

TRANSDERMAL DELIVERY OF AN EFFECTIVE NONSTEROIDAL ANTI-INFLAMMATORY DRUGS FOR PAIN MANAGEMENT IN ARTHRITIS

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ABSTRACT

Objective: The current research work has been carried out with the aim to develop a transdermal gel formulation of fenoprofen (a nonsteroidal anti-inflammatory drug used to treat pain associated in arthritis) which would overcome the gastrointestinal-related problems associated with oral administration of the drug. The present study aims at formulating transdermal gels using different concentrations of Carbopol, hydroxypropyl methylcellulose (HPMC), sodium alginate, and guar gum.

Methods: The formulated gels were subjected for various evaluation tests such as clarity, homogeneity, viscosity, drug content, pH, spreadability, and *in vitro* permeation studies. Drug-polymer interaction was studied by Fourier transmission infrared (FTIR) and differential scanning calorimetry (DSC). The *in vitro* permeation studies were performed in phosphate buffer 7.4 using Franz diffusion cell.

Results: The FT-IR and DSC studies showed no chemical interaction between drug and polymers used. All the formulated gels showed acceptable physical properties with respect to clarity, homogeneity, viscosity, drug content, pH, and spreadability. Among all the gel formulations, Carbopol gels containing fenoprofen showed good drug release compared to HPMC, sodium alginate, and guar gum. Optimized formulation was further subjected to kinetic studies which showed Higuchi model of drug release. The same formulation showed significant anti-inflammatory and analgesic activity, tested in Wistar albino rats. No signs of erythema, edema, flushing, and papules were observed when skin irritation test was performed. Stability studies under accelerated condition showed satisfactory results for the optimized formulation.

Conclusions: Thus, it was concluded from the results that the optimized formulation showed controlled and slow drug delivery. Animal studies were significant at $p < 0.05$ and 0.001 . The selected formulation was stable at various ambient temperatures.

Keywords: Fenoprofen, Transdermal, Carbopol 940, Hydroxypropyl methylcellulose, Sodium alginate, Guar gum.

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INTRODUCTION

Transdermal route has gained accolade as it has several advantages over conventional forms such as, avoids first pass metabolism and lowers gastrointestinal irritation that are associated with oral administration [1]. Easy termination of therapy enables a constant plasma level profile that results in decreased side effects are some other advantages [2]. The release of the drugs from topical preparations depends on the physicochemical properties of the drug and gels employed. Gels for dermatological use have many advantageous properties such as thixotropic, emollient, greaseless, easily spreadable, and easily removable [3]. Gelling agents when mixed with appropriate solvent entangle to form a three-dimensional colloidal network that limits fluid flow by entrapment and immobilization of the solvent molecules. One more advantage of network structure of gels is their resistance to deformation and hence its viscoelastic properties [4,5].

Fenoprofen is a propionic acid derivative with analgesic, anti-inflammatory, and antipyretic properties. Oral administration of fenoprofen can cause gastric mucosal damage, respiratory depression, and metabolic acidosis. Its high dose (300–600 mg, 3–4 times daily) causes more plasma fluctuation. To avoid invasive drug therapy and to eliminate frequent dosing with oral administration, a transdermal route has been studied as an alternative dosage form. The low molecular weight, less biological half-life, good analgesic activity, and anti-inflammatory activity make fenoprofen a good candidate for transdermal delivery [6-8].

Hence, the current research was planned to formulate and evaluate gel formulation containing fenoprofen using Carbopol 940, hydroxypropyl

methylcellulose (HPMC), sodium alginate, and guar gum. The prepared gels were evaluated for clarity, homogeneity, viscosity, drug content, pH, spreadability, and *in vitro* permeation studies. Finally, the optimized formulation was further subjected for animal studies and stability studies.

MATERIALS AND METHODS

Materials

Fenoprofen was obtained as gift sample from D.K Pharma, Mumbai, Carbopol 940, HPMC, sodium alginate, guar gum, ethanol, triethanolamine, and oleic acid were purchased from S.D. Fine Chemicals Limited, Mumbai. All the chemicals obtained were of analytical grade.

