

## FABRICATION OF DRUG DELIVERY SYSTEM FOR CONTROLLED RELEASE OF CURCUMIN, INTERCALATED WITH MAGNETITE NANOPARTICLES THROUGH SODIUM ALGINATE/POLYVINYLPIRROLIDONE-CO-VINYL ACETATE SEMI IPN MICROBEADS

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Received: 05 Apr 2020, Revised and Accepted: 13 Jul 2020

### ABSTRACT

**Objective:** The aim of the present work is to fabricate curcumin (CUR) encapsulated microbeads in the polymer matrix of sodium alginate (SA)/poly(vinylpyrrolidone)-co-vinyl acetate (PVP-co-VAc) intercalated with magnetite nanoparticles (MNPs) using glutaraldehyde (GA)/calcium chloride CaCl<sub>2</sub> as the crosslinker.

**Methods:** Magnetite nanoparticles (MNPs) were synthesized by a modified co-precipitation method. Curcumin encapsulated SA/PVP-co-VAc microbeads, intercalated with MNPs were prepared by simple ionotropic gelation technique. The formation of microbeads and uniform distribution of curcumin were characterized using spectroscopic methods. In addition, swelling and drug release kinetic studies of the microbeads were performed in simulated intestinal fluid (pH 7.4) and simulated gastric fluid (pH 1.2) at 37 °C.

**Results:** Microbeads formation was confirmed by Fourier Transform Infrared (FTIR). Differential Scanning Calorimetry (DSC) studies reveal that the peak at 181 °C of CUR was not observed in CUR loaded microbeads, which confirms that CUR was encapsulated at the molecular level in the polymer matrix. The X-Ray diffraction (X-RD) diffractograms of CUR shows 2 $\theta$  peaks between 12-28 °, which indicated the crystalline nature of CUR, these peaks are not found in CUR loaded microbeads, suggesting that the drug has been molecularly dispersed in the polymer matrix. The X-RD 2 $\theta$  peaks of MNPs are observed in the MNPs loaded microbeads, which confirms that MNPs are successfully loaded in the microbeads. The swelling studies and *in vitro* release studies were performed at pH 1.2 and 7.4. The results reveal that at pH 7.4 highest swelling and release was observed, which confirms that the developed microbeads are pH sensitive and are suitable for intestinal drug delivery. The drug release kinetics fit into the Korsmeyer-Peppas equation, indicating non-Fickian diffusion.

**Conclusion:** The results concluded that the present system as dependent on pH of the test medium and hence suggest suitability for intestinal drug delivery.

**Keywords:** Sodium alginate, Poly(vinylpyrrolidone)-co-vinyl acetate, Magnetite nanoparticles, Curcumin, Microbeads

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### INTRODUCTION

The best acceptable route for drug administration is an oral route, due to ease of administration and gastrointestinal physiology provides additional flexibility in the design of dosage form compared to other routes [1]. Normally, conventional oral drug administration does not provide a controlled release or target specificity [2]. Further, it has several drawbacks such as poor patient compliance, frequent dosing, poor bioavailability etc. To overcome these problems novel drug delivery systems (NDDS) such as controlled/sustained drug release systems with IPNs (Interpenetrating polymer networks) have been evolved [3, 6]. Polymers play a vital role in the development of controlled drug delivery systems because of their favourable and flexible features such as biocompatibility, non-toxic, biodegradability and also it can be easily produced at industrial scale [4].

Sodium alginate (SA) is anionic polysaccharide, composed of two different kinds of hexuronic acid residues such as  $\beta$ -D-mannuronic acid (M) and  $\alpha$ -L-guluronic acid (G) arranged as random or in an alternating manner [5, 7]. It has several biomedical and biotechnological applications due to its specific properties such as hydrophilicity, biodegradability, biocompatibility and non-toxicity [8-10]

Poly(vinylpyrrolidone)-co-vinyl acetate (PVP-co-VAc) is a water soluble block copolymer consisting of both hydrophobic and hydrophilic components in its structure, which facilitates the increased solubilisation of drugs [11]. The presence of hydrophobic part in PVP-co-VAc, acts as a good drug carrier for hydrophobic or less water-soluble drugs. The applications of PVP-co-VAc in biomedical applications are scanty. Previously Bailly *et al.*, [12] reported that Poly(N Vinylpyrrolidone)-block-poly(vinyl acetate) (PVP-b-PVAc) is a good interesting candidate for the delivery of hydrophobic drug (clofazimine).

Curcumin (CUR), a polyphenol of turmeric (diferuloylmethane), a natural bioactive compound, is obtained from *Curcuma longa* [13]. It has anti-inflammatory, anti-oxidant, antimicrobial, antispasmodic and antiproliferative activity against different cancer cells [14-16]. However, its biomedical applications are very scanty due to low aqueous solubility, limited bioavailability and potentially lipophilic characteristics [17], which have limited *in vivo* efficacy of curcumin [18]. To overcome this problem, various types of micro and nanocarriers have been proposed, such as micelles, nanoparticles, and iron oxide particles [19].

