

ISSN- 0975-7058

Vol 12, Special Issue 3, 2020

Full Proceeding Paper

POTENTIAL OF ETHANOL EXTRACT OF MAHKOTA DEWA LEAVES (*PHALERIA MACROCARPA* (SCHECFF.) BOERL.) TO INHIBIT INFLAMMATION IN MOUSE DISTAL COLON INDUCED BY DEXTRAN SODIUM SULFATE (DSS) AND AZOXYMETHANE (AOM)

MUHAMMAD ILHAM DHIYA RAKASIWI¹, KUSMARDI KUSMARDI^{2*}, ARI ESTUNINGTYAS³, ARYO TEDJO⁴

¹Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia, ²Department of Pathological Anatomy, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia, ³Department of Pharmacology and Therapeutics, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia, ⁴Department of Medical Chemistry, Faculty of Medicine, Universitas Indonesia, Jakarta, Indonesia Email: kusmardi.ms@ui.ac.id

Received: 02 Oct 2019, Revised and Accepted: 17 Feb 2020

ABSTRACT

Objective: To demonstrates the ability of P. macrocarpa leaf extract to reduce inflammation of the distal colon in DSS/AOM-induced mice.

Methods: *In vivo* experimental research using Balb/c mice induced by 0.2 ml azoxymethane (AOM) 0.1% once and 1% dextran sodium sulphate (DSS) for one week; additionally, ethanol extract of P. macrocarpa leaves, 25 mg and 50 mg, and 0.84 mg acetosal were given orally. The mice were sacrificed after 20 w. Histopathological examination (hematoxylin-eosin staining) was conducted by counting the average number of goblet cells per crypt, inflammatory focus and angiogenesis.

Results: Ethanol extract of P. macrocarpa leaves was able to prevent the decrease in the number of goblet cells (p<0.05). However, the administration of ethanol *P. macrocarpa* leaf extract could not reduce focal inflammation and angiogenesis in inflammation of the distal colon.

Conclusion: Ethanol extract of the Mahkota Dewa leaves is able to prevent inflammation of the distal colon by preventing the decrease in the number of goblet cells.

Keywords: Angiogenesis, Distal colon, Ethanol extract, Goblet cell, Phaleria macrocarpa leaf

 $@\ 2020\ The\ Authors.\ Published\ by\ Innovare\ Academic\ Sciences\ Pvt\ Ltd.\ This\ is\ an\ open\ access\ article\ under\ the\ CC\ BY\ license\ (http://creativecommons.org/licenses/by/4.0/)\ D01:\ http://dx.doi.org/10.22159/ijap.2020.v12s3.39490$

INTRODUCTION

UC is chronic gastrointestinal inflammation that occurs in the epithelial lining of the large intestine, which causes stomach pain, diarrhea and gastrointestinal bleeding [1]. The global UC incidence is 1.2–20.3 per 100,000 people annually, with a prevalence of 7.6 to 246 per 100,000 people. The incidence and prevalence of Inflammatory Bowel Disease (IBD) is high in European and American populations, whereas in Asian populations, the number of reported cases is low [2]. Patients with UC have a 50% chance of recurrence and 20–30% require colectomy for treatment [3].

UC is a disease caused by immunological mechanisms that occur in people with a genetic predisposition due to an immune system that cannot normally respond to intraluminal antigens; antigens that cause this immunological mechanism are commensal bacteria in the digestive tract [4]. Histopathological examination shows a change in cryptic architecture (shortening or branching crypts), lymphoplasmasitosis basalis and Paneth cell metaplasia [5].

Currently, the choice of UC management depends on the degree of the disease, distribution, causes, frequency of recurrence, extraintestinal manifestations, previous treatment and side effects of the drugs prescribed. The goal of UC management is to provide an improvement in conditions with minimal consumption of steroids while preventing disease complications. The first-line drug given for mild-to-moderate UC is 5-ASA; this drug has been studied by many researchers and shows positive effects in dealing with UC. Unfortunately, 5-ASA can cause a number of side effects, such as diarrhea, nausea, vomiting, headaches and fever, so special attention is required in the use of 5-ASA [6, 7].

Considering these serious health problems, there needs to be an effective, efficient treatment with minimal side effects. Indonesian people have long used many natural substances as medicines; one of these is the Mahkota Dewa (*Phaleria macrocarpa*) leaf. Numerous studies have shown the pharmacological effects of the extract of the P. macrocarpa, such as anti-cancer, antioxidant, antibacterial and anti-inflammatory effects. One of the active ingredients of the P.

macrocarpa extract is phalerin, which works by suppressing the expression of COX2 which causes decreased synthesis of prostaglandin so that the inflammatory reaction is reduced [8].

