

DEVELOPMENT AND VALIDATION OF NOVEL REVERSE-PHASE HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY METHOD FOR SIMULTANEOUS ESTIMATION OF ANDROGRAPHOLIDE AND ALOE-EMODIN

RASHMI PATIL, UTTARA JAISWAR, VANDANA JAIN*

Department of Quality Assurance, Oriental College of Pharmacy, Sanpada, Navi Mumbai, Maharashtra, India.
Email: vandana.jain@ocp.edu.in

Received: 22 October 2019, Revised and Accepted: 19 December 2019

ABSTRACT

Objective: The study aims to develop and validate a novel reverse-phase high-performance liquid chromatographic method for simultaneous estimation of andrographolide and aloe-emodin in herbal formulation and validate as per the International Conference on Harmonization (ICH) guidelines.

Methods: The analysis was carried on a Shimadzu LC Prominence-*i* 2030 model with the Lab Solution software. The column used for separation was Prontosil C18 (250×4.6 mm, 5 μ), with a mobile phase consisting of acetonitrile and 0.05% orthophosphoric acid (45:55), at a flow rate of 1 ml/min, column temperature was maintained at 28°C and effluents were monitored at 225 nm. The injection volume was 10 μl.

Results: The retention time of andrographolide and aloe-emodin was found to be 4.57±0.2 min and 12.29±0.2 min, respectively. The markers were resolved using linear responses that were obtained in concentration ranges of 0.5–60 μg/ml with correlation coefficient (r^2) of 0.9992 and 0.999 for andrographolide and aloe-emodin, respectively. The precision results were found to be satisfactory, which indicates that the method is precise. The recovery values lie in the range of 98–120% indicating the accuracy of the method.

Conclusion: A novel, simple, accurate, precise, and robust reverse-phase high-performance liquid chromatography method was developed for the simultaneous estimation of andrographolide and aloe-emodin. The developed method can be used for analysis of the formulations containing these phytoconstituents.

Keywords: Andrographolide, Aloe-emodin, Marketed formulation, International Conference on Harmonization guidelines.

© 2020 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>) DOI: <http://dx.doi.org/10.22159/ajpcr.2020.v13i2.36158>

INTRODUCTION

Ayurvedic medicines are polyherbal formulations and every herb consists of an array of chemical constituents.

Plants are considered as a conventional source for a large number of phytochemicals [1]. Ayurvedic medicines are polyherbal formulations and each herb consists of various chemical constituents [2]. The quality assessment of herbal formulations is vital to justify their acceptability in the modern system of medicine. The production and primary processing of herbal substance influences the quality of the active pharmaceutical ingredient [3]. Standardization of polyherbal formulations and the quantitative determination of markers in any polyherbal formulation is very challenging [4]. Standardization of herbal products can be achieved if they are evaluated using sophisticated techniques such as ultraviolet (UV)-visible, infrared, thin-layer chromatography, high-performance liquid chromatography (HPLC), high-performance thin-layer chromatography, gas chromatography-mass spectrometry, liquid chromatography-mass spectrometer, atomic absorption spectrometry spectrofluorimetric, and other such methods [5]. The literature survey reveals that various methods were developed for estimation of andrographolide and aloe-emodin alone or in combination with other markers [6-12], but no such HPLC analysis method for simultaneous estimation of andrographolide and aloe-emodin is reported.

This paper presents the development of a novel reverse-phase HPLC (RP-HPLC) method for the simultaneous estimation of andrographolide and aloe-emodin. The developed and validated method was applied for the standardization of marketed formulation for these two