Magnetite nanoparticles (MNPs) are increasingly being considered for a number of biomedical applications due to their inherent superparamagnetic properties, ultra-fine size and biocompatibility [20-22]. The functional properties of the MNPs can be tailored for specific biological functions, such as drug delivery [23], hyperthermia or magnetic targeting [24-26], magnetic resonance imaging (MRI) [27-29]. Among the MNPs, iron oxide nanoparticles (magnetite  $\gamma$ -Fe<sub>2</sub>O<sub>3</sub> or magnetite Fe<sub>3</sub>O<sub>4</sub>) are the most popular formulations. The large surface to volume ratio of MNPs renders relative high loading in biocompatible materials [28]. Owing to nanosize, MNPs can accumulate in tumor cells by the enhanced permeability and retention (EPR) effect [30]. Further malignant tumors have high heat sensitivity than normal tissues in the temperature range of 41-47 °C and the property is exploited for chemotherapeutic applications [31]. MNP drug delivery systems have incorporated small traditional molecules like paclitaxel, doxorubicin and methotrexate [22]. Natural macromolecules like curcumin have not been researched earlier with MNP combination for cancer therapy.

In the current work, SA/PVP-co-VAc microbeads were prepared by a simple gelation technique. The microbeads were characterized using fourier transform infrared spectroscopy, differential scanning

calorimetry, X-ray diffraction and scanning electron microscopy. In addition, swelling studies and drug release kinetics of the microbeads have been reported. The main objective of the present work is to encapsulate the curcumin molecules intercalated with magnetite nanoparticles into SA/PVP-co-VAc microbeads and to study the effect of MNPs on controlled drug delivery of curcumin for novel chemotherapeutic applications.

## MATERIALS AND METHODS

### Materials

Poly (vinylpyrrolidone)-co-vinylacetate, ferrous chloride ( $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$ ), and tetramethylammonium hydroxide were purchased from Sigma-Aldrich (USA). Sodium alginate, glutaraldehyde (GA), ferric chloride ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ), and calcium chloride were purchased from Sd. Fine chemicals, Mumbai, India. Curcumin was purchased from Loba Chemicals, Mumbai, India. Water used was of high purity grade after double distillation.

### Methods

#### Synthesis of magnetite nanoparticles (MNPs)

Magnetite nanoparticles (MNPs) were synthesized by modified co-precipitation method [32] using ferrous and ferric salts. 5.2 g of  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$  and 2.0 g of  $\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$  were weighed and transferred

into 250 ml round bottom flask and 100 ml of deoxygenated distilled water was added and stirred at 300 rpm for 2 h under nitrogen atmosphere. Then ammonium hydroxide solution was added slowly to the reaction mixture to raise the pH to 8.0. Then the solution turned from brown to black colour, which indicates the formation of magnetite nanoparticles. The magnetite nanoparticles formed were separated and washed with distilled water followed by 1 ml of tetramethylammonium hydroxide and then dried in air. Finally the developed MNPs were stored in airtight containers.

#### Synthesis of SA/PVP-co-VAc/MNPs microbeads

The magnetite nanoparticles loaded microbeads were prepared using simple ionotropic gelation technique. Exactly a weighed amount of magnetite nanoparticles (table 1) were added to 5 ml of double-distilled water and placed for sonication for 5 min. The resulting solution was gradually added to a blend ratio of SA/PVP-co-VAc (table 1), followed by addition of drug and GA (table 1) and stirred for 3 h.

The resulting solution is placed for sonication for 10 min to get a homogeneous solution. The resulting homogeneous suspension was added dropwise into  $\text{CaCl}_2$  solution under constant stirring at room temperature; the spherical beads formed instantly were kept aside for 20 min. The obtained wet beads were collected by decantation, washed with double distilled water to remove the drug attached on the bead surface, and finally were dried in air overnight at room temperature.

Table 1: Formulation and composition of all samples

Formulation code	Polymers (%w/v)		MNPs (mg)	Drug (mg)	GA (ml)
	SA	PVP-co-VAc			
F1	80	20	20	100	1
F2	60	40	20	100	1
F3	40	60	20	100	1
F4	80	20	20	150	1
F5	80	20	20	200	1
F6	80	20	20	100	2.5
F7	80	20	20	100	5
F8	80	20	00	00	1

(Results are expressed as mean $\pm$ SD, n=3)

### Characterization methods

#### Fourier transform infrared (FTIR) spectral analysis

Fourier Transmission infrared spectra of CUR, MNPs, placebo microbeads, and drug loaded microbeads were recorded using FTIR spectrophotometer (model Bomem MB-3000, with Horizon MBTM FTIR software) in the wavelength range of 400-4000  $\text{cm}^{-1}$  to find out the possible chemical interactions between polymers and drug.

#### Differential scanning calorimetry (DSC)

DSC curves of CUR, MNPs, placebo microbeads, and drug-loaded microbeads were recorded using thermogravimetric analyzer Rheometric Scientific, Model DSC-SP, UK. The analysis was performed by heating each sample from 40 °C to 600 °C at the heating rate of 10 °C/min under nitrogen atmosphere.

