

IN VITRO α -AMYLASE AND α -GLUCOSIDASE INHIBITOR ACTIVITY OF *ABUTILON INDICUM* LEAVES

GEETIKA PANT*, J. KRISHNA SAI, S. BABASAHEB, P. RAJASHEKAR REDDY AND SIBI G

Indian Academy Centre for Research and Post Graduate Studies, Bangalore, Karnataka, India

Received: 8 September 2013, Revised and Accepted: 25 September 2013

ABSTRACT

Postprandial hyperglycemia caused by the hydrolysis of carbohydrates by pancreatic α -amylase and intestinal α -glucosidase is a serious condition and inhibition of these enzymes is an effective way in the management of diabetes mellitus. *Abutilon indicum* (L.) Sweet leaves were used to determine their in vitro α -amylase and α -glucosidase inhibitory activity. Various concentrations from 10 $\mu\text{g ml}^{-1}$ to 160 $\mu\text{g ml}^{-1}$ were prepared in methanol along with the standard antidiabetic drug, acarbose. There was a dose dependent percent inhibition by the extract against α -amylase (7.12% - 41.31%) and α -glucosidase (8.01% - 36.13%). Logarithmic regression analysis revealed the IC_{50} of 129.89 $\mu\text{g ml}^{-1}$ (α -amylase) and 137.61 $\mu\text{g ml}^{-1}$ (α -glucosidase) with a potency and preference for α -amylase over α -glucosidase inhibition by the *A. indicum* leaf extract.

Keywords: *Abutilon indicum*, antidiabetic, α -amylase, α -glucosidase

INTRODUCTION

Diabetes mellitus is characterized by increased blood glucose levels resulting from absolute or lack of insulin secretion. Hydrolysis of starch by pancreatic α -amylase and uptake of glucose by intestinal α -glucosidase causes sudden rise in blood glucose levels or hyperglycemia in type 2 diabetes patients. Inhibition of these enzymes is an effective strategy for type 2 disease management [1]. Synthetic hypoglycemic agents are reported to cause gastrointestinal side effects [2, 3] and herbal medicines are getting more importance in the treatment of diabetes as they are free from side effects [4, 5]. Medicinal plants are used in the treatment of diabetes mellitus and were the only options before the discovery of insulin based on traditional practices [6].

Abutilon indicum (L.) Sweet belongs to the family Malvaceae is commonly called as country mallow. The leaves are evergreen, stipulate, fili form, ovate, acuminate, toothed and rarely subtrilobate. Petiole is cylindrical form, yellowish in colour, stellate and hairy. The flowers are yellow in color, peduncle jointed above the middle; pedicels jointed very near to top and the seeds are kidney shaped, tubercled or minutely stellate hairy, black or dark brown in colour [7, 8].

Various part of the plant has been reported to have medicinal properties that include analgesic [9] (Sharma *et al.*, 1989), anti-inflammatory [10, 11], antimicrobial [12, 13, 14], antidiarrhoeal [15], antimalarial [16], hepatoprotective [17, 18], antioxidant [19, 20] and antifertility [21] activities. Leaves of *A. indicum* have reported to possess hypoglycemic [22] and antidiabetic activities [23, 24].

Natural inhibitors of carbohydrate degrading enzymes especially from plant sources offer an attractive strategy for the control of postprandial hyperglycemia. This effort has been directed at investigating α -amylase and α -glucosidase inhibitors from *Abutilon indicum* leaves to reduce the progression of diabetes.

Materials and Methods

Preparation of plant extract

Leaves from *Abutilon indicum* were air-dried, pulverized and extracted with methanol (1:4 w/v) at 30°C for 24 hrs. The extract was filtered, centrifuged and the residue was subjected to subsequent extraction with the same solvent and stored at -20°C.

Phytochemical testing

The extract was subjected to preliminary qualitative tests to identify the phytoconstituents present in the leaves [25, 26].

 α -amylase inhibition assay

The α -amylase inhibitory activity was determined by following the method of Bernfield [27]. 500 μl each of *A. indicum* methanolic leaf extract and 0.02 mol l^{-1} sodium phosphate buffer (pH 6.9 with 0.006 mol l^{-1} NaCl) containing porcine pancreatic α -amylase (0.5 mg ml^{-1}) were incubated at 25°C for 10 minutes. This was followed by addition of 500 μl of starch solution (1%) in 0.02 mol l^{-1} sodium phosphate buffer to the reacting mixture and was incubated at 25°C for 10 min. The reaction was stopped with the addition of 1.0 ml of dinitrosalicylic acid followed by incubating in a boiling water bath for 5 min, and cooled to room temperature. The reaction mixture was then diluted by adding 10 ml of distilled water and absorbance measured at 540 nm in UV-Vis spectrophotometer.