Methods

Fourier transmission infrared (FTIR) studies

The FTIR spectra [9] of the pure drug fenoprofen and its physical mixture with various polymers such as Carbopol 940, HPMC, sodium alginate, and guar gum were recorded using FTIR spectrophotometer (PerkinElmer 1600 series, USA). The samples were prepared by potassium bromide press pellet technique and scanned for the absorbance at $4000-400\text{ cm}^{-1}$.

Differential scanning calorimetry (DSC)

The DSC thermograms of the pure drug fenoprofen and its physical mixture with various polymers such as Carbopol 940, HPMC, sodium alginate, and guar gum were recorded using PerkinElmer 1600 series USA instrument, to know any interaction between drug

and the various polymers used. All the samples were placed in a sealed aluminum pans and scanned for the endothermic peaks. The heating rate selected was from 50°C to 300°C at an increase of 10°C per min [10].

Preparation of transdermal gels

All the gelling agents were optimized for the optimum concentration at which they formed the gels. It was found that Carbopol, HPMC, sodium alginate, and guar gum formed proper gels at 1%, 2%, 4%, and 6 % w/w, respectively.

Carbopol gel was prepared by soaking required quantity of Carbopol 934 in water for a period of 3 h. Then, specified amount of drug was dissolved in ethanol, to this required quantity of propylene glycol was added and stirred. This drug solution was transferred to Carbopol container and stirred for 20 min. Drug containing Carbopol gel was neutralized using triethanolamine with continuous stirring. The dispersion was then allowed to hydrate and swell for 1 h; finally, the desired pH of 6.8–7 was adjusted using triethanolamine with continuous stirring until a homogeneous gel was formed. Hence, prepared Carbopol gel was allowed to equilibrate for 24 h at room temperature until further evaluation [11-13].

HPMC, sodium alginate, and guar gum were optimized for polymer concentration. Transdermal gels containing HPMC, sodium alginate, and guar gum were prepared by dispersing 2% HPMC, 4% sodium alginate, and 6% guar gum in water by continuous stirring for a period of 2 h. Fenopfen was dissolved in ethanol and the drug solution was added gently to various polymeric solutions with continuous stirring until homogeneous gel was formed. Finally, all the gels were allowed to equilibrate at room temperature for 24 h until further evaluation [14-16]. Detailed formulation is shown in Table 1.

Evaluation of transdermal gels

Physical appearance and clarity

The prepared fenopfen gels were inspected visually for their color and transparency [17]. Clarity of various formulations was determined by visual inspection under black and white background and it was graded as turbid +; clear ++; and very clear (glassy) +++.

Homogeneity

All the formulated gels were tested for homogeneity [18] by visual inspection after the gels have been set in the container. They were tested for their appearance and presence of any aggregates.

Viscosity

Viscosity [19] was determined using Brookfield viscometer. Viscosity measurements were carried out at room temperature (25–27°C) using Brookfield Viscometer (Model RVTDV II, Brookfield Engineering Laboratories, Inc., Stoughton, MA).

Measurement of pH

The pH [20] of various gel formulations was determined using digital pH meter (Elico). One gram of gel was dissolved in 100 ml distilled

water and stored for 2 h. The electrode was inserted into the sample 10 min before taking the reading at room temperature. The measurement of pH of each formulation was done in triplicate and average values were calculated.

Drug content

A specified quantity (100 mg) of formulated gel was taken and dissolved in 100 ml of phosphate buffer pH 7.4. The volumetric flask containing gel was shaken for a period of 2 h on mechanical shaker to get absolute solubility of drug. This solution was filtered and estimated spectrophotometrically at 272 nm using phosphate buffer pH 6.8 as blank [21].

Spreadability

The spreadability [22] of the gel was determined using the following technique: 0.5 g gel was placed within a circle of 1 cm diameter pre-marked on a glass plate over which a second glass plate was placed. A weight of 500 g was allowed to rest on the upper glass plate for 5 min. The increase in the diameter due to spreading of the gels was noted. Average of triplicate readings was taken.