#### Thermogravimetric analysis (TGA)

Thermogravimetric analysis of CUR, MNPs, placebo microbeads, and drug-loaded microbeads were carried out using thermogravimetric analyzer Rheometric Scientific, Model DSC-SP, UK. About 10-12 mg of sample was placed into alumina crucible and the thermograms were recorded between 40 °C to 600 °C at a heating rate of 10 °C/min under nitrogen atmosphere.

#### X-Ray diffraction (X-RD) analysis

The X-ray diffraction of CUR, placebo microbeads, drug-loaded microbeads, MNPs and MNPs/drug-loaded microbeads were performed by a wide-angle X-ray scattering diffractometer (Panalytical X-ray Diffractometer, model-X'pert Pro) with Cu-K  $\alpha$  radiation ( $\lambda = 1.54060$ ) at a scanning rate of 10°/min to determine the crystallinity.

#### Scanning electron microscopy (SEM) analysis

The morphological characterization of microbeads and MNPs was observed using SEM (JOEL MODEL JSM 840A) with an accelerated voltage of 20 kV.

#### Swelling measurements

The swelling behavior of different formulations was determined gravimetrically under both pH 1.2 and pH7.4.

#### Encapsulation efficiency

The percentage encapsulation efficiency of CUR loaded microbeads was estimated according to the formula and method reported in previous literature [30]. A known mass of drug-loaded microbeads (20 mg) was immersed into 100 ml of phosphate buffer solution (pH 7.4 containing 5 % absolute ethyl alcohol) for 24 h and then vigorously stirred to ensure the complete extraction of CUR from the microbeads. Supernatants were filtered and analyzed by ultraviolet (UV) spectrophotometer (LabIndia, Mumbai, India) at the  $\lambda_{\text{max}}$  of 470 nm with placebo microbeads as blank correction. Concentration of drug was determined using calibration curve constructed by series of CUR standard solutions and the percentage of encapsulation efficiency was determined using the following formula.

$$\text{Encapsulation efficiency percentage (EE \%)} = \frac{W_t}{W_i} \times 100$$

Where  $W_t$  is the total amount of drug in the microspheres and  $W_i$  is the total quantity of drug added initially during the preparation.

#### In vitro drug release studies

In vitro drug release studies for different formulations were performed using a dissolution tester containing eight baskets at 37 °C in 900 ml of phosphate buffer solution (pH-7.4) at a rotation speed of 50 rpm

to replicate intestinal atmosphere. The accurately weighed 100 mg of microbeads were used for the study. At regular intervals of time, 5 ml aliquot samples were withdrawn and analyzed using UV

spectrophotometer at fixed  $\lambda_{\max}$  value of 470 nm, and the released drug amount was obtained using concentration versus absorbance calibration curve.

## RESULTS AND DISCUSSION

### FTIR analysis

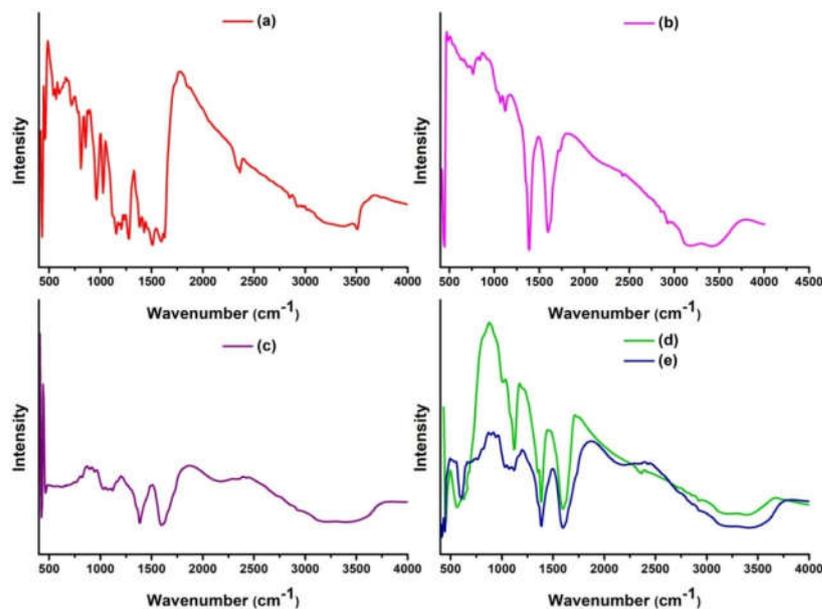


Fig. 1: FTIR spectra of CUR (a), placebo microbeads (b), drug-loaded microbeads (c), MNPs (d) and MNPs-drug-loaded microbeads (e)