 α -glucosidase inhibition assay

The α -glucosidase inhibitory activity was determined according to the method described by Apostolidis *et al.*, [28]. 50 μl of the extract and 100 μl of yeast α -glucosidase solution were incubated at 25°C for 10 min followed by the addition of 50 μl of 5 mM l^{-1} *p*-nitrophenyl- α -D-glucopyranoside solution in 0.1 mol l^{-1} phosphate buffer (pH 6.9). The reacting mixture was then incubated at 25°C for 5 min and the absorbance was read at 405 nm. Acarbose was used a positive control and the inhibitory activity of α -amylase and α -glucosidase were calculated by using the following formula,

$$\% \text{ Inhibition} = \frac{(\text{Abs}_{\text{Control}} - \text{Abs}_{\text{Sample}}) / \text{Abs}_{\text{Control}} \times 100}$$

The IC_{50} values defined as the concentration of the extract that inhibited 50% of the enzyme activity were determined from plots of percent inhibition versus log inhibitor concentration and calculated by logarithmic regression analysis from the mean inhibitory values.

RESULTS

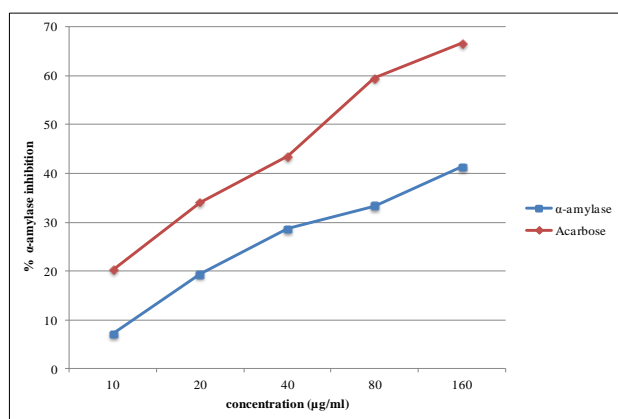
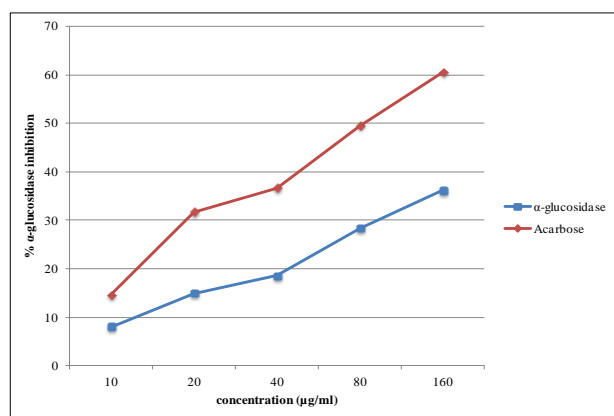
Phytochemical screening of *A. indicum* leaf extracts revealed the presence of alkaloids, flavonoids, phenolics, saponins, steroids, glycosides and carbohydrates (Table 1). Aminoacids, anthraquinones, proteins and tannins were absent in the extract.

Table1: Phytochemical analysis of *A. indicum* leaf extract

Phytoconstituents	Test	Methanolic extract
Alkaloids	Mayer's test	+
Aminoacids	Millon's test	-
Anthraquinones	Borntrager's test	-
Carbohydrates	Molish's test	+
Flavonoids	Ammonia test	+
Glycosides	Keller Kiliani test	+
Phenolics	Ferric chloride test	+
Proteins	Millon's test	-
Saponins	Froth test	+
Steroids	Liebermann-Burchard's test	+
Tannins	Ferric chloride test	-

(+) positive; (-) negative

Increasing concentrations (10, 20, 40, 80 and 160 $\mu\text{g ml}^{-1}$) of *A. indicum* methanolic leaf extracts were prepared for their inhibitory enzyme activities. Figure 1 and 2 shows the % inhibition of α -amylase and α -glucosidase activity of *A. indicum* methanolic leaf extract along with the standard, acarbose. There was a dose-dependent increase in percentage inhibitory activity against α -amylase with 7.12% at a concentration of 10 $\mu\text{g ml}^{-1}$ and 41.31% at 160 $\mu\text{g/ml}$ concentration. In the case of α -glucosidase, 8.01% and 36.13% inhibition was observed at 10 $\mu\text{g ml}^{-1}$ and 160 $\mu\text{g ml}^{-1}$ concentrations. The positive control, acarbose has exerted the highest potent inhibitory action against α -amylase (66.62%) and α -glucosidase (60.62%).

