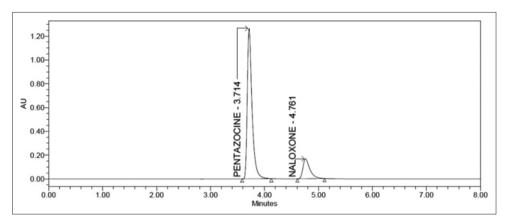


Fig. 2: Chromatogram of pentazocine and naloxone with optimized conditions

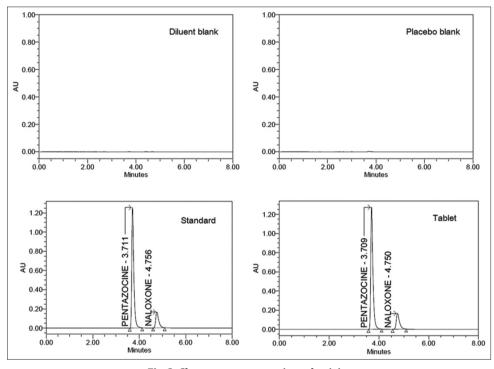


Fig. 3: Chromatograms proving selectivity

	Table 1: System	suitability data	for pentazocine	and naloxone
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Statistical parameter	Retention time	Area	Plate count	Tailing factor	Resolution
Pentazocine					
Mean*	3.712	7655800	9203	1.166	-
SD**	0.0011	37675.9570	211.7114	0.0089	-
RSD***	0.031	0.492	2.301	0.767	-
Criteria limits [30]	RSD-≤2%	RSD−≤2%	>2000	≤2.0	≤2.0
Naloxone					
Mean*	4.757	1466529	7063	5.246	1.268
SD**	0.0014	6299.2275	140.7434	0.0594	0.0083
RSD***	0.030	0.430	1.993	1.133	0.660
Criteria limits [30]	RSD-≤2%	RSD−≤2%	>2000	≤2.0	≤2.0

*Mean of five determinations, **standard deviation, ***relative standard deviation

Robustness

- Five variation parameters were studied to demonstrate robustness:
- Change in ratio of methanol by ±5.0%
- Change in pH of buffer by ±0.1 unit
- Change in flow rate by ±0.1 ml/min

- Change in detection wavelength by ±2 nm
- Change in column temperature by ±2.0°C.

The peak area, plate count, resolution, and tailing factor values of pentazocine and naloxone obtained from variation parameters were

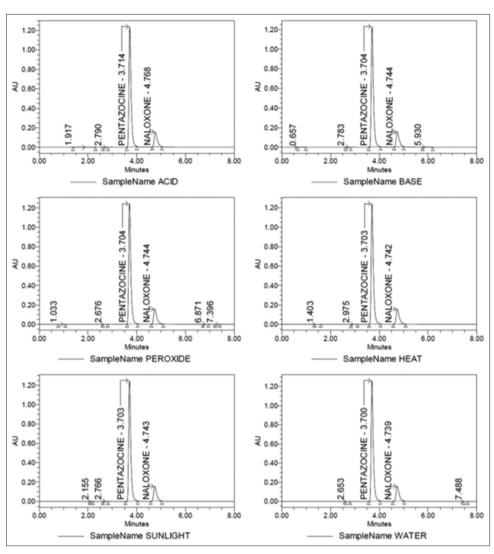


Fig. 4: Chromatograms proving stability indicating and specificity

Table 2: Precision data for pentazocine and naloxone

Inj. No.*	Peak area				
	Pentazocine	Naloxone			
1	7641945	1464317			
2	7655208	1451732			
3	7644772	1462000			
4	7646482	1463570			
5	7641857	1451692			
6	7653061	1450658			
Mean**	7647221	1457328			
SD***	5675.3883	6590.9768			
RSD****	0.074	0.452			

*Injection number, **mean of five determinations, ***standard deviation, ****relative standard deviation

compared with the acceptance limits. The results were within the acceptance criteria (Table 5).