markers. The selected formulation is a well-known formulation and is indicated against various complications such as jaundice, liver disorder, indigestion, hepatotoxicity, liver enlargement, gastroenteritis, fungal infections, gastritis, and other conditions [13]. This tablet consists of various medicinal plants namely, Bhringaraj (*Eclipta alba*), Revandchini (*Rheum emodi*), Sarapunkha (*Tephrosia purpurea*), Kalmegh (*Andrographis paniculata*), Kasni (*Cichorium intybus*), Giloy (*Tinospora cordifolia*), Hareetaki (*Terminalia chebula*), and Bhumymlaki (*Phyllanthus niruri*). Andrographolide from *A. paniculata* is reported to possess abortifacient, anti-inflammatory, antibacterial, antipyretic, antithrombotic, antiviral, antineoplastic, cardioprotective, choleric, digestive, expectorant, hepatoprotective, hypoglycemic, immune enhancement, laxative, and sedative activity [14]. Andrographolide is effective against liver damage caused by paracetamol or galactosamine. It also played a hepatoprotective role by reducing a lipid peroxidation product malondialdehyde [15]. aloe-emodin from *R. emodi* possess antioxidant, antimicrobial, antifungal, anticancer, antiulcer, antidyslipidemic, hepatoprotective, wound healing, and immune-enhancing activity [16]. Aloe-emodin also possesses multiple antiproliferative and anticarcinogenic properties in a host of cancer cell lines [17].

METHODS

Instrument

RP-HPLC Shimadzu LC Prominence-*i* 2030 model consisting of UV detector and autosampler was employed for the method development and validation. Software used was Lab Solution. UV-visible spectrophotometer was used for obtaining maximum wavelength (λ_{max}) of the compounds of interest.

Standards and reagents

Andrographolide and aloe-emodin standards were obtained from Yucca Enterprises, Mumbai, Maharashtra, India. Marketed formulation of Livfit Tablet of Alembic Pharmaceuticals Ltd. was procured from the local market of Mumbai, Maharashtra, India. All the chemicals used were of HPLC grade, which were procured from Thermo Fisher Scientific, India Pvt. Ltd., Powai, Mumbai.

Chromatographic conditions

HPLC (Shimadzu, Prominence-i 2030 model) with Lab Solution software was employed in this method. Prontosil C18 (250×4.6 mm, 5 μ) column was used for analysis. The mobile phase used was acetonitrile: 0.05% orthophosphoric acid (45:55), at the flow rate of 1.0 ml/min and the injection volume was kept 10 μ l. The column temperature was set at 28°C. Andrographolide and aloe-emodin were detected at 225 nm using a UV detector.

Selection of wavelength

Standard solutions of andrographolide and aloe-emodin were prepared and scanned by a UV spectrophotometer. The range of detection was kept from 200 to 400 nm and the overlay spectra of andrographolide and aloe-emodin obtained are shown in Fig. 1. 225 nm was selected as the detection wavelength for the analysis of andrographolide and aloe-emodin as both the markers showed appreciable absorption at 225 nm.

Preparation of standard solutions

Hundred mg of each marker (andrographolide and aloe-emodin) was transferred individually in two volumetric flasks of 100 ml and the volume was made up with methanol to obtain solutions of 1000 μ g/ml. These were used as stock solutions and were used after suitable dilutions.

Preparation of working solutions

Working solutions were prepared from the standard solution of markers. A combined solution of markers having a concentration of 100 μ g/ml was prepared from the stock solution. This was further diluted to get dilutions of 0.5, 1, 5, 10, 20, 50, and 60 μ g/ml which were used to construct a calibration curve.

Preparation of sample solution

Ten tablets were triturated and about 2 g powder was weighed and was subjected to reflux using methanol as extracting solvent. Triturated powder was transferred into the 100 ml round bottom flask and 100 ml methanol was added to it and was placed in a heating mantle and extraction was continued for 20 min. The solution was further filtered using the Whatman Filter Paper to get a clear solution and the volume

was made up to 100 ml using methanol. This solution was sonicated before injection.

RESULTS AND DISCUSSION

Method development

A series of trials was carried out using various mobile phases such as acetonitrile: Phosphate buffer (60:40) having different pH, acetonitrile: 0.1% orthophosphoric acid (50:50), acetonitrile: 0.05% orthophosphoric acid (40:60) to develop RP-HPLC method for simultaneous estimation of andrographolide, and aloe-emodin in the marketed formulation. Finally, acetonitrile: 0.05% orthophosphoric acid (45:55) was selected as the mobile phase based on better peak resolution and peak symmetry. Prontosil C18 column (250×4.6 mm, 5 μ) was used for analysis and injection volume was kept 10 μ l. The flow rate was 1.0 ml/min and the run time was 15 min. The column temperature was set at 28°C and the detection was carried out at 225 nm. The retention time (RT) of andrographolide and aloe-emodin obtained was found to be 4.57±0.2 min and 12.29±0.2 min, respectively. Chromatograms of standard and sample of andrographolide and aloe-emodin are shown in Figs. 2 and 3. The optimized chromatographic conditions are tabulated in Table 1.