**Fig 1: α -amylase inhibitory activity of *A. indicum* leaf extract****Fig 2: α -glucosidase inhibitory activity of *A. indicum* leaf extract**

Our in vitro studies demonstrated an appreciable α -amylase inhibition with an IC_{50} value of 129.89 $\mu\text{g ml}^{-1}$ and a weak α -glucosidase inhibitory with an IC_{50} value of 137.61 $\mu\text{g ml}^{-1}$ activity of *A. indicum* leaf extract.

DISCUSSION

Many bioactive principles from plants have been reported to have hypoglycemic effect [4, 29] which includes alkaloids [30, 31], flavonoids [32, 33], phenolics [34], triterpenoids [35] and carbohydrates [36]. In this study, alkaloids, flavonoids, phenolics, steroids and carbohydrates were identified from the *A. indicum* methanolic leaf extracts. A number of other phytochemicals have been identified from the leaves of *A. indicum* in earlier studies [37, 38, 24]. There is a positive correlation between plant phenolics and antidiabetic activities [39] and the presence of phenolics in *A. indicum* leaves might have influenced its hypoglycemic activity.

α -amylase catalyzes the hydrolysis of starch and α -glucosidase catalyzes the final step in carbohydrate digestion which leads to postprandial hyperglycemia. Inhibitors of α -amylase and α -glucosidase are useful in the control of hyperglycemia as they delay carbohydrate digestion and causing reduced glucose absorption rate which consequently reduce the postprandial plasma glucose rise [40]. These inhibitors have been found useful in the control of diabetes mellitus over many years [41, 42]. Many scientists have investigated the plants containing various phytochemicals that exhibit additive and synergistic interaction in antidiabetic properties which exert positive health-promoting effects [43]. In this present study, in vitro α -amylase and α -glucosidase inhibitor activity of methanolic extract of *Abutilon indicum* (L.) Sweet leaves were evaluated. The retardation and delay of carbohydrate absorption with a plant-based α -amylase and α -glucosidase inhibitor offers a prospective therapeutic approach for the management of type 2 diabetes mellitus. The IC_{50} values show that *A. indicum* has a potency and preference for α -amylase over α -glucosidase inhibition. A significant inhibitory activity was observed against α -amylase (7.12 - 41.31 $\mu\text{g ml}^{-1}$) at different concentrations than the α -glucosidase inhibition (8.01 - 36.13 $\mu\text{g ml}^{-1}$). The different inhibition kinetics observed between α -amylase and α -glucosidase could be due to structural differences related to the origins of the enzymes [44].

CONCLUSION

This study justifies the hypoglycemic activity of *Abutilon indicum* leaves and can be used in the management of diabetes. However, the active principles responsible for inhibitory action of α -amylase and α -glucosidase need to be identified and characterized for the development of indigenous botanical resources for novel hypoglycemic drug development.

REFERENCES

1. Kwon YI, Apostolidis E, Kim YC, Shetty K. Health benefits of traditional corn, beans and pumpkin: In vitro studies for hyperglycemia and hypertension management. *J Med Food* 2007; 10: 266 - 275.
2. Cheng AYY, Fantus IG, Oral antihyperglycemic therapy for type 2 diabetes mellitus. *Canadian Medicinal Association Journal*. 2005; 172(2): 213-226.
3. Hanefeld M, The role of acarbose in the treatment of non-insulin-dependent diabetes mellitus. *J Diabetes Complications*. 1998; 12:228-237.
4. Grover JK, Yadav S, Vats V, Medicinal plants of India with anti-diabetic potential. *J Ethnopharmacol*. 2002; 81: 81-100.
5. Mukherjee PK, Maiti K, Mukherjee K, Houghton PJ, Leads from Indian medicinal plants with hypoglycemic potentials. *J Ethnopharmacol*. 2006; 106(1): 1-28.
6. Ribnicky DM, Poulev A, Watford M, Cefalu WT, Raskin I, Antihyperglycemic activity of Tarralin, an ethanolic extract of *Artemisia dracunculul* L. *Phytomedicine*. 2006; 13: 550-557.
7. Kirtikar KR, Basu BD. 1994. *Indian Medicinal Plants*, 2nd Edn, Vol. I, Dehradun. 314-317.
8. Prajapati ND, Purohit SS, Sharma AK, Kumar TA, *Handbook of Medicinal Plants*, Agrobios, (India) Jodhpur. 2003; 3.
9. Sharma PV, Ahmed ZA, Sharma VV, Analgesic constituent of *Abutilon indicum*. *Indian drugs*. 1989; 26 (7): 333.
10. Nadkarni KM, 1954. *The Indian Materia Medica* 3rd Edn., Popular Prakashan, Bombay, Vol. 1.