In vitro drug release studies

Drug permeation through rat abdominal skin was carried out after getting approval from the Institutional Animal Ethical Committee, bearing registration number 557/02/C/Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA). It was conducted as per the principles and guidelines of CPCSEA.

Franz diffusion cell was used to carry out *in vitro* permeation study through rat abdominal skin [23]. The skin was mounted between the compartments of the diffusion cell with stratum corneum facing the donor compartment. One gram of gel containing fenopfen was placed in the donor cell. Twenty milliliters of phosphate buffer (pH 7.4) solution were used as receptor medium that was maintained at 37°C and magnetically stirred at 300 rpm. At appropriate time interval, 1 ml of the sample was withdrawn from the receptor compartment and the same amount of fresh buffer solution was added to maintain sink condition. Simultaneously, a blank diffusion study was carried out in a similar manner using blank gel without fenopfen. Each experiment was carried out in triplicate. The sample was analyzed spectrophotometrically at a wavelength of 272 nm and the concentration of drug in receptor compartment was noted.

Mathematical kinetic assessment for drug release mechanisms

Release kinetics is an integral part for the development of a dosage form because, if the kinetics of drug release is known, one can also establish *in vitro-in vivo* correlation. Mathematical approach is one of the scientific methods to optimize and evaluate the error in terms of deviation in the release profiles of formulated products during the formulation development stage. Mathematical model approaches important in research and development because of its simplicity and their interrelationships may minimize the number of trials in final optimization, thereby improving the formulation development process. The permeation profile of the optimized formulation was fitted to different kinetic models.

Table 1: Formulation of various transdermal gels

S. No.	Ingredients	F1	F2	F3	F4	F5	F6	F7	F8
1.	Fenopfen (mg)	100	100	100	100	100	100	100	100
2.	Carbopol 374 (%)	0.5	1	-	-	-	-	-	-
3.	Hydroxypropyl methylcellulose (gm)	-	-	1	2	-	-	-	-
4.	Sodium alginate (gm)	-	-	-	-	3	4	-	-
5.	Guar gum (gm)	-	-	-	-	-	-	5	6
6.	Oleic acid (%wrt drug)	10	10	10	10	10	10	10	10
7.	Triethanolamine (%)	1	1	1	1	1	1	1	1
8.	Water	qst							

wrt: With respect to, qst: Quantity sufficient to

Qt versus t (zero order)
 Log (Q₀-Qt) versus t (first order)
 Qt versus square root of t (Higuchi)
 log %Qt versus log t (Korsmeyer-Peppas)

Where, Qt is the amount of drug released at time t, Q₀ is amount of drug release at time 0, and t is time in hour.

Animal studies

All the animal studies were carried out after getting approval from the Institutional Animal Ethical Committee, bearing registration number 557/02/C/CPCSEA. It was conducted as per the principles and guidelines of CPCSEA.

Anti-inflammatory activity

Wistar albino rats were divided into two groups of six animals each. The ventral surface of the animals was depilated and one group was treated as control and the other group was treated as test. About 2% w/v formalin solution was used as chronic inflammogen to induce inflammation in all animals by subcutaneous route. A mark was made on hind paw just behind tibiotarsal junction so that every time the paw was dipped in the mercury column up to the fixed mark to ensure constant paw volume. The test gel containing the dose of fenopropfen equivalent the body weight of the animal was applied to the animal of test group. The paw volume of all the animals, both control and test groups, was measured using plethysmograph at selected interval of time. Finally, the percentage reduction in edema volume was calculated and the activity of the formulation was statically analyzed by Student's "t" test [24].

Analgesic activity

Twelve albino mice were selected and divided into two groups each having six rats. The ventral surface of the animals was depilated and divided into Group 1 (control) and Group 2 (test). A blank gel containing no drug was applied to control group. The selected gel formulation containing the dose of fenopropfen equivalent to body weight was applied on the ventral surface of the test animal group. About 0.6%v/v acetic acid was injected intraperitoneally to all the animals (1 ml/100 g of body weight of animal) to induce writhes. The number of writhes produced in 15 min was noted. The activity of the formulation was statistically analyzed by Student's "t" test [25].