Method validation

The developed method was validated for parameters such as linearity, specificity, precision, accuracy, robustness, and solution stability as per the International Conference on Harmonization (ICH) guidelines [18].

Specificity

It is performed to ensure the identification, purity testing, and quantification of marker compound from the ayurvedic formulation under analysis. Specificity was confirmed by comparing the RTs and UV spectra of the standards with the component obtained in chromatograms of the extract of tablets. The developed method was found to be specific as there was no interference of any other constituents at the RTs of both markers andrographolide and aloe-emodin as depicted in Figs. 2 and 3.

Linearity

Linearity was evaluated by analyzing the plot area as a function of the concentration of analyte. Andrographolide and aloe-emodin showed a linear response in the concentration range of 0.5–60 μ g/ml. The linearity was constructed by plotting peak area versus concentration of analyte. The linearity was validated by the high value of correlation coefficients (r^2) 0.9992 and 0.999 for andrographolide and aloe-emodin, respectively, which meets the acceptance criteria for method validation. The results are tabulated in Table 2 and plots obtained are given in Figs. 4 and 5.

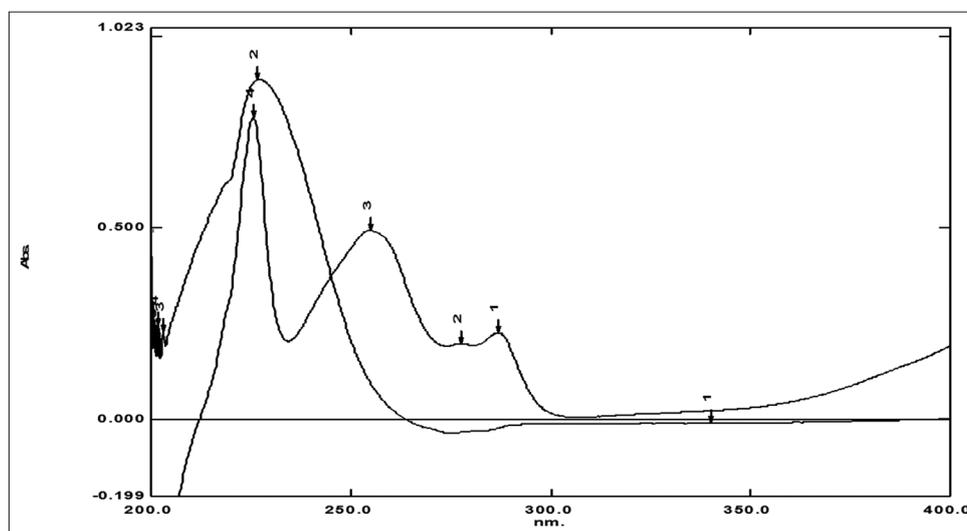


Fig. 1: Ultraviolet overlap spectrum of andrographolide and aloe-emodin

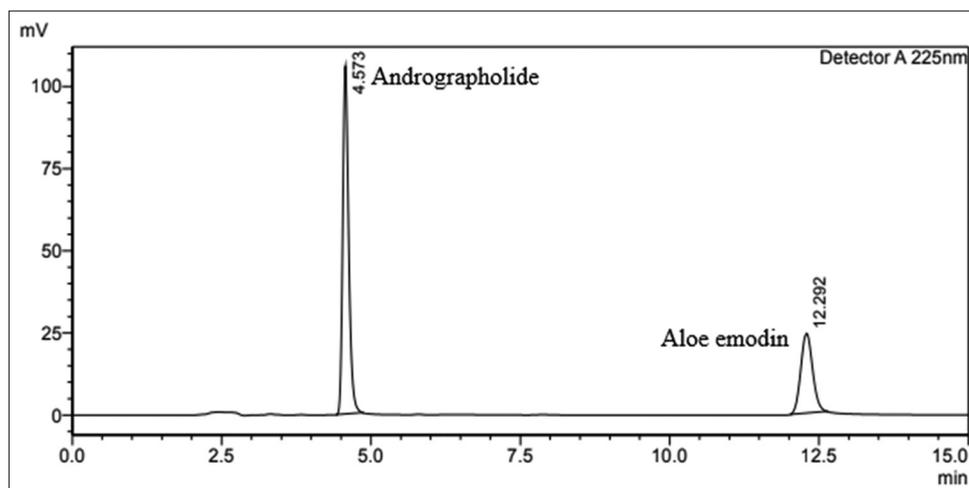


Fig. 2: High-performance liquid chromatography chromatogram of a standard mixture of andrographolide and aloe-emodin obtained using optimized conditions