The FTIR spectral studies were used to confirm the formation of microbeads. FTIR spectra of CUR (a), placebo microbeads (b), drug loaded microbeads (c), MNPs (d) and drug-loaded MNPs microbeads (e) are shown in fig. 1. FTIR spectra of placebo microbeads (fig. 1b), a peak observed at 3341  $\text{cm}^{-1}$  is responsible for O-H stretching frequency, a peak at 1609  $\text{cm}^{-1}$  indicates C=O stretching frequency. In addition to the above, peaks at 1387  $\text{cm}^{-1}$  and 1121  $\text{cm}^{-1}$  corresponds to C-N stretching and bending vibrations. The FTIR spectra of CUR (fig. 1a), shows a broad peak at 3493  $\text{cm}^{-1}$  assigned to O-H stretching frequency of phenolic group. The peak at 2923  $\text{cm}^{-1}$  corresponds to C-H stretching vibrations, the peaks at 1596 and 1513  $\text{cm}^{-1}$  were assigned to stretching vibration of benzene ring skeleton and mixed (C=O) and (C=C) vibration respectively. The band at 1272  $\text{cm}^{-1}$  corresponds to Ar-O stretching vibrations [33]. On comparing the drug-loaded microbeads (fig. 1c) with placebo microbeads (fig. 1b) a new peak was observed in the drug-loaded microbeads at 1029  $\text{cm}^{-1}$  and also a broad peak appeared at 1601  $\text{cm}^{-1}$  due to the carbonyl group of CUR, which confirmed that the drug molecules are successfully encapsulated in the microbeads. The FTIR spectra of MNPs (fig. 1d) show a peak at 565  $\text{cm}^{-1}$  due to Fe-O stretching frequency, peaks at 1598  $\text{cm}^{-1}$  and 3421  $\text{cm}^{-1}$  are due to bending and stretching vibrations of O-H group which are attached to iron atoms on the surface [34]. In the case of MNPs-drug-loaded microbeads (fig. 1e), similar to drug-loaded microbeads all peaks were observed in MNPs-drug-loaded microbeads, along with a new peak observed at 611  $\text{cm}^{-1}$  due to the presence of Fe-O group, which confirms that the MNPs are successfully loaded in the microbeads.

### DSC analysis

Typical DSC thermograms of CUR (a), placebo microbeads (b), drug-loaded microbeads (c), MNPs-drug-loaded microbeads (d) and MNPs (e) are depicted in fig. 2.

The DSC curve of placebo microbeads (fig. 2b) show two endothermic peaks at 87 and 205 °C. The DSC curve of CUR (fig. 2a) shows an endothermic peak at 181 °C due to its melting, whereas no

such peak was observed in drug-loaded microbeads (fig. 2c), which confirms that the drug has been molecularly dispersed in the microbeads. DSC curve of MNPs (fig. 2e) show an exothermic peak at 572 °C due to phase transition from  $\gamma\text{-Fe}_2\text{O}_3$  to  $\alpha\text{-Fe}_2\text{O}_3$ , whereas no such peak was observed in MNPs-drug-loaded microbeads (fig. 2d), which indicates the stability of MNPs-drug-loaded microbeads due to the interaction between  $\gamma\text{-Fe}_2\text{O}_3$  and polymer matrix [35].

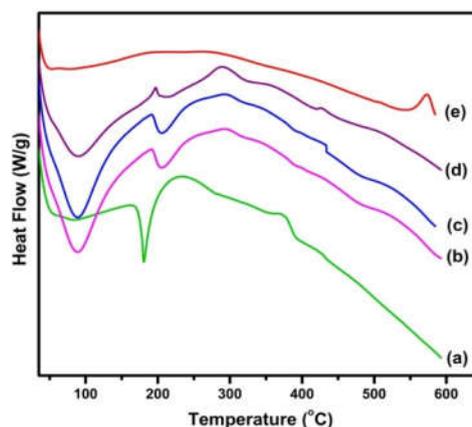


Fig. 2: DSC curves of CUR (a), placebo microbeads (b), drug loaded microbeads (c), MNPs-drug loaded microbeads (d) and MNPs (e)

### TGA analysis

TGA thermograms of CUR (a), placebo microbeads (b), drug-loaded microbeads (c), MNPs-drug-loaded microbeads (d) and MNPs (e) are

displayed in fig. 3. The TGA curve of CUR (fig. 3a) showed a weight loss between the ranges of 50-204 °C due to loss of residual water, followed by a weight loss at 210-398 °C due to the decomposition of CUR. The TGA curve of placebo microbeads (fig. 3b) shows three weight loss steps. The first step was observed in the region of 50-193 °C with weight loss of 23 % due to evaporation of water physically adsorbed on the polymer matrix. The second weight-loss step was observed in the range of 197-313 °C with weight loss of 26 % due to the decomposition of polymer matrix and the last step with weight loss of 19 % in the region of 317-600 °C due to the complete decomposition of polymer matrix. The TGA curve of drug-loaded microbeads (fig. 3c) also shows three consecutive weight loss steps. The first weight loss of 22% was found in the region of 50-190 °C and is attributed to the loss of water bounded on the surface of polymer matrix. The second and third steps were observed in the range of 195-305 °C and 309-600 °C with a weight loss of 30 and 20 % respectively due to the decomposition of the polymer matrix. TGA curve of MNPs (fig. 3e) shows a weight loss of 7% over the range of 50-600 °C indicating that no significant weight loss took place. In the case of MNPs-drug loaded microbeads (fig. 3d), TGA curves show three weight loss steps. The first weight loss step was observed over the range of 50-191 °C with weight loss of 34% due to the loss of free and bound water from the polymer matrix. The second and third weight loss steps were found in the region of 197-301 °C and 305-600 °C, with weight loss of 20 and 16 % due to the decomposition of polymer matrix. From the TGA results, it is suggested that the drug-loaded microbeads and MNPs-drug-loaded microbeads show an overall improvement in the thermal stability of microbeads.