Skin irritation test

Skin irritation tests were performed for optimized fenopropfen transdermal gel on the albino rats to find out any irritation problems which could reject its suitability for topical use. Approximately 1 g of gel was topically applied for the albino rats across a 2 square inch area and was observed for any show any signs of erythema, edema, flushing, and papules for a period 24 h [26].

Stability studies of optimized formulation

Stability studies were conducted according to the ICH guidelines [27] by storing the best selected formulation at room temperature and at 40 ± 2°C/75% RH in stability chamber for 3 months. The formulation was analyzed for the change in appearance, pH, and drug content by procedure stated earlier.

RESULTS AND DISCUSSION

FTIR studies

The principal peaks of fenopropfen were observed at 3647 cm⁻¹ [N-H stretch], 3070 cm⁻¹ [C-H aromatic], 2420 cm⁻¹ [O-H bending], 1423 cm⁻¹ [aromatic C=C stretch], and 1566 cm⁻¹ [C=O stretch]. FTIR of physical mixture showed similar peaks indicating no interaction between drug and the polymers employed in formulation. Figs. 1 and 2 show the spectra of pure drug and physical mixture, respectively.

DSC

The thermogram of pure fenopropfen exhibited a sharp endothermic peak at 125.08°C corresponding to its melting point (118–120°C). Similarly, the thermograms of physical mixtures of fenopropfen with excipients under study exhibited the endothermic peak in the vicinity of its melting point range indicating the absence of any drug excipient interaction. The DSC thermograms are depicted in Fig. 3.

Evaluation of transdermal gels

Physical appearance and clarity

The prepared fenopropfen gel formulations were transparent in Carbopol 934, white viscous in HPMC, and brownish gummy in sodium alginate and guar gum. All gels were found to be transparent and were free from the presence of particles. Results are tabulated in Table 2.