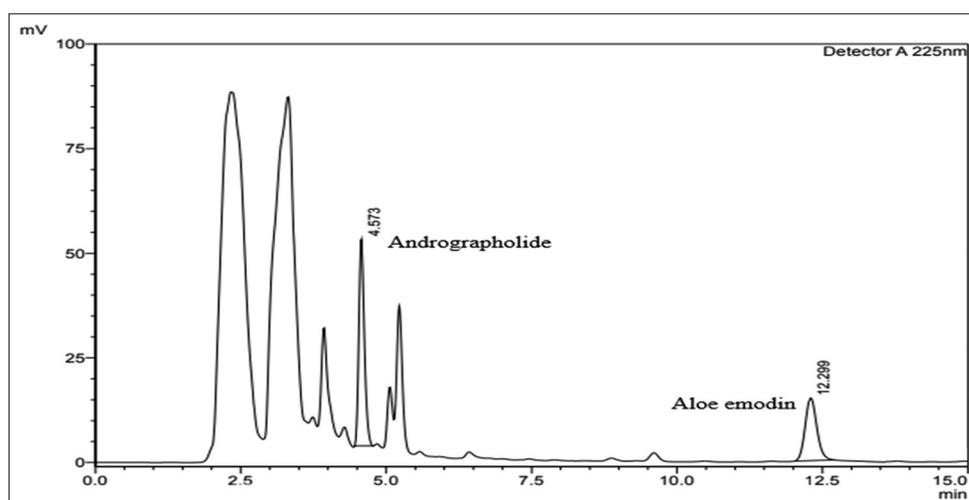


Fig. 3: Chromatogram of extract of marketed formulation

Table 1: Optimized chromatographic conditions for andrographolide and aloe-emodin

Parameters	Optimized conditions
Column	Prontosil C18, (250×4.6 mm, 5 μ)
Mobile phase	Acetonitrile: 0.05% orthophosphoric acid (45:55)
Detector	UV detector
Detection wavelength	225 nm
Column temperature	28°C
Injection volume	10 μl
Flowrate	11.0 ml/min
Run time	15 min
Retention time	4.57 and 12.29 min

UV: Ultraviolet

Limit of detection (LOD)

The LOD of an individual analytical method is the lowest amount of analyte in a sample which can be detected but not necessarily quantified as an exact value. The LOD is expressed as $LOD=3.3 \sigma/S$, where σ =standard deviation of intercepts of the calibration curve and S is the Slope of the calibration curve.

Limit of quantification (LOQ)

LOQ is a parameter of quantitative assays for low levels of compounds (markers) in extracts. The LOQ is expressed as $LOQ=10 \sigma/S$.

LOD and LOQ of andrographolide were found to be 0.14 and 0.44 μg/ml, respectively, and that of aloe-emodin was found to be 0.13 and 0.40 μg/ml, respectively. A low LOD and LOQ value indicates that the method is sensitive.

Quantification of markers

The amount of andrographolide and aloe-emodin present in the formulation was calculated using linear regression analysis. Quantification of the markers was done by performing HPLC analysis of test solutions. The area obtained for each of the markers from formulation was extrapolated on the calibration curve of the respective marker. The results are shown in Table 3.