#### SEM analysis

The topographical images of microbeads and MNPs are investigated

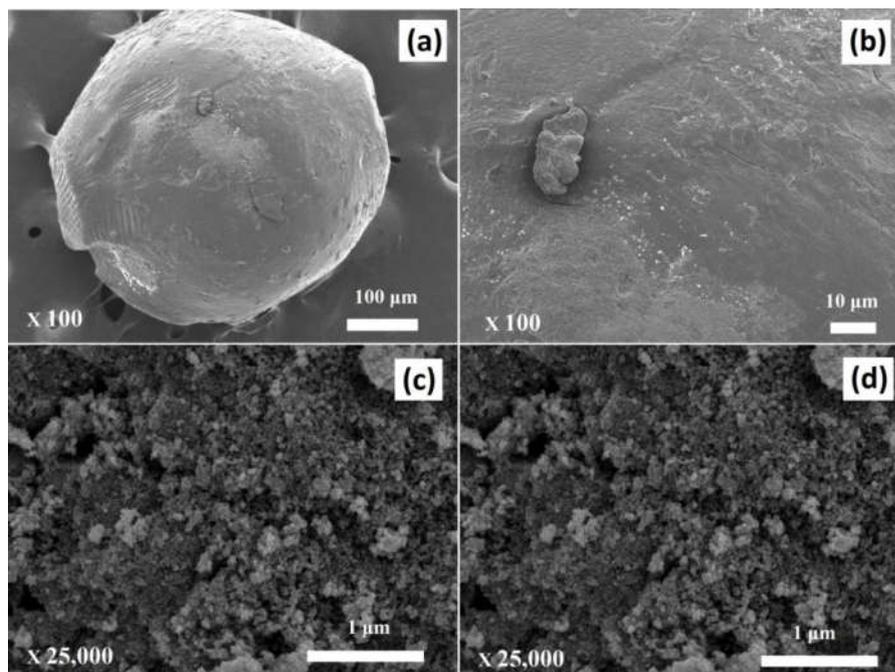


Fig. 4: SEM analysis of drug and MNPs loaded microspheres (a, b), synthesized magnetic nanoparticles (c, d)

#### X-RD analysis

X-RD patterns of CUR (a), MNPs (b), placebo microbeads (c), drug-loaded microbeads (d) and MNPs-drug-loaded microbeads (e) are displayed in fig. 5. The X-RD pattern of MNPs (fig. 5b) show a peak at 35.70° due to the crystalline plane with miller indices of {3 1 1}. The peaks at 30.33°, 43.44°, 54.23°, 57.31° and 62.91° match the magnetite diffraction pattern reported by Banerjee et al., [36]. The results suggest that the developed

MNPs were Fe<sub>3</sub>O<sub>4</sub> nanoparticles. The X-RD pattern of CUR (fig. 5a) shows the most intensive peaks in the 2θ region of 12-28°, suggesting its crystalline nature. Whereas these peaks have not appeared in drug-loaded microbeads (fig. 5d), suggesting that the drug has been molecularly dispersed in the polymer matrix. X-RD pattern of MNPs-drug-loaded microbeads (fig. 5e) show peaks similar to that of MNPs at 30.21°, 35.89°, 57.51° and 62.97° indicating that the MNPs are successfully loaded in the microbeads.

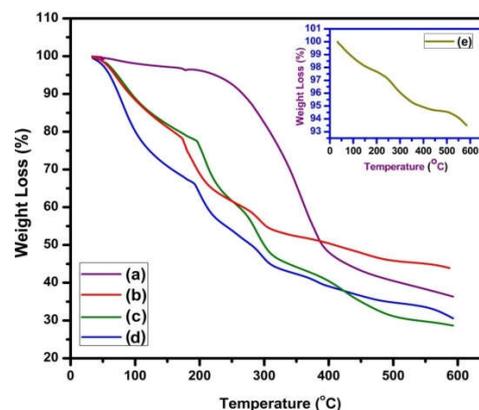


Fig. 3: TGA curves of CUR (a), placebo microbeads (b), drug loaded microbeads (c), MNPs-drug-loaded microbeads (d) and MNPs (e)