11. Tripathi P, Chauhan NS, Patel JR, Anti-inflammatory effect of *Abutilon indicum* extract. Nat Prod Res. 2012; 26(17): 1659-1651.
12. Mehta BK, Neogi R, Kotra S, Mall OP, Antimicrobial activity of *Abutilon indicum*. Fitoterapia, 1997; 68(3): 273-274.
13. Pandey DK, Tripathy RD, Tripathy NN, Dixit SN, Antifungal activity of some seed extracts with special reference to that of *Pimpinella diversifolia* DC. Indian Journal of Crude Drug Research. 1983; 21(4): 177-182.
14. Vairavasundaram RP, Senthil K, Antimycotic activity of the components of *Abutilon indicum* (Malvaceae), Drug Invention Today. 2009; 1(2): 137-139.
15. Chandrashekhar VM, Nagappa AN, Channesh TS, Habbu PV, Rao KP, Antidiarrhoeal activity of *Abutilon indicum* leaf extracts. Journal of Natural Remedies. 2004; 4(1): 12-16.
16. Rahuman AA, Gopalakrishnan G, Venkatesan P, Kannappan G, Isolation and identification of mosquito larvicidal compound from *Abutilon indicum* (Linn.) Sweet. Parasitol Research. 2008; 102: 981-988.
17. Porchezian E, Ansari SH, Hepatoprotective activity of *Abutilon indicum* on experimental liver damage in rats. Phytomedicine. 2005; 12(1-2): 62-64.
18. Dash GK, Samanta A, Kanungo SK, Shau SK, Suresh P, Ganpathy S, Hepatoprotective activity of leaves of *Abutilon indicum*. Indian Journal of Natural Products. 2000; 16(2): 25-27.
19. Srividya AR, Dhanabal SP, Jeevita S, Vishnuvarthan VJ, Rajeshkumar R, Relationship between Antioxidant Properties and Chemical Composition of *Abutilon indicum* Linn. Indian J Pharm Sci. 2012; 74(2): 163-167.
20. Yasmin S, Kashmiri MA, Asghar MN, Ahmad M, Mohy-ud-Din A, Antioxidant potential and radical scavenging effects of various extracts from *Abutilon indicum* and *Abutilon muticum*. Pharm. Biol. 2010, 48(3): 282-289.
21. Johri RK, Pahwa GS, Sharma SC, Zutshi U, Determination of estrogenic/ antiestrogenic potential of antifertility substances using rat uterine peroxidase assay. Contraception. 1991; 44(5): 549-557.
22. Seetharam YN, Chalageri G, Setty SR, Bheemachar, Hypoglycemic activity of *Abutilon indicum* leaf extracts in rats. Fitoterapia. 2002; 73 (2): 156-159.
23. Krisanapun C, Lee SH, Peungvicha P, Temsiririrkkul R, Baek SJ, Antidiabetic activities of *Abutilon indicum* (L.) Sweet are mediated by enhancement of adipocyte differentiation and activation of the GLUT1 Promoter. Evid Based Complement Alternat. Med. 2011; 167684.
24. Krisanapun C, Peungvicha P, Temsiririrkkul R, Wongkrajang Y, Aqueous extract of *Abutilon indicum* Sweet inhibits glucose absorption and stimulates insulin secretion in rodents, Nutr. Res. 2009; 29(8): 579-587.
25. Trease GE, Evans WC, Pharmacognosy, 15th ed. 2002; Springer, Berlin.
26. Sofowora A, Screening plants for bioactive agents. In: Medicinal Plants and Traditional Medicinal in Africa, 2nd ed., Spectrum Books Ltd., Sunshine House, Ibadan, Nigeria, 1993; pp. 134-156.
27. Bernfield P, Enzymes of starch degradation and synthesis. Adv Enzymol. 1951; 12: 379 - 380.
28. Apostolidis E, Kwon YI, Shetty K, Inhibitory potential of herb, fruit, and fungus enriched cheese against key enzymes linked to type 2 diabetes and hypertension. Inn Food Sci Emerg Technol. 2007; 8: 46-54.