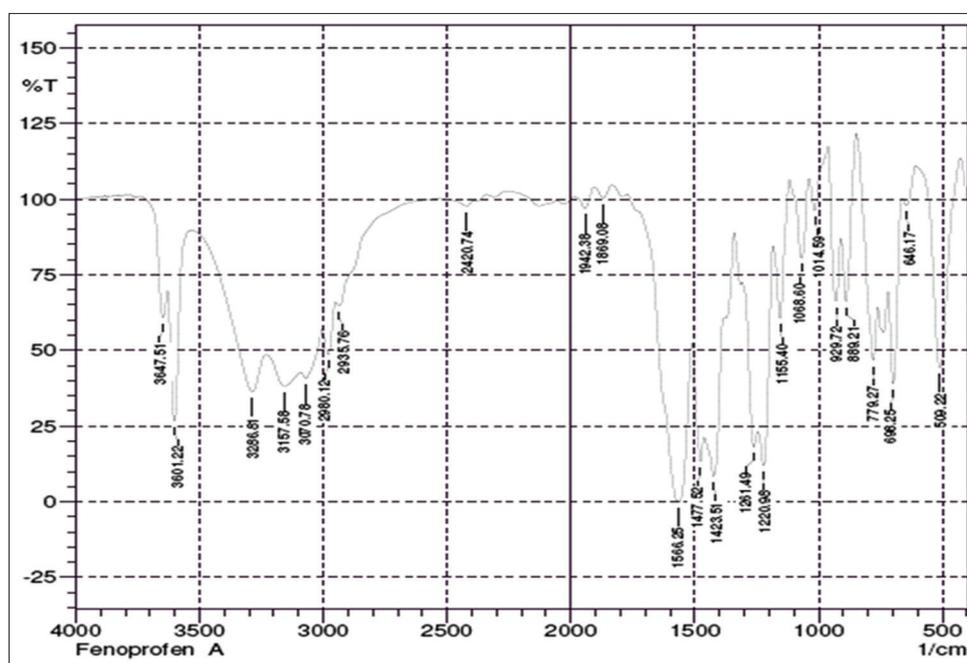


Fig. 1: Fourier transmission infrared of fenopropfen drug alone

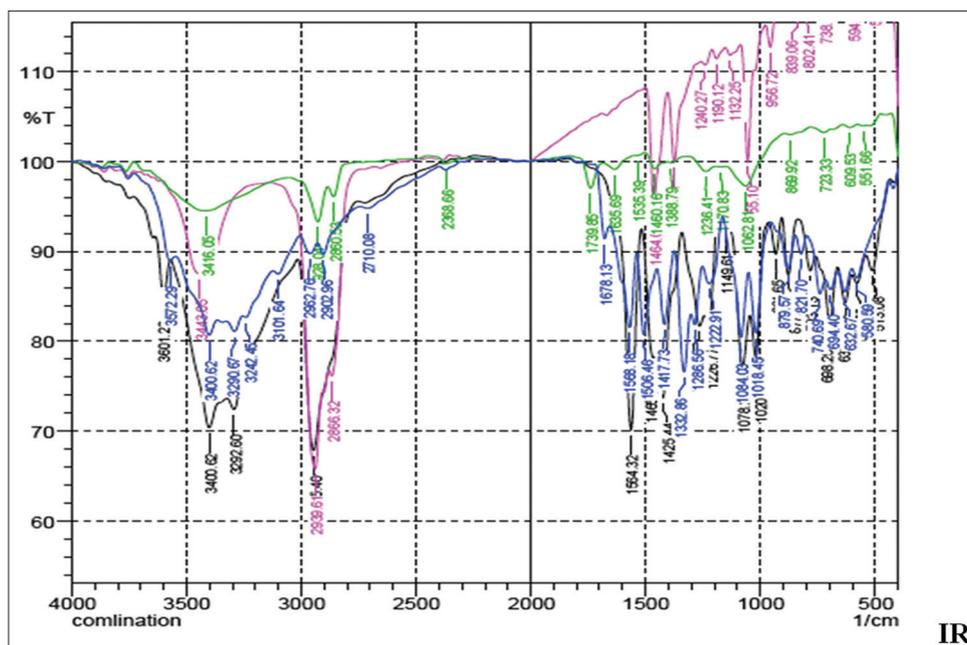


Fig. 2: Fourier transmission infrared spectrum of pure drug fenoprofen with polymers used

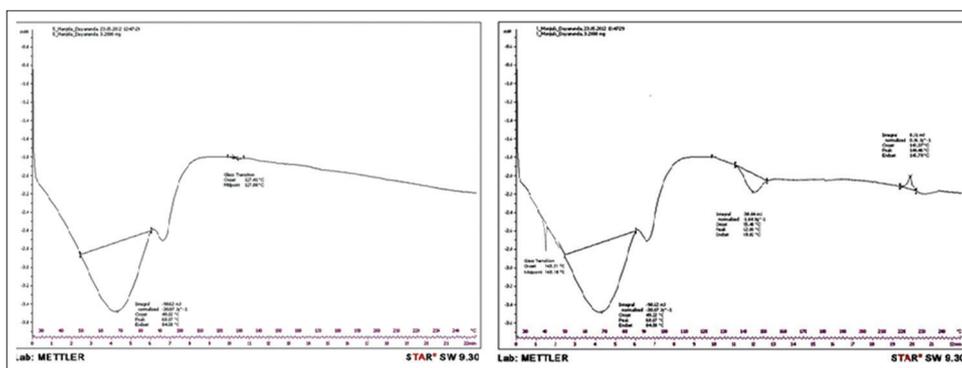


Fig. 3: Differential scanning calorimetry thermograms (a) fenoprofen drug alone (b) physical mixture of drug and polymers used (Carbopol 940, hydroxypropyl methylcellulose, sodium alginate, and guar gum)

