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Research Article

ASSOCIATION BETWEEN MATRIX METALLOPROTEINASE-1 –1607 1G/2G POLYMORPHISM AND CHRONIC PERIODONTITIS IN INDONESIAN POPULATION

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ABSTRACT

Objective: This study aimed to identify the distribution of matrix metalloproteinase (MMP)- $1 - 1607 \ 1G/2G$ alleles and genotypes in subjects with chronic periodontitis and healthy subjects in a sample of Indonesian population and assess the possible association of this polymorphism with susceptibility to chronic periodontitis.

Methods: Genomic DNA samples were obtained from 200 Indonesian males aged 33–78 years old, comprising 100 chronic periodontitis patients and 100 healthy controls. DNA fragments were amplified by a polymerase chain reaction and analyzed by restriction fragment length polymorphism. Results were analyzed by Chi-square test.

Results: The frequency of the 2G allele was high both in subjects with periodontitis (87%) and in controls (91%). Analysis of MMP-1 genotype (–1607 1G/2G) showed no significant difference between the chronic periodontitis and healthy groups (p>0.05).

Conclusion: The result found no association between MMP-1 –1607 1G/2G polymorphism and susceptibility to chronic periodontitis in Indonesian subjects.

Keywords: Periodontitis, Matrix metalloproteinases-1, Genetic polymorphism, Indonesia.

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INTRODUCTION

Periodontitis is an inflammatory disease of the supporting dental tissue that is initiated by specific microorganisms or groups of specific microorganisms, ultimately causing periodontal apparatus destruction by means of loss of clinical attachment (CAL) [1-4]. In addition, we have investigated the prevalence of periodontal diseases in Indonesia, finding that chronic periodontitis was the most common form of periodontal disease at 75.2%; a weak positive correlation was found between age and chronic periodontitis [5]. Taken together, these findings suggest that genetics might contribute to susceptibility to periodontal disease.

Matrix metalloproteinases (MMPs) are a family of proteolytic enzymes that play a major role in periodontal tissue degradation. At present, about 28 types of MMPs have been identified, some of which are found in inflamed periodontal tissues and gingival crevicular fluid [6]. Studies showed that the 2G allele significantly increases the transcription activity of MMP-1 compared to the 1G allele [6]. However, the results are inconsistent across studies. Variation in the disease, sample selection, race, and ethnic background may all contribute to the inconsistent findings across the previous studies. Therefore, further study is needed to examine the distribution of MMP-1 genotypes in other populations to confirm if it is indeed a genetic marker of chronic periodontitis.

The present study aimed to identify the distribution of MMP-1 alleles and genotypes in Indonesian subjects with chronic periodontitis and healthy controls and also to identify the possible association of the MMP-1 promoter polymorphism with susceptibility to chronic periodontitis.

METHODS

Subjects and sampling

Two-hundred Indonesian male adults (aged 33–78 years old) participated in this study, including 100 subjects with chronic periodontitis and 100 subjects with healthy periodontal conditions as controls recruited through the dental faculty hospital (RSGM-P FKG UI). Periodontal examinations were performed by a periodontal specialist resident and then standardized [7]. Patients who had probing depth >3 mm, clinical attachment loss >5 mm, and/or bleeding on probing positive were selected as chronic periodontitis subjects. Dental and/or panoramic radiographs were used to confirm the diagnosis of chronic periodontitis. Patients with systemic diseases and a history of antibiotic use within the past 6 months were excluded from this study. Peripheral blood samples were obtained from direct venipuncture, transferred to ethylenediaminetetraacetic acid (EDTA) tubes, and stored at -20° C until DNA extraction using the technique by Auerkari *et al.* [8,9]. This study was approved by the Ethics Committee of the Faculty of Dentistry, University of Indonesia.

