

PHYTOCHEMICAL ANALYSIS AND ANTIOXIDANT PROPERTIES OF LEAF EXTRACTS OF *CARICA PAPAYA*

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ABSTRACT

Objective: The objective of the present study aimed at investigating the phytochemical and antioxidant properties of *Carica papaya* leaf extracts.

Methods: As phytochemicals are biologically active compounds and a powerful group of plant chemicals, believed to stimulate the immune system along with antioxidants, the molecules which hinder oxidation of other molecules by the process of inhibiting or by generating the oxidizing chain reactions and preventing diseases. The total phenolic content (TPC) was determined by Folin-Ciocalteu method and total flavonoid contents (TFC) were determined aluminum chloride method and antioxidant by 2,2,1-diphenyl-1-picrylhydrazyl method.

Results: The results of phytochemical screening revealed the presence of bioactive compounds such as alkaloid, carbohydrates, and amino acids and TPC and TFC varied among the different solvent extracts, in which methanolic extracts showed highest amount of phytochemicals and TPC and TFC and antioxidants compared to other solvents.

Conclusion: The isolation and purification of specific bioactive compound may account as natural and promising medicines in exploration of new drug.

Keywords: *Carica papaya* Linn, Phytochemicals, Antioxidants.

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INTRODUCTION

Plants are considered to be the good source for the exploration and discovery of new pharmaceutical compounds as well as medicines, which can be the potential drug for humans as they act as intermediate for synthesis of useful drug [1]. Plants possess various phytochemicals with several bioactivities such as anti-inflammatory, antioxidant, and anticancer [2] and one among them such as *Carica papaya* is widely used in the treatment of many ailments in Ayurvedic as well as herbal and folk medicine.

C. papaya Linn, is an evergreen shrub or small tree, is a member of family Caricaceae, represented with four genera and four species in India. This plant originated in Southern Mexico and Costa Rica and distributed as a plantation crop in India, Sri Lanka, Hawaii, Australia, and in tropical and subtropical regions [3]. Papaya, the herbaceous perennial plant is also known as Papaya melon tree, Pawpaw or papau, Kapaya, Lapaya, Papyas, Papye, Tapayas, and Fan mu gua. The entire papaya plant possesses various phytonutrients and can be considered for commercial, pharmaceutical, and industrial applications. The papaya plant is best with a large variety of phytonutrients and antioxidant, antimicrobial, and anti-dengue properties [4].

Phytochemicals are the natural, non-nutritive plant chemicals with defensive properties against cancer by protecting the cells from damage. Most of the phytochemicals possess the biological antioxidant capacity that protects our cells against the oxidative damage and reduces the risk of certain types of cancer. These phytochemicals tend to prevent the adhesion of pathogens to the human cell wall by physically binding to it. Some of the important phytochemicals found in *C. papaya* are Lycopene, Benzylglucosinolate, Beta-carotenoid, Benzylisothiocyanate, chlorogenic acid, caffeic acid, protocatechuic acid, Quercetin, etc. [5,6].

Antioxidants are substances that prevent oxidative damage to the target molecule. An antioxidant can scavenge the free radicals because of their singlet oxygen quenching and redox hydrogen donating features. In recent days, the usage of synthetic antioxidants has been taken over by natural antioxidants as it could be safer without any side effects. In recent decades, due to the various pharmacological actions of the medicinal plants, many researchers are showing interest in studying the antioxidant phytochemicals such as phenols, flavonoids, and tannins which have been recognized for their potential role in preventing human diseases [7-10].

METHODS

Sample collection

Fresh leaves of *C. papaya* were collected from Mysuru and authenticated by the Department of Water and Health, JSS AHER, Mysore.

Preparation of *C. papaya* leaf extract

The collected plant materials were washed with running tap water to avoid surface contaminations and shade dried for about 15 days. The dried leaves were cut into small pieces and macerated into a fine powder; dried powder was soaked with different organic solvents such as hexane, ethyl acetate, methanol, and ethanol and was subjected to solvent extraction using the Soxhlet apparatus. Then, the extracted sample was stored at 4°C for further analysis.