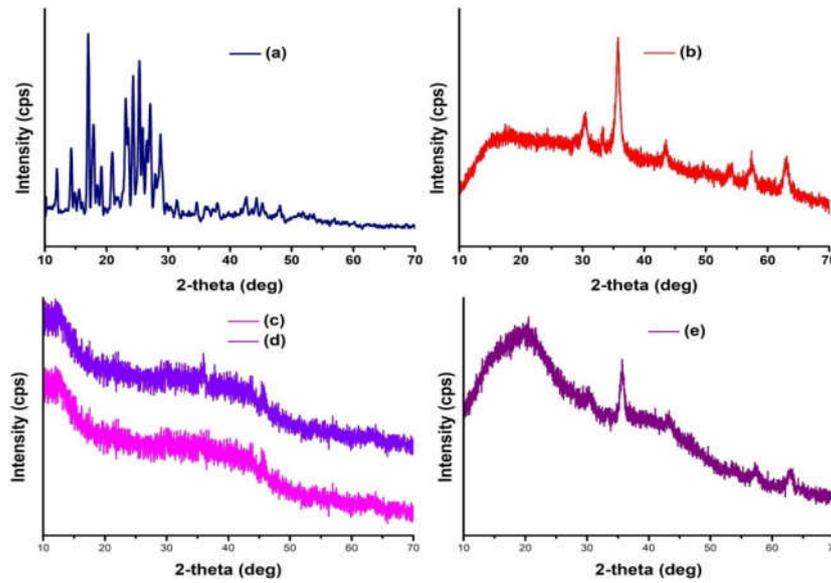


Fig. 5: X-RD patterns of CUR (a), MNPs (b), placebo microbeads (c), drug loaded microbeads (d) and MNPs-drug loaded microbeads (e)

**Encapsulation efficiency**

The percentage encapsulation efficiency of developed CUR encapsulated microbeads was found to be between 39 % and 50 % (table 2). Three different concentrations of CUR, i.e., 100, 150, and 200 mg were encapsulated with constant amounts of MNPs and GA. As the % of drug loading is increased, % EE also increased from 45 to 48%; this trend may be due to higher drug concentration, which causes entrapment of more number of drug molecules in the polymeric matrix leading to higher %EE. With increasing crosslinker

concentration, % EE decreased. For instance, with an increase of GA from 1 to 5 ml, % EE decreased from 45 to 39%. Such a decrease in trend could be attributed to decrease in free volume spaces in the polymeric matrix due to increased crosslinking density, thereby reducing encapsulation efficiency. Further with increasing the % of PVP-co-VAc from 20 to 60% (The SA percentage decreased from 80 to 40%) the % EE increased from 45 to 50, this may be due to more interactions between hydrophobic groups of CUR and PVP-co-VAc, whereas in SA the presence of ionic COO groups inhibited such interactions.

Table 2: Encapsulation efficiency (% EE) of all samples, (Results are expressed as mean±SD, n=3)

S. No.	Formulation code	% EE
1	F1	45±1.3
2	F2	47±0.8
3	F3	50±1.4
4	F4	46±1.1
5	F5	48±1.5
6	F6	42±1.3
7	F7	39±0.6

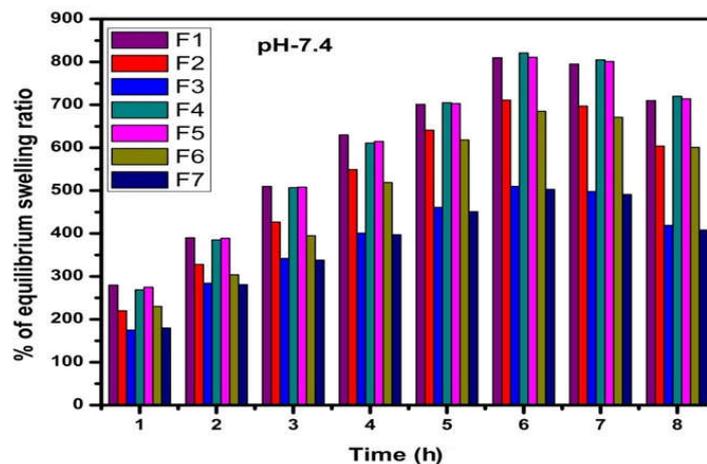


Fig. 6a: Equilibrium swelling ratio for all profiles at pH 7.4, (Results are expressed as mean±SD, n=3)

### Swelling studies

Swelling properties play a crucial role in biomedical applications such as diffusion of drugs, nutrients and other water-soluble metabolites and also for controlled release in drug delivery systems. In order to find out the suitability of microbeads for gastrointestinal drug delivery, swelling studies were performed under both pH 7.4 (simulated intestinal fluid) and 1.2 (simulated gastric fluid) at 37 °C and the results are displayed in fig. 6a and 6b, respectively. The

swelling results indicate that the swelling degree is high in the case of pH 7.4 rather than in the case of pH 1.2. This is because, in lower pH (1.2), -COOH groups of polymer molecule forms a hydrogen bonding with the solvent molecules, which results in the shrinking of polymer network consequently swelling degree decreases. Whereas at higher pH (7.4) the ionic-COO<sup>-</sup> groups of polymer molecules repel the water molecules due to the existence of ionic repulsions. Therefore there is no chance for the formation of hydrogen bonding, which ultimately increases the swelling degree at higher pH (7.4) conditions [37].