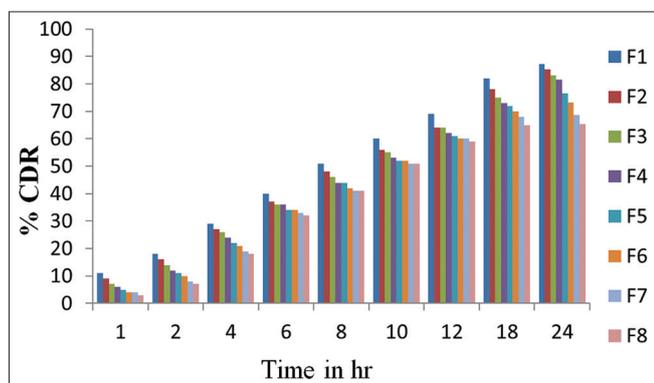


Fig 4: Cumulative drug release from F1 to F8 at different time intervals

Homogeneity

All the formulations were found to be smooth and homogeneous with the absence of lumps as shown in results (Table 2).

Viscosity

Viscosity of various formulated gels was found in the range of $0.492 \pm 0.28 - 0.689 \pm 0.33$ poise, results are depicted in Table 3.

pH

The pH of the formulated gels was found to be in the range of 5.7–6.9, which lies in the normal pH range of the skin, so the formulations would not produce any skin irritation. pH of all formulation is tabulated in Table 3.

Drug content

The uniformity in drug distribution throughout the gels was estimated by drug content studies. The drug content of the gel formulations was in the range of $82.2 \pm 0.38 - 93.5 \pm 0.19$ showing content uniformity, Table 3.

Spreadability

Spreadability diameter for various formulated gels showed good spreadability, i.e., all the gels were easily spreadable. The results are shown in Table 2.

In vitro drug release studies

In vitro permeation studies of fenopropfen gels containing different concentrations of Carbopol (0.5% and 1%), HPMC (1% and 2%), sodium alginate (3% and 4%), and guar gum (5% and 6%) were carried out using Franz diffusion cell containing pH 7.4 phosphate buffer as medium and measuring drug concentration by ultraviolet spectrophotometer. The percentage cumulative drug release at different time intervals is depicted in Table 4 and Fig. 4. The percentage drug release for the formulations containing drug and Carbopol (F1 and F2) was found to be 87.23% and 85.28%, respectively, for formulations containing drug and HPMC (F3 and F4) was found to be 84.15% and 83.57%, respectively, for formulations containing drug and sodium alginate (F5 and F6) was found to be 79.49% and 78.18%, respectively, and that for guar gum formulations (F7 and F8) was found to be 76.56% and 75.32%, respectively. In general, it was observed that an increase in polymer content was associated with a corresponding decrease in the percentage drug release among Carbopol, HPMC, sodium alginate, and guar gum. This could be attributed to two

reasons, first, extensive swelling of the polymer which created a thick gel barrier for drug diffusion [28]. Second, as polymer concentration increased viscosity also increased, which has a negative relation to release of active drug from formulations and its penetration through the diffusion barriers [29]. Thus, both high concentration of polymer and high viscosity compete each other in controlling the release of drug substance from the formulations.

Drug release kinetics

In vitro drug release data of all formulations from F1 to F8 were fitted to zero-order, first-order, Higuchi, and Korsmeyer–Peppas equations to ascertain the pattern of drug release. The results are depicted in Table 5. When zero-order and first-order drug release kinetics were compared, the R^2 values were found to be higher in zero order. Hence, this concluded that all the formulations followed zero-order kinetics. Whereas, in case of mechanism of drug release, between Higuchi and Korsmeyer–Peppas equation, the R^2 value was found to be higher in Korsmeyer–Peppas equation and release exponent “ n ” < 1. This indicates

Table 2: Evaluation of physical appearance, clarity, homogeneity, and spreadability of various gel formulations