Phytochemical analysis

Phytochemical screening was performed for the presence of alkaloids, carbohydrates, amino acids, glycosides, protein, phenolic compounds, and tannins from respective solvents such as hexane, ethyl acetate, methanol, and ethanol, according to standard procedure [11,12].

Stock preparation

200 mg of *C. papaya* extract was dissolved in 10 ml of each solvent and 1 ml of each solvent was used as a standard for various phytochemical tests.

Alkaloids

Wagner's test – For 1 ml of the sample solution, few drops of Wagner's reagent were added along the sides of the test tube. The appearance of a reddish-brown precipitate indicates the presence of alkaloids [13].

Carbohydrates

Benedict's test – For 1 ml of the sample solution, few drops of Benedict's reagent were added and heated for 2 min. The appearance of a colored precipitate indicates the presence of carbohydrates.

Amino acids

Ninhydrin test – For 1 ml of the sample solution, two drops of ninhydrin reagent were added. The formation of purple color indicates the presence of amino acids.

Glycosides

Kellar-Killiani test – For 1 ml of sample solution 1 ml of glacial acetic acid, few drops of ferric chloride and concentrated sulfuric acid were added. The appearance of a reddish-brown ring at the junction of liquids indicates the presence of glycosides [13].

Phenolic compounds and tannins

Ferric chloride test – For 1 ml of extract, few drops of neutral 5% ferric chloride were added. The appearance of a dark green color indicates the presence of phenolic and tannin compounds.

Protein

Biuret test – For 1 ml of extract, one drop of 2% copper sulfate and 1 ml of ethanol and potassium hydroxide were added. The presence of the pink color of the ethanolic layer indicates the presence of protein.

Saponins

For 1 ml of extract, few drops of distilled water were added and shaken vigorously. The appearance of foam indicates the presence of saponins [13].

Quinones

For 1 ml of extract, few drops of concentrated hydrochloric acid were added. The formation of a yellow precipitate indicates the presence of quinones.

Oxalate

For 1 ml of extract, few drops of glacial acetic acid were added. The appearance of greenish-black coloration indicates the presence of oxalate.

Anthocyanins

For 1 ml extract, 2 ml of hydrochloric acid and 1 ml of ammonia were added. The color change from pink-red to blue-violet indicates the presence of anthocyanins.

Determination of total phenolic content (TPC)

The TPC of the *C. papaya* sample was determined spectrophotometrically according to the Folin-Ciocalteu colorimetric method with slight modifications [11,12].

A standard solution of gallic acid was prepared using the distilled water in the concentration of 1 mg/ml. Different working standards were prepared to obtain the standard calibration curve, followed by diluting with distilled water with 3 ml and 0.5 ml of FC reagent (1:1) and incubated at room temperature for about 15 min, and then 2 ml

of 7% sodium carbonate was added. Similar steps were followed for estimating phenolic content in the sample extract and the absorbance was measured at 765 nm against blank using spectrophotometer. All experiments were made in triplicates and the TPC was determined using the standard gallic acid calibration curve.

Determination of total flavonoid content (TFC)

TFC of the *C. papaya* sample was determined spectrophotometrically according to the aluminum chloride method with slight modifications [11,12].

A standard solution of quercetin was prepared using methanol in the concentration of 1 mg/1 ml. Different working standards were prepared to obtain a standard calibration curve using 1ml with methanol and 0.5 ml of 1.2% AlCl₃, 0.5 ml of sodium acetate and incubated for 30 min. Similar steps were followed for estimating flavonoid content in the sample extract and absorbance was measured at 415 nm against blank using spectrophotometer. All determinations were made in triplicates and the TFC was determined using the standard quercetin calibration curve.

Antioxidant assay**2,2,1-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity DPPH**

The free-radical scavenging activity was determined by DPPH proposed by Zadeh et al., 2008, with slight modifications [14].