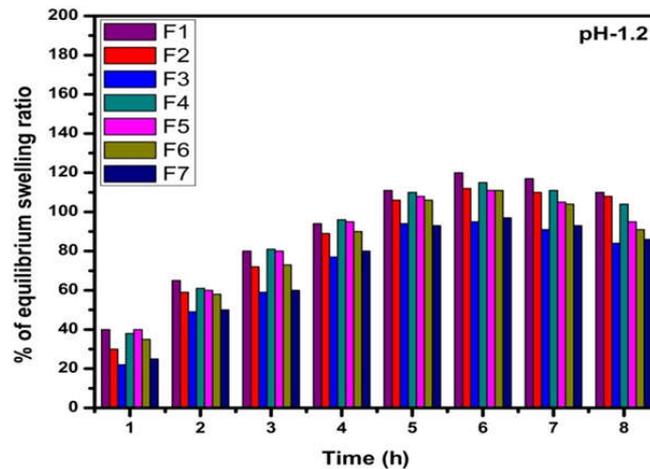


Fig. 6b: Equilibrium swelling ratio for all profiles at pH 1.2, (Results are expressed as mean±SD, n=3)

### In vitro studies

The *in vitro* drug release studies of microbeads under study are investigated under both pH 1.2 and pH 7.4 at 37 °C. The cumulative % of drug release for all profiles at pH 1.2 are displayed in fig. 7. The *in vitro* release studies for all profiles at pH 7.4 are discussed in terms of polymer

blend composition (fig. 8), drug loading (fig. 9) and crosslinker variation (fig. 10). The results reveal that the release rate is higher at pH 7.4 rather than at pH 1.2, this is because at pH 7.4 the carboxylate groups have less interactions with buffer medium, therefore, the network becomes more slack; hence, the entrapped drug molecules easily leach out from the network when compared to pH 1.2.

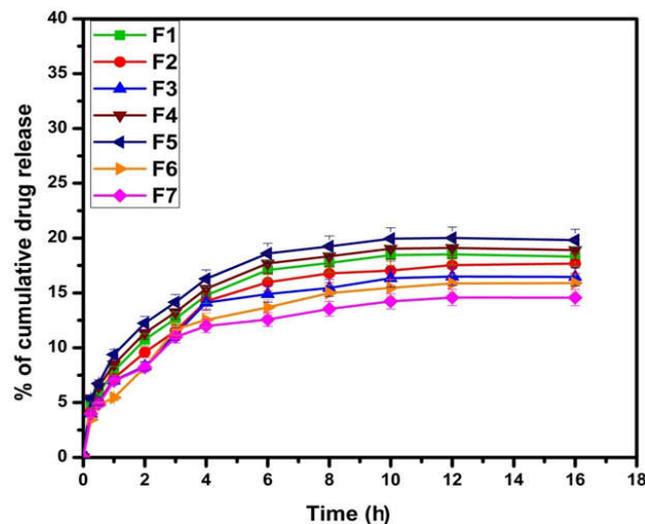


Fig. 7: Cumulative % of drug release in pH 1.2 at 37 °C, (Results are expressed as mean±SD, n=3)

### Polymer variation

Cumulative % of drug release was influenced by changing the polymer blend ratio at constant drug content (100 mg) and crosslinker content (1 ml). From the fig. 8 it was clearly observed that as the content of PVP-co-VAc increased, the cumulative % of

drug release decreased. This trend is observed in the formulations of F1 (80:20), F2 (60:40) and F3 (40:60).

This is due to the presence of hydrophobic part (vinyl acetate) in the polymer matrix, consequently decreasing the leaching of an entrapped drug from the polymer matrix in to the buffer media.

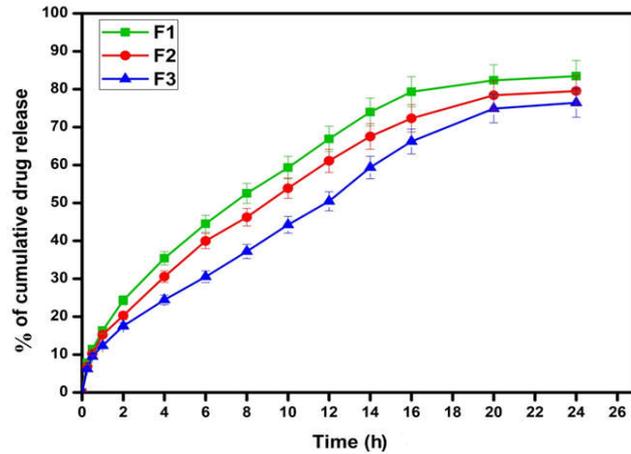


Fig. 8: Effect of polymer blend composition on % cumulative drug release in pH 7.4 at 37 °C (Results are expressed as mean±SD, n=3)

**Drug variation**

The effect of drug content on cumulative % of drug release rate was observed by varying the drug content at constant polymer blend ratio (80:20) and crosslinker content (1 ml). The effect of drug content was observed in the formulations F1 (100 mg), F4 (150 mg) and F5 (200 mg) and the results are displayed in fig. 9. The

formulation F5 shows a higher release rate than the F4 and F1; thus it was concluded that the release depends on the drug present in the matrices i.e., higher the drug, higher is the drug release rate and vice-versa. In other words, lower the amount of drug, lower the drug release rate was observed due to the availability of more free void spaces through which a lesser number of drug molecules could be transported [38].