Degradation study

Degradation investigation outcomes are shown in Table 6. Degradation of pentazocine and naloxone was seen in the stress conditions applied. Highest degradation of pentazocine and naloxone was observed in dry heat and acid conditions applied. Lowest degradation of pentazocine and naloxone was observed with neutral and peroxide conditions applied. The chromatograms of pentazocine and naloxone after degradation

Table 3: Recovery data for pentazocine

Level	Amount (µg/ml)	Recovery	Mean*	
spiked (%)	Spiked Determine		(%)	(%)	
50	99.000	99.31	100.31	100.28	
	99.000	99.20	100.20		
	99.000	99.32	100.32		
100	198.000	198.86	100.43	100.42	
	198.000	198.87	100.44		
	198.000	198.77	100.39		
150	297.000	296.99	100.00	100.12	
	297.000	298.62	100.54		
	297.000	296.44	99.81		

*Mean of three determinations

are presented in Fig. 4. The times of elution of pentazocine, naloxone, and degradants are different. These findings proved that the method is stability indicating and specific.

Tablet analysis

The method was applied to evaluate the content of pentazocine and naloxone in tablets. The recovered values were 100.237% for pentazocine and 101.013% for naloxone, indicating the method's reliability (Table 7). The RSD values were 0.150% for pentazocine and 0.178% for naloxone (Table 7), indicating the method's reproducibility.

Table 4: Recovery data for naloxone

Level spiked (%)	Amount (µg/ml)		Recovery	Mean* (%)
	Spiked	Determined	(%)	
50	0.980	0.99	101.15	101.15
	0.980	0.99	101.16	
	0.980	0.99	101.14	
100	1.960	1.98	101.14	101.08
	1.960	1.99	101.31	
	1.960	1.98	100.78	
150	2.940	2.96	100.75	100.81
	2.940	2.97	100.90	
	2.940	2.96	100.78	

*Mean of three determinations

Table 5: Robustness data for pentazocine and naloxone

Parameter	Retention	time	Plate count		Tailing factor		Resolution	
Drug	Pen	Nal	Pen	Nal	Pen	Nal	Pen	Nal
Flow 1	3.086	3.952	8275	6661	1.13	1.22	-	5.05
Flow 2	3.364	4.296	8891	7103	1.16	1.25	-	5.16
Temp 1	4.092	5.227	9227	7425	1.11	1.25	-	5.25
Temp 2	4.628	5.929	9482	7494	1.14	1.28	-	5.34
Ratio 1	3.086	3.952	8275	6661	1.13	1.22	-	5.05
Ratio 2	4.092	5.227	9227	7425	1.21	1.25	-	5.25
PH 1	3.712	4.755	9072	7006	1.18	1.27	-	5.20
PH 2	3.708	4.751	8951	6948	1.17	1.27	-	5.18
Nm 1	3.714	4.758	9161	7013	1.17	1.29	-	5.22
Nm 2	3.713	4.756	9557	7366	1.16	1.26	-	5.35

Pen: Pentazocine, Nal: Naloxone, Flow 1: 0.9 ml/min, Flow 2: 1.1 ml/min, Temp 1: 23°C, Temp 2: 27°C, Ratio 1: Methanol ration 45% by volume, Ratio 2: Methanol ratio 35% by volume, PH 1: 3.9 units, PH 2: 4.1 units, Nm 1: Wavelength 246 nm, Nm 2: Wavelength 250 nm

Table 6: Degradation data for pentazocine and naloxone

Deg. with	Pen area	Nal area	Pen % assay	Nal % assay	Pen % deg	Nal % deg
Acid	7243689	1334725	94.52	90.57	5.48	9.43
Base	7340455	1361014	95.79	92.35	4.21	7.65
Peroxide	7421869	1444737	96.85	98.04	3.15	1.96
Heat	7198634	1355646	93.93	91.99	6.07	8.01
Sunlight	7505340	1393436	97.94	94.55	2.06	5.45
Water	7530071	1438596	98.26	97.62	1.74	2.38