Polymerase chain reaction (PCR)

MMP-1 genotype was determined using specific forward and reverse primers for 1G/2G alleles. The sequences of the PCR primers were forward 5'-TCGTGAGAATGTCTTCCCATT-3' and reverse 5'-TCTTGGATTGATTTGAGATAAGTGAAATC-3'. The target size of the PCR product is 118 bp. These primer pairs have been used in the previous studies [8,9]. PCR was carried out in a total volume of 20 μ L containing 0.75 μ L genomic DNA, 0.75 μ L of each primer (PT. Genetika Science Indonesia, Jakarta, Indonesia), 7.75 μ L ddH₂O, and 10 μ L PCR ready mix (KAPA Biosystems, Massachusetts, K1006, USA). The PCR cycling conditions were 1 min at 95°C, followed by 35 cycles of 30 s at 95°C, 30 s at 55°C, and 30 s at 72°C, with a final extension of 72°C for 5 min.

Restriction fragment length polymorphism (RFLP)

This study used restriction enzyme XmnI to digest the 1G alleles, while 2G alleles do not contain the recognition site due to the guanine (G) insertion [4,10]. A total volume of 18 μ L RFLP mixture contained 10 μ L PCR product, 1 μ L XmnI (1000 U/mL, CutSmart^M, New England Biolabs, Inc.), 2 μ L nuclear exact buffer, and 5 μ L ddH₂O. The mixtures were incubated at 37°C overnight. Restriction enzyme XmnI digested 1G allele and created two fragments of 89 bp and 29 bp, but it could not digest 2G allele due to the insertion of the guanine base. This single-nucleotide polymorphism (SNP) caused mismatches in the reverse primer annealed sequence for the XmnI site.

Gel electrophoresis

This study used 1.5% agarose gel (TopVision Agarose, Thermo Fisher Scientific Inc., Massachusetts, R0491, USA) for PCR product and 3% for RFLP product, dissolved in 1x TAE buffer (0.089 M Tris, 0.089 M acetic acid, and 0.5 M EDTA pH 8.0). 1.5 μ L of GelRedTM was added as a dye for the DNA fragments. As a DNA marker, 3 μ L 50 bp DNA ladder (InvitrogenTM, Thermo Fisher Scientific Inc., Massachusetts, 10416014, USA) was added to one well. Electrophoresis was performed at 80 V and 400 mA for 45 min for PCR products.

Statistical analysis

Statistical analysis was performed using the SPSS 17.0 statistical package (SPSS Inc., Chicago, IL, USA). Differences in allele and genotype frequencies between the periodontitis and control groups were assessed by Chi-square test. p<0.05 was considered statistically significant. The risk associated with individual alleles and genotypes was calculated as the odds ratio (OR) with 95% confidence interval (95% CI).

RESULTS

All PCR products were detected and visualized by electrophoresis at 118 bp (Figs. 1 and 2). Thus, 1G/1G homozygotes showed a single 89 bp band, 2G/2G homozygotes displayed a single 118 bp band, and 1G/2G heterozygotes exhibited two bands of 118 bp and 89 bp. The polymorphism was in Hardy–Weinberg equilibrium.

The allele and genotype frequencies are presented in Tables 1 and 2. Table 1 shows that the 2G allele was most common in both the chronic periodontitis (87%) and control subjects (91%). The presence of the 2G allele was not related to increased risk of chronic periodontitis (p=0.201, OR=0.662, 95% CI=0.350–1.250). Table 2 shows that the 2G/2G genotype occurred in 86% of chronic periodontitis subjects, which is slightly lower than in control subjects (87%). This genotype was not significantly associated with increased risk of chronic periodontitis (p=0.076, OR=0.386, 95% CI=0.131–1.140).