S. No.	Formulation codes	Physical appearance and clarity	Homogeneity	*Spreadability×cm/s
1.	F1	White, viscous, transparent +++	Homogeneous	19.12±0.46
2.	F2	White, viscous, transparent +++	Homogeneous	21.23±0.35
3.	F3	White, viscous, translucent ++	Homogeneous	26.16±0.72
4.	F4	White, viscous, translucent ++	Homogeneous	29.42±0.88
5.	F5	Brownish, gummy ++	Homogeneous	24.56±0.21
6.	F6	Brownish, gummy ++	Homogeneous	25.17±0.89
7.	F7	Light brown, gummy ++	Homogeneous	22.65±0.69
8.	F8	Light brown, gummy ++	Homogeneous	24.32±0.87

+++ : Excellent, ++ : Good. *Average of triplicate readings

Table 3: Evaluation of viscosity, pH, drug content, and flux of various gel formulations

S. No.	Formulation codes	*Viscosity (poise)	*pH	*Drug content (%)	*Flux $\mu\text{g}/\text{h}\times\text{cm}^2$
1.	F1	0.562±0.76	7.3±0.29	93.5±0.19	7.81±0.35
2.	F2	0.593±0.81	7.5±0.55	92.1±0.24	7.32±0.51
3.	F3	0.613±0.25	6.9±0.81	89.5±0.96	6.94±0.48
4.	F4	0.689±0.33	7.1±0.80	88.6±0.87	6.28±0.61
5.	F5	0.552±0.91	6.8±0.35	85.9±0.39	5.82±0.74
6.	F6	0.587±0.53	6.3±0.86	85.3±0.47	5.48±0.82
7.	F7	0.492±0.28	6.5±0.47	82.2±0.38	4.93±0.79
8.	F8	0.498±0.76	6.7±0.51	82.8±0.59	4.16±0.53

*Average of triplicate readings

Table 4: Percent drug release of various gels containing fenopropfen

Time in h	Formulation codes							
	F1	F2	F3	F4	F5	F6	F7	F8
1	11.12	9.12	7.62	6.13	5.23	4.13	4.55	3.57
2	18.22	16.31	14.16	13.54	13.22	12.26	11.12	11.78
4	29.13	27.25	26.83	24.25	24.15	21.37	19.23	18.82
6	40.42	37.13	36.19	36.44	34.27	34.43	33.34	32.91
8	51.34	48.65	46.53	44.29	44.35	42.52	41.45	41.47
10	60.32	56.32	55.76	53.71	52.57	52.61	51.54	51.73
12	69.71	64.55	64.62	62.13	61.62	60.26	60.53	59.52
18	82.27	78.23	75.85	73.64	72.73	70.37	68.62	65.24
24	87.23	85.28	84.15	83.57	79.49	78.18	76.56	75.32

The standard deviation of all formulations was found to be in the range of ± 0.19 – ± 0.32