Pen: Pentazocine, Nal: Naloxone, Deg: Degradation

Table 7: Quantification of pentazocine and naloxone in tablets

Statistical parameter	Label claim (mg)	Determined (mg)	Recovered (%)
Pentazocine			
Mean*	50	50.137	100.273
SD**	-	0.0750	0.1501
RSD***	-	0.150	0.150
Naloxone			
Mean*	0.5	0.505	101.013
SD**	-	0.0008	0.1795
RSD***	-	0.178	0.178

*Mean of three determinations, **standard deviation, ***relative standard deviation

CONCLUSION

A simple and rapid RP-HPLC method was developed to quantify pentazocine and naloxone in bulk and tablets. The results of validation are satisfactory and adequate to quantify pentazocine and naloxone simultaneously. The results of forced degradation studies established the specificity and stability indicating nature of the method. The method offers adequate selectivity and accuracy for routine evaluation and quality control in laboratories for pentazocine and naloxone.

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AUTHORS' CONTRIBUTION

This work was done by RKK under the supervision of RS.

CONFLICTS OF INTEREST

The authors declared that they have no conflicts of interest.

REFERENCES

- 1. Gasparre G, Abate C, Carlucci R, Berardi F, Cassano G. The σ_1 receptor agonist (+)-pentazocine increases store-operated Ca⁺ entry in MCF7 σ_1 and SK-N-SH cell lines. Pharmacol Rep 2017;69:542-5.
- Craft RM, McNiel DM. Agonist/antagonist properties of nalbuphine, butorphanol and (-)-pentazocine in male vs. Female rats. Pharmacol Biochem Behav 2003;75:235-45.
- Hoskin PJ, Hanks GW. Opioid agonist-antagonist drugs in acute and chronic pain states. Drugs 1991;41:326-44.
- Wang N, Wang L, Gao Y, Zhou H, Wang J. Analgesic effect of preoperative pentazocine for laparoscopic cholecystectomy. Cureus 2016;8:e948.
- Xu HQ, Xing JM, Jia R. Clinical observation of pentazocine for postoperative intravenous analgesia in patients with lumbar herniation. Zhongguo Gu Shang 2010;23:838-40.
- Dhanalakshmi K. Opioid Analgesics. In: Pain Management. Vol. 2., Ch. 113. Netherlands: Elsevier Publications; 2007. p. 939-64.
- Chimbar L, Moleta Y. Naloxone effectiveness: A systematic review. J Addict Nurs 2018;29:167-71.
- van Dorp E, Yassen A, Dahan A. Naloxone treatment in opioid addiction: The risks and benefits. Expert Opin Drug Saf 2007;6:125-32.