MATERIALS AND METHODS

Sample size

The number of samples used in this study was calculated using the Federer formula $(t-1) (n-1) \ge 15$. The number of groups in this study was 5 groups, so the number of replications in each group was:

- (5-1) (n-1) ≥ 15 heads
- (n-1)/4 15/4
- (n-1) ≥ 3175
- n ≥ 4.75

Based on the calculation of the formula above, five mice were obtained for each group; in total, 25 Balb/c mice, which were randomly divided into five groups, were used.

Material

Dextran sodium sulfate (DSS) with a molecular weight of 36,000– 50,000 was obtained from Regent Science Industry Limited (RSC), Hong Kong. The ethanol extract of P. macrocarpa leaves was obtained from the Biopharmaca IPB Study Center, Bogor, Indonesia.

Experiment design

In vivo experimental research using Balb/c mice divided into five groups. Each group consists of 5 mice:

1. The normal group (N) consisted of mice that were not given any treatment.

2. The negative group (K-) consisted of mice given 0.2 ml AOM 0.1% w/v ip once for seven days and then given a 1% w/v DSS solution through drinking water every day for seven days *ad libitum*.

3. Positive group (K+) consisted of mice given 0.2 ml AOM 0.1% w/v ip once for seven days and then given an oral suspension of acetosal 0.4% w/v in the amount of 0.21 ml (equivalent to 0.84 mg of acetosal) and a 1% w/v DSS solution with drinking water every day for seven days *ad libitum*.

4. High dose group (DT) consisted of mice given 0.2 ml AOM 0.1% w/v ip once for seven days and then given an extract of 25% w/v P. macrocarpa orally in the amount of 0.2 ml (equivalent to 50 mg extract) and 1% DSS w/v solution with drinking water every day for seven days *ad libitum*.

5. The low dose group (DR) consisted of mice given 0.2 ml AOM 0.1% w/v ip once for seven days, and then given 12.5% w/v P. macrocarpa leaf extract orally in the amount of 0.2 ml (equivalent to 25 mg extract) and a 1% DS w/v solution with drinking water every day for seven days *ad libitum*.

Mice were sacrificed after 20 w of being given treatment according to their group. Distal colonic tissue was then stained with hematoxylin-eosin for histopathological observation.

Histopathological examination

Histopathological observations were made at the Pathological Anatomy Laboratory FKUI, Jakarta, Indonesia. Observations focused on calculating the number of goblet cells per crypt, the focus of inflammation and angiogenesis in six predetermined fields. Observations were made in a double-blind manner. Observations were made at 400-times magnification and then analyzed using ImageJ software (NHI).

Ethical approval

This study passed the ethical review from the Faculty of Medicine University of Indonesia Health Research Ethics Committee with letter number 0891/UN2. FI/ETIK/2018.

Statistical analysis

Statistical analysis was performed using the One-Way ANOVA test, followed by Tukey's Post Hoc test for goblet cell variables and inflammatory focus. Angiogenesis variables were analyzed using the Kruskal-Wallis test because the data distribution was not normal.

RESULTS

Effect of administration of *P. macrocarpa* leaf extract on the number of goblet cells

Goblet cells play a protective function in the mucosal lining of the colon by secreting mucosa [9–11]. Damage to the mucous layer causes an increase in permeability to bacteria and toxins that can damage epithelial cells and cause systemic inflammation, such as UC [12]. In UC, there is a depletion of goblet cells due to inflammation processes. This is related to a decrease in glycosylation of mucin accompanied by the absence of the MUC2 and MUC3 genes in goblet cells [13]. Goblet cell depletion in UC is also associated with impaired induction of Hath1 and KLF4 differentiation factors during inflammation [14]. The effect of P. macrocarpa extract on observed goblet cell counts with hematoxylin-eosin staining using 400-times magnification is shown in fig. 1.

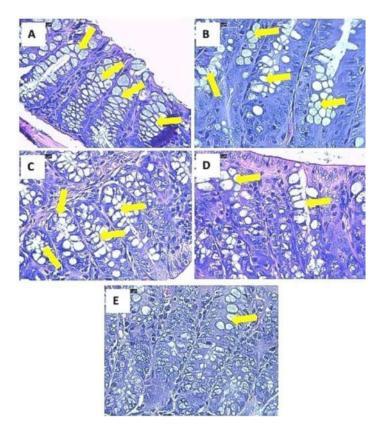


Fig. 1: Effect of administration mahkota dewa extract to the number of goblet cells: (A) Normal group, (B) DSS/AOM+Aspirin 0.86 mg, (C) DSS/AOM+50 mg extract, (D) DSS/AOM+Extract 25 mg and (E) DSS/AOM

The administration of P. macrocarpa leaf extract was able to prevent a significant decrease in the number of goblet cells (p<0.05) (fig. 2). The results of this study are supported by a number of other studies. A study by Suprapti *et al.* examined the effect of the P. macrocarpa leaf extract on colon inflammation in mice. From this study, it was

concluded that the P. macrocarpa leaf extract can reduce colonic tissue damage and prevent an increase in the expression of COX-2, NOS and β -catenin [15]. A study by Maharani *et al.* showed that the P. macrocarpa leaf extract with a dose of 100 mg, 200 mg, and 300 mg can inhibit a decrease in the number of goblet cells in the colon [16].