Precision

The system precision was carried out by injecting six injections of standards of andrographolide and aloe-emodin and method precision was performed by injecting a sample of the same concentration 6 times. The percent relative standard deviation (%RSD) was calculated from the area obtained from the chromatogram. The standard analysis of the results proved that %RSD of the peak areas obtained was <2%; hence, the developed method was found to be precise. The data of precision are tabulated in Tables 4 and 5.

Accuracy (recovery)

Recovery of andrographolide and aloe-emodin from formulation was checked by spiking a known quantity of standards at three concentration

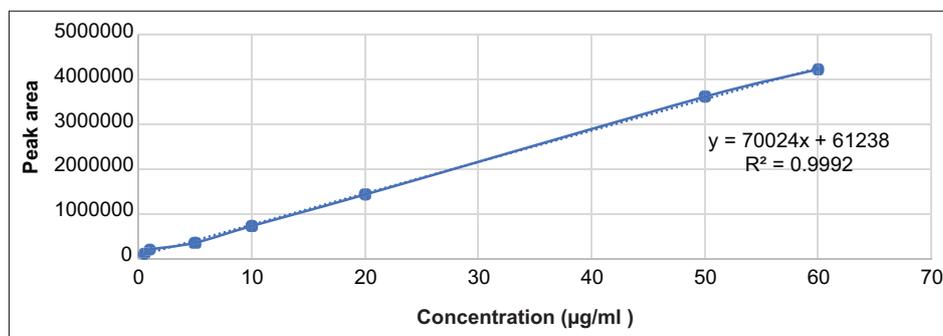


Fig. 4: Calibration curve of andrographolide

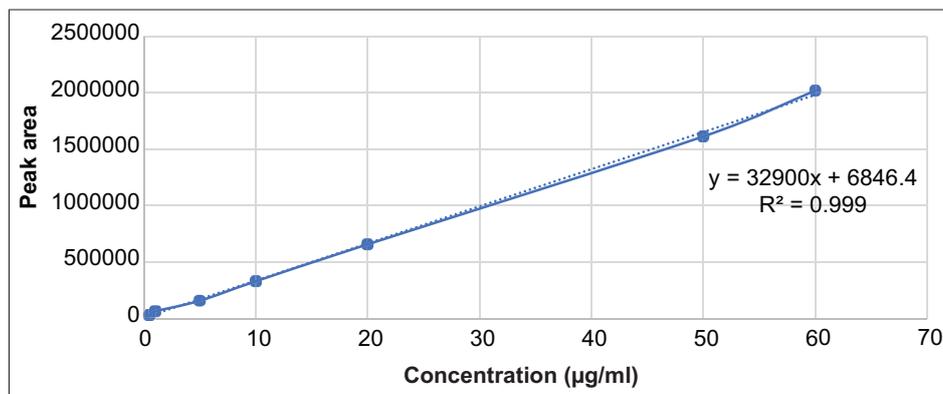


Fig. 5: Calibration curve of aloe-emodin

Table 2: Linear regression data obtained from calibration curves of andrographolide and aloe-emodin

Concentration (µg/ml)		Area	
Andrographolide	Aloe-emodin	Andrographolide	Aloe-emodin
0.5	0.5	109,805	30,416
1	1	202,898	62,888
5	5	357,499	157,427
10	10	731,227	329,612
20	20	1,433,500	656,922
50	50	3,619,588	1,612,455
60	60	4,232,730	2,018,109
Slope	Slope	70,024	32,900
Intercept	Intercept	61,238	68,464
Correlation	Correlation	0.9992	0.999

Table 3: Analysis of markers in formulation

Markers	%w/w content
Andrographolide	0.0023
Aloe-emodin	0.0115

Table 4: System precision results

S. no.	Andrographolide (10 µg/ml)	Aloe-emodin (20 µg/ml)
	Peak area	Peak area
1.	741,227	656,922
2.	741,708	654,925
3.	744,624	649,835
4.	749,309	653,309
5.	741,889	652,233
6.	741,264	652,590
Mean±SD	743,337±3188	653,302±2425
%RSD	0.43	0.37