- Lynn RR, Galinkin JL. Naloxone dosage for opioid reversal: Current evidence and clinical implications. Ther Adv Drug Saf 2018;9:63-88.
- Connors NJ, Nelson LS. The evolution of recommended naloxone dosing for opioid overdose by medical specialty. J Med Toxicol 2016;12:276-81.
- Elzey MJ, Fudin J, Edwards ES. Take-home naloxone treatment for opioid emergencies: A comparison of routes of administration and associated delivery systems. Expert Opin Drug Deliv 2017;14:1045-58.
- FDA Approved Drug Products. Available from: https://www. accessdata.fda.gov/scripts/cder/daf/index.cfm?event=overview. process&ApplNo=074736. [Last accessed on 2019 Jun].
- Pentazocine and Naloxone (Oral Route). Drugs and Supplements, Mayoclinic. Available from: https://www.mayoclinic.org/drugssupplements/pentazocine-and-naloxone-oral-route/description/drg-20074147. [Last accessed on 2019 Jun].
- Pentazocine and Naloxone Pentazocine Hydrochloride and Naloxone Hydrochloride Tablet. Dailymed. Available from: https://dailymed. nlm.nih.gov/dailymed/drugInfo.cfm?setid=a28450a0-ac93-4235-b9a6-58cdf24773cb. [Last accessed on 2019 Jun].
- Sastry CS, Rekha TV, Satyanarayana A. Spectrophotometric determination of pentazocine in pharmaceutical formulations. Indian J Pharma Sci 1998;60:55-8.
- Revanasiddappa HD, Veena MA. A sensitive spectrophotometric determination of ritodrine, pentazocine, isoxsuprine hydrochlorides and amoxicillin in pure and pharmaceutical samples. E-J Chem 2008;5:100-6.
- Poprzen V, Radulović D. UV-densitometry determination of pentazocine hydrochloride in tablets after low temperature extraction. Vojnosanit Pregl 2001;58:267-71.
- Kelly JW, Stewart JT, Blanton CD. HPLC separation of pentazocine enantiomers in serum using an ovomucoid chiral stationary phase. Biomed Chromatogr 1994;8:255-7.
- Hiedaki M, Kengo O, Ken-Ich H, Makoto S, Kazuhiro I, Hideo N, et al. Quantification of pentazocine in human plasma by HPLC with electrochemical detection. J Liquid Chromatogr 1992;15:3247-60.
- Moeller N, Dietzel K, Nuernberg B, Geisslinger G, Brune K. Highperformance liquid chromatographic determination of pentazocine in plasma. J Chromatogr 1990;530:200-5.
- 21. Ali RA, Ali AE, Saraji M. Rapid determination of pentazocine in human plasma and urine by a potentiometric method. Anal Lett

2009;42:571-83.

- 22. Seno H, Kumazawa T, Ishii A, Matsushima H, Watanabe-Suzuki K, Suzuki O, *et al.* Determination of pentazocine in human whole blood and urine by gas chromatography/surface ionization organic mass spectrometry. J Mass Spectrom 2000;35:33-8.
- Gil-Alegre ME, Barone ML, Torres-Suárez AI. Extraction and determination by liquid chromatography and spectrophotometry of naloxone in microparticles for drug-addiction treatment. J Sep Sci 2005;28:2086-93.
- Mostafa ST, Mohamed EM, Mahmoud MA. Determination of naloxone hydrochloride in dosage form by high-performance liquid chromatography. J Liquid Chromatogr 1983;6:1491-97.
- Panchagnula R, Sharma P, Khandavilli S, Varma MV. RP-HPLC method and its validation for the determination of naloxone from a novel transdermal formulation. Farmaco 2004;59:839-42.
- Reid RW, Deakin A, Leehey DJ. Measurement of naloxone in plasma using high-performance liquid chromatography with electrochemical detection. J Chromatogr 1993;614:117-22.
- Franklin M, Odontiadis J. Determination of naloxone in human plasma by high-performance liquid chromatography with coulometric detection. J Chromatogr B Biomed Appl 1996;679:199-203.
- Fang WB, Chang Y, McCance-Katz EF, Moody DE. Determination of naloxone and nornaloxone (noroxymorphone) by high-performance liquid chromatography-electrospray ionization- tandem mass spectrometry. J Anal Toxicol 2009;33:409-17.
- 29. International Conference on Harmonization, Stability Testing of New Drug Substances and Products (Q1AR2). Geneva, Switzerland: Proceedings of the International Conference on Harmonization; 2003.
- International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use, ICH Harmonized Tripartite Guideline. Validation of Analytical Procedures: Text and Methodology Q2 (R1). Geneva, Switzerland: ICH; 2005.
- Sharma S, Goyal S, Chauhan K. A review on analytical method development and validation. Int J Appl Pharm 2018;10:8-15.
- Hemant KJ, Umakant SJ. Development and validation of RP-HPLC method for estimation of darunavir ethanolate in bulk and tablets. Int J Pharm Pharm Sci 2015;7:386-9.
- Babu GR, Rao AL, Rao JV. A rapid RP-HPLC method development and validation for the quantitative estimation ribavirin in tablets. Int J Pharm Pharm Sci 2015;7:60-3.