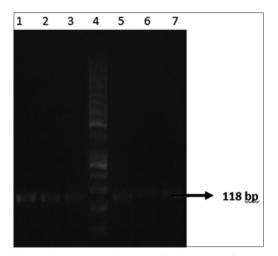


Fig. 1: Electrophoresis visualization of polymerase chain reaction products. Single bands were seen at 118 bp (lanes 1–3, 5–7) and the size marker (lane 4) is a 50 bp ladder

DISCUSSION

Periodontitis is a multifactorial disease; aside from interaction between periodontal pathogens and host immunoinflammatory response, genetic factors may be associated with the pathogenesis of periodontal disease [2,3]. The loss of clinical attachment (CAL) in periodontitis is caused by the destruction of periodontal tissues initiated by group(s) of Gram-negative and anaerobic bacteria that colonize the subgingival [4]. The host responds to this situation by activating immune cells and releasing inflammatory cytokines and enzymes. By contrast, chronic periodontitis is the slow, progressive destruction of the periodontal apparatus and is more likely related to the accumulation of dental plaque [1].

It is known that MMP-1, also known as collagenase, is an MMP family that can cleave interstitial collagen types I and III, which are the predominant types of interstitial collagen in the periodontium, and it is found at higher

Table 1: Distribution of matrix metalloproteinases-1 alleles in control and chronic periodontitis subjects

Alleles	Control		Chronic periodontitis		p-value	Odds ratio (95% CI)
	n	%	n	%		
1G	18	9.0	26	13.0	0.201	
2G	182	91.0	174	87.0		0.662 (CI=0.350-1.250)

CI: Confidence interval

Table 2: Distribution of matrix metalloproteinases-1 genotypes in control and chronic periodontitis subjects

Genotypes	Control		Chronic periodontitis		p-value	Odds ratio (95% CI)
	n	%	n	%		
1G/1G	5	5.0	12	12.0	0.076	
1G/2G	8	8.0	2	2.0		
2G/2G	87	87.0	86	86.0		0.386 (CI= 0.131-1.140)

CI: Confidence interval

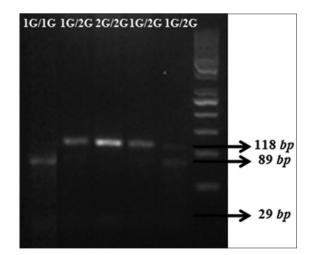


Fig. 2: Electrophoresis visualization of matrix metalloproteinases-1 –1607 1G/2G polymorphisms performed by XmnI. XmnI digested the 1G allele and created two fragments of 89 bp and 29 bp. 1G/1G homozygotes appeared as two bands of 89 bp and 29 bp, 2G/2G homozygotes appeared as a single 118 bp band, and 1G/2G heterozygotes appeared as three bands of 118 bp, 89 bp, and 29 bp. The size marker is a 50 bp ladder levels in periodontitis sites [4,6]. MMP-1 is secreted by connective tissue cells and macrophages. Genetic variation in the promoter region of the MMP-1 gene can influence transcription levels of MMP-1, thereby altering susceptibility to and severity of periodontitis [4]. With respect to an SNP located at -1607 bp, where there is a guanne (G) base insertion, creates a mismatched Ets-binding site. A study of the Chinese population showed that the 2G allele may be associated with aggressive periodontitis [4], while a study of the Brazilian population concluded that it was not [10]. This contradiction may occur because polymorphisms, especially SNPs, are population specific [11].

To the best of our knowledge, this is the first study of MMP-1 polymorphism and periodontitis risk in Indonesian population. Our findings did not suggest a relationship between MMP-1 genotype distribution and risk of chronic periodontitis, which is consistent with findings reported by both Astolfi *et al.* (2006) and Itagaki *et al.* (2004) [10,12-14].

However, our study is limited by the sample size of 200 males. Given the complex relationship between genetic polymorphisms and periodontal destruction, further study with a larger sample is needed to confirm this finding.

CONCLUSION

No association between MMP-1 -1607 1G/2G polymorphism and susceptibility to chronic periodontitis was found in Indonesian subjects. However, further studies need to be conducted.

ACKNOWLEDGMENTS

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CONFLICTS OF INTEREST

All authors have none to declare.

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