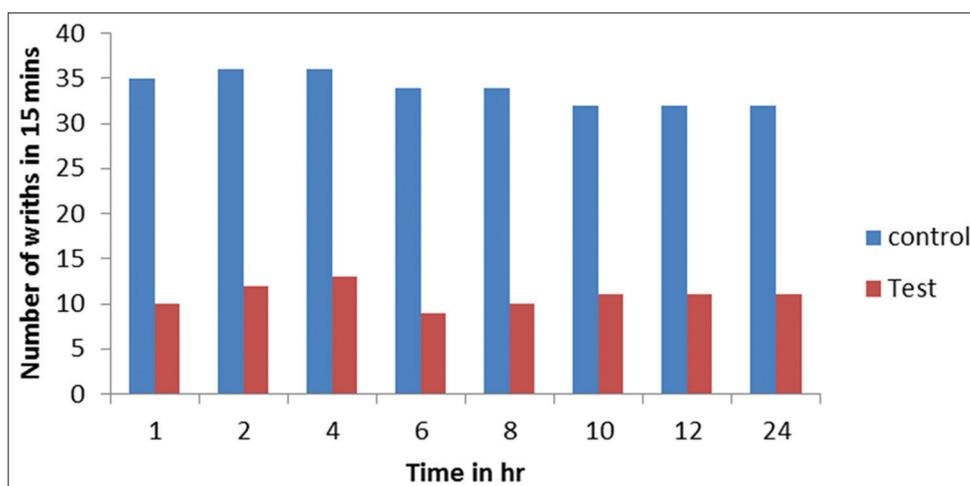


Fig. 5: Analgesic activity of F2 formulation

Table 5: Drug release kinetics of formulations F1 to F8

Formulation codes	Release kinetics			
	Zero order	First order	Korsmeyer-Peppas	Higuchi diffusion model
	R ²	R ²	R ²	R ²
F1	0.992	0.965	0.993	0.967
F2	0.995	0.961	0.996	0.969
F3	0.994	0.959	0.993	0.972
F4	0.993	0.962	0.999	0.974
F5	0.991	0.953	0.994	0.959
F6	0.996	0.958	0.997	0.951
F7	0.995	0.961	0.991	0.967
F8	0.993	0.960	0.992	0.971

Table 6: Anti-inflammatory activity of F2 formulation

S. No.	Time in h	Control mean	Test formulation	
			Mean	% reduction in edema
1.	1	0.279	0.248	10.4
2.	2	0.342	0.319	5.53
3.	4	0.373	0.301	11.59
4.	6	0.409	0.298	21.93
5.	8	0.431	0.278	32.21
6.	10	0.481	0.251	44.55
7.	12	0.533	0.218	56.89
8.	24	0.652	0.203	63.8

Table 7: Skin irritation test of F2 formulation

S. No.	Time in h	F1 formulation		
		F	P	E
1.	1	-ve	-ve	-ve
2.	2	-ve	-ve	-ve
3.	4	-ve	-ve	-ve
4.	6	-ve	-ve	-ve
5.	8	-ve	-ve	-ve
6.	10	-ve	-ve	-ve
7.	12	-ve	-ve	-ve
8.	24	-ve	-ve	-ve

-ve: No allergic manifestation observed, F: Flushing of skin, P: Papules and wheals, E: Erythema and edema

that all the formulations followed non-Fickian diffusion. Hence, it was concluded that all the formulations followed zero-order drug release with non-Fickian diffusion [30].

Animal studies

Since F2 formulation was found to be good compared to other formulations with respect to appearance, clarity, homogeneity, viscosity, spreadability, and also controlled drug release was observed in this formulation, so this particular formulation was further subjected to animal studies.

Anti-inflammatory activity of F2 formulation

Anti-inflammatory activity of F2 formulation showed 63.8% reduction of formalin-induced paw edema at the end of 24 h, as shown in Table 6. The data were analyzed by P-STAT package where F2 showed a significant anti-inflammatory activity at $p < 0.05$.

Analgesic activity of F2 formulation

Same formulation F2 was subjected for analgesic activity by acetic acid-induced writhing method for a period of 24 h. The data analyzed with P-STAT package was found to be significant at $p < 0.05$ and 0.001. The analgesic activity is depicted in the form of bar graph of number of writhes per 15 min plotted against time, as shown in Fig. 5.

Skin irritation test of F2 formulation

The animals subjected for hypersensitivity studies did not show any signs of erythema, edema, flushing, and papules for a period of 24 h (Table 7).

Stability studies

Stability studies of the selected formulation (F2) showed no significant change in its physical appearance, pH, and drug content when stored at room temperature and at $40 \pm 2^\circ\text{C}/75\% \text{RH}$ for a period of 3 months.

CONCLUSIONS

On the whole, it can be concluded that the various gels containing fenopfen were found to be satisfactory with respect to physic

chemical properties. There was also controlled drug release seen as the polymer concentration increased. Animal studies were significant at $p < 0.05$ and 0.001 . The selected formulation was stable at various ambient temperatures. Further, there is a scope for pharmacokinetic studies and its correlation with pharmacodynamics activity in higher animals.

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AUTHORS' CONTRIBUTIONS

Manjula D and Abhishek Raj designed the research work, Josephine Leno Jenita helped in animal studies, and Shanaz Banu assisted in drafting the manuscript.

CONFLICTS OF INTEREST

The authors have declared that they have no conflicts of interest with respect to current research.

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