A standard solution of ascorbic acid was prepared in the concentration of 2 mg/ml in methanol. The standard calibration curve was obtained using different working standards and was made up of 3 ml with methanol. DPPH solution (500 µl) was then added and mixed vigorously. Similar steps were followed for the sample extract. The reaction mixture was incubated for about 45 min in dark condition and absorbance was measured at 517 nm using a spectrophotometer. All determinations were made in triplicates and the standard curve was obtained using ascorbic acid.

The % DPPH which was scavenged (% DPPH_{sc}) was calculated using the formula:

$$\text{Scavenging effect (\%)} = 1 - \frac{\text{Absorbance of sample at 517 nm}}{\text{Absorbance of control at 517 nm}} \times 100$$

RESULTS

Qualitative analysis conducted to evaluate the phytochemical profile of *C. papaya* leaves extract.

Phytochemical screening of *C. papaya* leaves extracts shows (Figs. 1-4), the presence of alkaloids, proteins, glycosides, phenol, tannin, saponin, quinine, oxalate, and anthocyanin. The presence of alkaloid, carbohydrate, glycoside, phenols, tannin, saponin, and oxalate shows the greater intensity of their presence in methanolic, ethanolic, hexane, and ethyl acetate extract (Table 1). In the methanolic extract, all the bioactive compounds such as alkaloids, glycosides, phenol, tannin, saponin, quinine, and oxalate are present expect proteins and anthocyanin.

The overall result shows that methanol extracts possess a greater number of bioactive compounds when compared to other solvents.

TPC

The total phenol content was determined by Folin-Ciocalteu method and reported as gallic acid equivalents (GAE) concerning the standard curve. The standard taken was gallic acid in a concentration of 1 mg/ml. The concentration of sample extract was evaluated by comparing it to the standard Graph 1. The concentration of TPC presents in the leaf extract of *C. papaya* in methanol extract is 13.7 mgGAE/g and hexane extract is 13.6 mgGAE/g, ethanol extract is 13.5 mgGAE/g, and ethyl acetate extract is 9.486 mgGAE/g, respectively, to the GAE of the sample [Table 2].

Table 1: Qualitative analysis of *Carica papaya* leaves extract: Phytochemical screening

S.No.	Sample	Alkaloid	Carbohydrate	Amino acid	Glycoside	Phenols tannin	Proteins	Saponin	Quinine	Oxalate	Anthocyanin
1.	Methanol	+	+	+	+	+	-	+	+	+	-
2.	Ethanol	+	+	-	+	+	-	+	+	-	-
3.	Hexane	+	+	-	-	+	-	-	-	+	-
4.	Ethyl acetate	+	+	-	-	-	-	+	-	-	-

Table 2: Quantification of TPC and TFC in different solvent extracts of *Carica papaya* leaves

S. No.	Solvent	TPC	TFC
1.	Methanol	0.083±0.003	0.392±0.006
2.	Hexane	0.078±0.006	0.445±0.025
3.	Ethyl acetate	0.071±0.004	0.422±0.02
4.	Ethanol	0.061±0.002	0.196±0.08

TPC: Total phenolic content, TFC: Total flavonoid contents

TFC

The TFC was determined by aluminum chloride method and represented as quercetin equivalents (QAE) concerning the standard curve. The standard taken was quercetin in a concentration of 1 mg/ml. The concentration of sample extract was evaluated by comparing it to the standard Graph 2.

The concentration of flavonoids presents in the leaf extract of *C. papaya* in methanol extract is 21.60 mgQAE/g, hexane extract is 26.48 mgQAE/g, ethyl acetate extract is 27 mgQAE/g, and ethanol extract is 15.65 mgQAE/g extract, respectively, to the QAE of the sample [Table 2].