%RSD: Percentage relative standard deviation, SD: Standard deviation

Table 5: Method precision results

S. no.	Andrographolide	Aloe-emodin
	Peak area	Peak area
1.	125,903	158,946
2.	124,982	157,982
3.	125,467	158,268
4.	125,948	158,923
5.	125,120	157,689
6.	123,832	159,893
Mean±SD	125,209±781	158,617±802
%RSD	0.62	0.51

%RSD: Percentage relative standard deviation, SD: Standard deviation

levels (i.e., 80%, 100%, and 120% of the quantified amount) to the test samples in triplicate using HPLC. This way, accuracy was performed and calculated for nine determinations over a specified range and mean recovery was calculated. The acceptance limit for percent recovery ranges from 98 to 102%. The mean % recovery was found to be within the range, which indicates that the method is accurate. The percentage of recovery results is tabulated in Tables 6 and 7.

Table 6: % recovery results for andrographolide

Level (%)	Sample (µg/ml)	Standard added (µg/ml)	Total amount	Area (n=3)*	Recovery	% recovery
80	4.62	3.69	8.31	637535	8.23	99
100	4.62	4.62	9.24	697055	9.08	98.26
120	4.62	5.54	10.16	769880	10.12	99.6

*n: number of injections

Table 7: % recovery results for aloe-emodin

Level (%)	Sample (µg/ml)	Standard added (µg/ml)	Total amount	Area (n=3)	Recovery	% recovery
80	23.1	18.48	41.58	1370880	41.46	99.71
100	23.1	23.1	46.2	1519588	45.98	99.52
120	23.1	27.72	50.82	1681127	50.89	100.1

Table 8: Robustness results of andrographolide and aloe-emodin

Parameter	Deviation	%RSD			
		Andrographolide		Aloe-emodin	
		Area	RT	Area	RT
Flow rate (ml/min)	0.8 ml	0.45	0.37	0.41	0.46
	1.2 ml	0.92	1.23	0.78	0.65
Column temperature	27°C	0.48	0.26	0.60	0.38
	29°C	0.87	0.40	0.35	0.21
Wavelength	224 nm	0.32	0.48	0.95	0.33
	226 nm	0.58	0.29	0.98	0.17

RT: Retention time, %RSD: Percentage relative standard deviation

Table 9: Solution stability of andrographolide and aloe-emodin

Time	%RSD	
	Andrographolide (20µg/ml)	Aloe-emodin (20µg/ml)
Initial	0.40	0.52
6 h	0.37	0.66
12 h	0.44	0.71
24 h	0.50	0.82

%RSD: Percentage relative standard deviation

Robustness

Robustness of the analytical method was evaluated by making deliberate changes in the chromatographic conditions such as flow rate of mobile phase (± 0.2 ml/min), wavelength (± 1 nm), and column temperature ($\pm 1^\circ\text{C}$). Each marker was analyzed in triplicate in order. The %RSD was found to be within limits this ensured that the method is robust. The results of robustness are shown in Table 8.

Solution stability

The solution of andrographolide and aloe-emodin was injected at different time intervals for evaluating the stability of the solution. The %RSD was calculated. The solution stability of 24 h showed that the solution can be used over 24 h without any degradation. The results are depicted in Table 9.

CONCLUSION

A novel HPLC method was developed and validated for the simultaneous estimation of andrographolide and aloe-emodin. This method was validated according to the ICH Q2 (R1) guidelines in the terms of linearity, precision, LOD, LOQ, accuracy, and robustness. The developed method is simple, linear, robust, precise, and accurate for the determination of andrographolide and aloe-emodin and can be used for the analysis of both the markers in the formulation. Peaks of both the markers were sharp and well resolved. Both the markers were quantified from formulation under study. Hence, the proposed

method can be applied for routine qualitative and quantitative analysis of andrographolide and aloe-emodin in an ayurvedic formulation containing these phytoconstituents.

ACKNOWLEDGMENT

The authors are thankful to the Oriental College of Pharmacy for providing necessary facilities for research.

AUTHORS' CONTRIBUTIONS

All the authors have contributed equally in performing analysis and writing the manuscript.

CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest in this research work.

AUTHORS' FUNDING

The authors are thankful to Oriental College of Pharmacy for funding the project.

REFERENCES

1. Arunachalam KD, Kuruva JK, Hari S, Annamalai SK, Baskaran KV. HPTLC finger print analysis and phytochemical investigation of *Morinda tinctoria* roxb leaf extracts by HPLC and GC MS. Int J Pharm Pharm Sci 2015;7:360-6.
2. Shaikh H, Jain V. A novel, simple, rapid RP-HPLC method for simultaneous estimation of ferulic acid, quercetin, piperine and thymol in Ayurvedic formulation. Int J Appl Pharm 2018;10:303-8.
3. EMEA. Quality of Herbal Medicinal Products. Guidelines. European Agency for the Evaluation of Medicinal Products (EMA): London; 1998.
4. Ong ES. Extraction methods and chemical standardization of botanicals and herbal preparations. J Chromatogr B Analyt Technol Biomed Life Sci 2004;812:23-33.
5. Lokhande S, Chougule A, Patil S, Patil V. Need of herbal drug standardization. Int Ayurvedic Med J 2015;3:874-7.
6. Kotagiri R, Kanaujia A, Singh P, Thakur D. Validated RP-HPLC method for the quantification of andrographolide in toxiroak premix, a polyherbal mycotoxin inhibitor. Int J Pharm Sci Res 2013;4:2623-8.
7. Xu W, Yang W. Assay of andrographolide bulk drug by HPLC. Zhongguo Zhong Yao Za Zhi 2010;35:2113-5.
8. Bhoje SG, Kuber VV, Nagore DH, Gaikwad PS, Patil MJ. Development and validation of RP-HPLC method for simultaneous analysis of andrographolide, phyllanthin and hypophyllanthin from herbal hepatoprotective formulation. Acta Chromatogr 2013;25:159-69.
9. Kumaran KS, Thirugnanasambantham P, Vishwanathan S, murthy MS. An HPLC method for the estimation of andrographolide in rabbit serum. Indian J Pharmacol 2003;35:109-12.
10. Mandrioli R, Mercolini L, Ferranti A, Fanali S, Raggi MA. Determination of aloe emodin in *Aloe vera* extracts and commercial formulations by HPLC with tandem UV absorption and fluorescence detection. Food Chem 2011;126:387-93.
11. Tabin S, Gupta RC, Bansal G, Kamili AN. Comparative HPLC analysis

- of emodin, aloe emodin and rhein in *Rheum emodi* of wild and *in vitro* raised plants. J Pharmacogn Phytochem 2016;5:121-30.
12. Zaffaroni M, Mucignat C, Pecera T, Zagotto G, Frapolli R, D'Iucalci M, et al. High-performance liquid chromatographic assay for the determination of aloe emodin in mouse plasma. J Chromatogr B Analyt Technol Biomed Life Sci 2003;796:113-9.
 13. New Livfit Tablet Uses, Side effects, Reviews, and Precautions Alembic Pharma Tablet Wise India. Available from: <https://www.tabletwise.com/new-livfit-tablet>. [Last accessed on 2019 Sep 18].
 14. Joseph J, Solomon J. *Andrographis paniculata*: A review of its traditional uses, phytochemistry and pharmacology. Med Aromat Plants 2014;3:2-14.
 15. Handa SS, Sharma A. Hepatoprotective activity of andrographolide against galactosamine and paracetamol intoxication in rats. Indian J Med Res 1990;92:284-92.
 16. Malik MA, Bhat SA, Bilquees F, Sheikh BA, Siddiqui S, Shrivastava P. *Rheum emodi* as valuable medicinal plant. Int J Gen Med Pharm 2016;5:35-44.
 17. Sanders B, Ray AM, Goldberg S, Clark T, Clark T, McDaniel HR, et al. Anti-cancer effects of aloe-emodin: A systematic review. J Clin Transl Res 2017;3:283-96.
 18. ICH Harmonised, Tripartite Guideline Validation of Analytical Procedures: Text and Methodology Q2(R1), Geneva; 2005. p. 1-13.