29. Oboh G, Ademiluyi AO, Faloye YM, Effect of combination on the antioxidant and inhibitory properties of tropical pepper varieties against α -amylase and α -glucosidase activities in vitro. J Med Food. 2011; 14:1152-1158.
30. Dineshkumar B, Mitra A, Mahadevappa M, Antidiabetic and hypolipidemic effects of mahanimbine (carbazole alkaloid) from *Murraya koenigii* (rutaceae) leaves. Int J Phytomed. 2010; 13:22-30.
31. Sharma B, Salunke R, Balomajumder C, Daniel S, Roy P, Anti-diabetic potential of alkaloid rich fraction from *Capparis decidua* on diabetic mice. J Ethnopharmacol. 2010; 13:457-462.
32. Panda S, Kar A, Apigenin (4',5,7-trihydroxyflavone) regulates hyperglycaemia, thyroid dysfunction and lipid peroxidation in alloxan-induced diabetic mice. J Pharm Pharmacol. 2007; 13:1543-1548.
33. Yoshikawa M, Wang T, Morikawa T, Xie H, Matsuda H, Bioactive Constituents from Chinese Natural Medicines. XXIV. Hypoglycemic Effects of *Sinocrassula indica* in Sugar-Loaded Rats and Genetically Diabetic KK-Ay Mice and Structures of New Acylated Flavonol Glycosides, Sinocrassosides A1, A2, B1, and B2. Chem Pharm Bull. 2007; 55:1308-1315.
34. Manickam M, Ramanathan M, Jahromi MA, Chansouria JP, Ray AB, Antihyperglycemic activity of phenolics from *Pterocarpus marsupium*. J Nat Prod. 1997; 13: 609-610.
35. Tan MJ, Ye JM, Turner N, Hohnen-Behrens C, Ke CQ, Tang CP, et al, Antidiabetic activities of triterpenoids isolated from bitter melon associated with activation of the AMPK pathway. Chem Biol. 2008; 13:263-273.
36. Contreras C, Roman R, Perez C, Alarcon F, Zavala M, Perez S, Hypoglycemic activity of a new carbohydrate isolated from the roots of *Psacalium peltatum*. Chem Pharm Bull. 2005; 13:1408-1410.
37. Lakshmayya, Nelluri NR, Kumar P, Agarwal NK, Gouda TS, Setty SR, Phytochemical and pharmacological evaluation of leaves of *Abutilon indicum*. Indian Journal of Traditional Knowledge. 2003; 2(1): 79-83.
38. Ramasubramanaraja R, Pharmacognostical phytochemical including GC-MS investigation of ethanolic leaf extracts of *Abutilon indicum* (Linn), Asian Journal of Pharm. Ana. 2011; 1(4): 88-92.
39. Anam K, Widharna RM, Kusriani D, α -glucosidase inhibitor activity of *Terminalia* species. Int J Pharmacol, 2009; 5: 277-280.
40. Tarling CA, Woods K, Zhang R, Brastianos HC, Brayer GD, Andersen RJ, et al, The Search for Novel Human Pancreatic α -Amylase Inhibitors: High-Throughput Screening of Terrestrial and Marine Natural Product Extracts. Chem BioChem. 2008; 9: 433-438.
41. Layer P, Rizza RA, Zinsmeister AR, Carlson GL, DiMaggio EP, Effect of a purified amylase inhibitor on carbohydrate tolerance in normal subjects and patients with diabetes mellitus. Mayo Clin Proc. 1986; 61(6): 442-447.
42. Tundis R, Loizzo MR, Menichini F, Natural products as α -amylase and α -glucosidase inhibitors and their hypoglycemic potential in the treatment of diabetes: an update. Mini Rev Med Chem. 2010; 10(4):315-331.
43. Samad A, Shams MS, Ullah Z, Wais M, Nazish I, Sultana Y, et al, Status of herbal medicines in the treatment of diabetes: a review. Current Diabetes Reviews. 2009; 5(2):102-111.
44. Chiba S, Molecular mechanism in α -glucosidase and glucoamylase. Biosci Biotechnol Biochem. 1997; 61:1233-1239