Antioxidant activity of *C. papaya*

DPPH assay DPPH radical scavenging activity

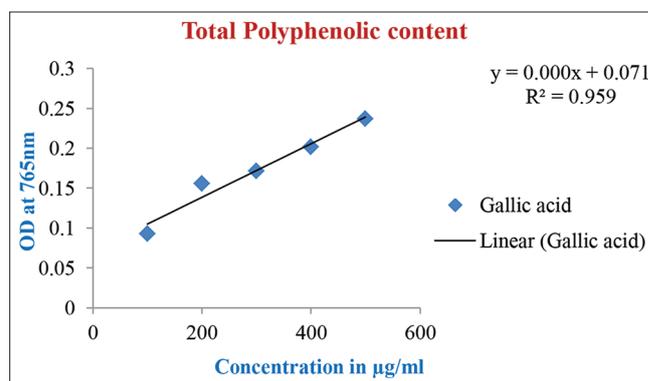
IC₅₀ (Inhibitory Concentration) of the sample extract was determined by comparing it to the standard value of ascorbic acid. The result of the antioxidant activity of *C. papaya* with methanolic extract by DPPH assay shows that the presence of free radicals is more and directly proportional to the concentration of the sample. It means higher the concentration higher will be the percentage of free radicals in the *C. papaya* leaves methanolic extract.

The Graph 3 indicates that the percentage of radical scavenging activity of the methanolic extracts at different concentrations with 50% of DPPH scavenging activity was found to be 80 µg/ml, as the concentration of the sample extract was evaluated compared to the standard value. The results show that methanolic solvent extracts analyzed for the DPPH scavenging activity show the higher radical inhibition activity which can be comparable to standard ascorbic acid.

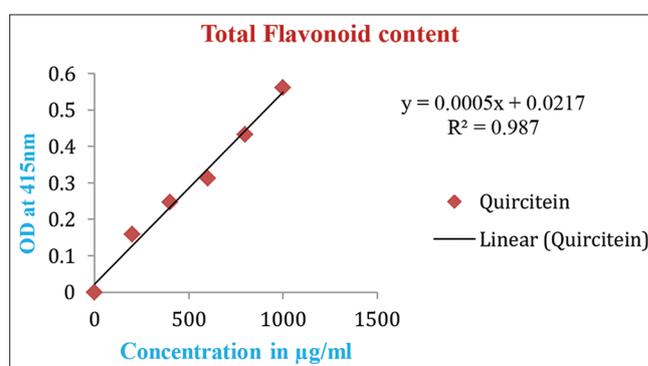
DISCUSSION

Medicinal plants constitutes as an important natural wealth of the country by playing a significant role in primary health of mankind. They importantly serve as raw material for manufacturing medicines as therapeutic drugs [15]. *C. papaya* is used as a natural medicinal plant, recognized for its antimicrobial, anti-amoebic, antifungal, and hypolipidemic activity. Mainly papaya is recognized as "King of medicine," due to the presence of a large number of important phytonutrients loaded in it, and also referred to as "Powerhouse of nutrients" [16,17].

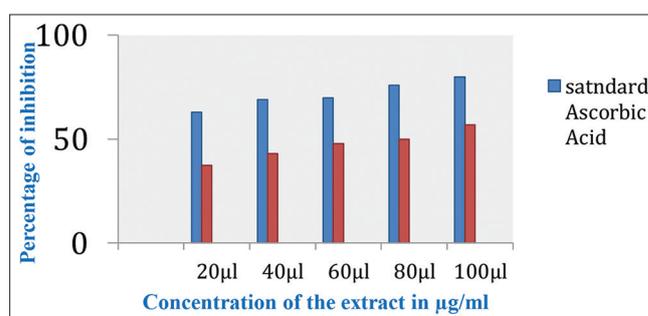
The plant based medicines which have been used in treatment from ancient times reveals in some cases desirable effect were not achieved because the biological action of the herbal medicine of phytoconstituents may vary, as well as the amount of phytoconstituent in a plant can vary according to the age of plant, time of collection, and also environmental conditions [18].