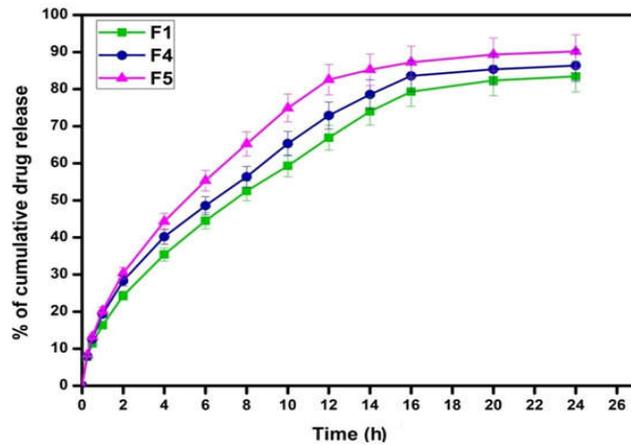


Fig. 9: Effect of drug content on cumulative % of drug release in pH 7.4 at 37 °C (Results are expressed as mean±SD, n=3)

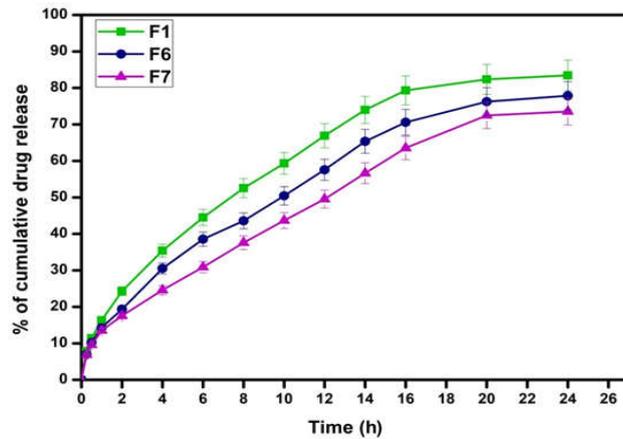


Fig. 10: Effect of crosslinker content on cumulative % of drug release in pH 7.4 at 37 °C (Results are expressed as mean±SD, n=3)

Table 3: Results of % of release kinetics parameters (k and n) of drug in different IPN microbeads formulations

Sample	Korsmeyer-peppas	
	n	r <sup>2</sup>
F1	0.526	0.998
F2	0.530	0.998
F3	0.506	0.995
F4	0.540	0.994
F5	0.562	0.996
F6	0.552	0.997
F7	0.493	0.997

### Crosslinker variation

The effect of crosslinker on the % of drug release rate for formulations F1, F6 and F7 are investigated and the results are displayed in fig. 10. It was clearly noticed that the % of drug release rate of F1 (GA-1 ml) is higher than the F6 (GA-2.5 ml) and F7 (GA-5 ml), because as the crosslinker content increases the % of drug release rate decreases. As the crosslinker content increases, the polymer network becomes more rigid; hence the drug molecules have less chance to escape from the polymer matrix; therefore, the drug release rate decreases [39].

### Drug release kinetics

To confirm the release mechanism, the *in vitro* release results were fitted to the following Korsmeyer-Peppas equation.

$$M_t/M_\infty = kt^n$$

Where  $M_t/M_\infty$  represents the fractional drug release at time  $t$ ,  $k$  is a constant characteristic of the drug-polymer system and  $n$  is the release exponent indicating the type of drug release mechanism. In the present study, the values 'n' are obtained in the range of 0.493-0.562, leading to a shift of transport from Fickian to anomalous type. These results, along with correlation coefficients,  $r^2$  are presented in table 3.

### CONCLUSION

In the present study, SA/PVP-co-VAc/MNPs microbeads were fabricated by a simple ionotropic gelation technique. The microbeads were confirmed by FTIR and DSC studies. Chemical stability and molecular level dispersion of CUR in microbeads were confirmed by FTIR, DSC, TGA, and X-RD, respectively. SEM studies reveal that the beads were spherical in shape with a rough surface and the average diameter of microbeads was found to be 1080 to 1030  $\mu\text{m}$ . The swelling and *in vitro* release studies performed at both pH 1.2 and 7.4 reveal that at pH 7.4 highest swelling and release was observed. *In vitro* releases studies were fitted into the Korsmeyer-Peppas equation, leading to a shift of transport from Fickian to anomalous type. Since the therapeutic efficacy of CUR and MNPs are established previously, based on the drug release studies of CUR loaded SA/PVP-co-VAc/MNPs microbeads, it is observed that the combination of MNPs has not effected the drug release kinetics of CUR. Hence it is suggested that the developed SA/PVP-co-VAc/MNP microbeads were potentially good carriers for drug delivery of CUR and augment chemotherapeutic procedures.

### ACKNOWLEDGEMENT

The authors MCS Subha and K Chowdoji Rao acknowledge the financial support provided by UGC in the form of UGC-BSR faculty fellowship.

### AUTHORS CONTRIBUTIONS

All the author have contributed equally.

### CONFLICT OF INTERESTS

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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