Graph 1: Standard gallic acid calibration curve. (Values are expressed as mean±SE mg of gallic acid equivalent per gram of dry weight, that is, GAE/g of the extract triplicates of each sample extract was recorded)



Graph 2: Standard quercetin calibration curve. (Values are expressed as mean±SE mg of quercetin equivalent per gram of dry weight, that is, QAE/g of the extract triplicates of each sample extract was recorded)



Graph 3: 2,2,1-Diphenyl-1-picrylhydrazyl scavenging activity of methanolic extracts of *Carica papaya* leaves. (Values are expressed as mean±SE mg of ascorbic acid per gram of dry weight, that is, extract triplicates of each sample extract were recorded)

An attempt to study the *in vitro* antioxidant, antimicrobial, and anti-inflammatory of *C. papaya* seed extract with methanolic fractions,



Fig. 1: Phytochemical analysis of ethanol extract



Fig. 3: Phytochemical analysis of hexane extracts



Fig. 2: Phytochemical analysis of ethyl acetate



Fig. 4: Phytochemical analysis of methanol extract

revealed the TPC and the TFC in the methanolic fraction was high compared to other solvent extracts, the antioxidant activity showed the radical scavenging activity of methanolic extracts at different concentration with 50% of scavenging activity, a comparison of the ability of various seed extract have proved their limited DPPH scavenging activity compared to methanol [19].

The antioxidant activity was conducted to compare the total antioxidant activity, TPC, and TFC from the different parts of the papaya tree including their ripe and unripe fruit, seeds, and the young leaves. The result showed that the highest antioxidant activity through β -carotene assay was observed in unripe fruit ($90.67 \pm 0.29\%$), followed by young leaves, ripe fruit, and the seed. In brief, taken to account all the parameter measured, antioxidant was highly remarkable in the sequence of young leaves > unripe fruit > seed [20].

The investigation reports of quantitative analysis of total phenolic and TFCs in extracts of *Oroxylum indicum* L. Kurz. reveal that the TPC and TFC varied among the different solvent extracts used and among them methanolic extracts of seed showed the highest amount compared to other solvent extracts and insists their ratio and distribution in different parts of the tree that could be used for treating various ailments [2].

An increase in the demand for the natural bioactive compounds from the medicinal plants has provided an opportunity for the extraction of either pure or standardized extracts for various therapeutic activities. *C. papaya* leaves are a boon from nature for food as well as therapeutic applications.

CONCLUSION

Bioactive compounds can be studied by extraction and isolation, also with defining their structure and by analyzing it in laboratory models as *in vitro* and *in vivo* studies and importance was given for identification and characterization of the specific phytochemical which is primarily responsible for biological activity.

The current study which was aimed at investigating the presence of biologically active phytochemicals and antioxidant properties of *C. papaya* leaves extract reveals that samples with various solvents such as methanol, ethanol, hexane, and ethyl acetate have shown the presence of phytochemicals constituents such as alkaloid, carbohydrates, and amino acids.

Among the used solvent extracts, the *C. papaya* leaves with methanolic fraction showed the presence of more phytochemicals and have effective phenolic, flavonoid content, and exhibited strongest antioxidant properties which can effectively scavenge reactive oxygen species compared to other solvent extracts used.

Hence, *C. papaya* can be considered as an important and potential natural source for various pharmaceutical and medicinal applications. Interestingly, the broad spectrum of phytochemicals and antioxidants presents in them can be regarded as the reservoir of naturally occurring diverse bioactive molecules and papaya as herbal medicine can be furnished for the quantitative and qualitative extraction for exploring the new promising biomolecules for pharmaceutical applications.

AUTHOR CONTRIBUTION

Nandini G and Kanthesh BM conceptualized the study. Nandini G and Nagalambika Prasad conducted the experiments. Nandini G drafted the Manuscript. Gopenath G and Murugesan Karthikeyan, Ashok Gnanasekaran, Ranjith MS, and Pradeep Palanisamy helped with the Manuscript and Discussion.

DECLARATION AND COMPETING INTEREST

The authors declare that there are no conflicts of interest.

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