Effect of administration of *P. macrocarpa* leaf extract on the amount of angiogenesis

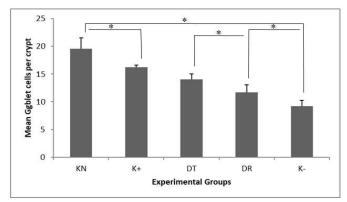
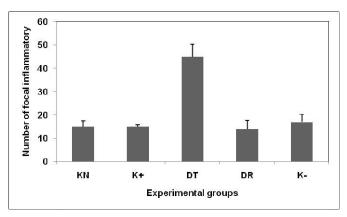


Fig. 2: Administration of Dewa Dewa leaf extract to normal group goblet (KN) cells, (K+) Aspirin 0.86 mg, (DT) DSS/AOM+50 mg extract, (DR) DSS/AOM+25 mg extract and (K-) DSS/AOM

The administration of DSS to the colon was able to increase the number of angiogenesis. In this study, the administration of the *P*.



macrocarpa leaf extract did not reduce the amount of angiogenesis in the DSS/AOM-induced distal colitis (p = 0.895).

Fig. 3: Effect of administration of dewa dewa leaf extract to the number of angiogenesis (KN) normal group, (K+) Aspirin 0.86 mg, (DT) DSS/AOM+50 mg extract, (DR) DSS/AOM+25 mg extract and (K-) DSS/AOM

Effect of administration of *P. macrocarpa* leaf extract on the amount of inflammatory focus

The focus of active inflammation is defined as cryptic damage due to neutrophil infiltration. The discovery of neutrophil infiltration in crypts and cryptic abscesses indicates an inflammatory focus. Research by Osmond *et al.* showed that the focus of inflammation leads to clinical diagnosis in UC [17, 18]. In UC patients, active inflammation is characterized by the presence of neutrophils in the stool, whereas the severity of the disease is characterized by neutrophil infiltration [19].

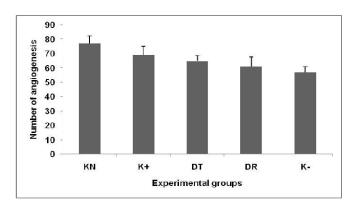


Fig. 4: Effect of administration of dewa dewa leaf extract to the number of focal inflammatory (KN) normal group, (K+) Aspirin 0.86 mg, (DT) DSS/AOM+50 mg extract, (DR) DSS/AOM+25 mg extract and (K-) DSS/AOM

4th International Conference and Exhibition on Indonesian Medical Education and Research Institute 2019

In this study, the administration of *P. macrocarpa* leaf extract did not show a reduction in the amount of inflammation in focus. The amount of inflammatory focus in the control group did not differ significantly with the group given the extract (p = 0.082) (fig. 4).

DISCUSSION

Phaleria macrocarpa is a tropical plant of the Thymelaceae family that commonly grows in Papua, Indonesia. Extracts of *P. macrocarpa* leaves are used for a number of pharmacological activities because of its anti-tumor, anti-inflammatory and anti-fungal effects. Anti-inflammatory activity of *P. macrocarpa* are due to its contents, including tannins, terpenoids, saponins, flavonoids and phenols [8]. The aim of this study is to determine the histopathological effect of distal colon inflammation by inhibition of the ethanol extract of *P. macrocarpa* leaves.

Goblet cells play a protective function in the mucosal lining of the colon by secreting mucosa [9–11]. Damage to the mucous layer causes increased permeability to bacteria and toxins that can damage epithelial cells and cause systemic inflammation, such as UC [12]. In UC, a depletion of the number of goblet cells occurs due to inflammation processes [13]. The results of this study show that there were significant differences in the number of distal colonic goblet cells between the groups given the extract of the *P. macrocarpa* leaves and the group that was only given AOM/DSS (p<0.05). Giving 25 mg and 50 mg dosages of *P. macrocarpa* leaf extract can prevent a decrease in the number of goblet cells after the inflammation process occurs; there is a significant difference between the 25 mg and 50 mg doses. In addition, high-dose (50 mg) *P. macrocarpa* leaf extract shows the same anti-inflammation effect (inhibitory decrease in the number of goblet cells) as acetosal.

The results of this study are supported by a number of other studies. The study by Suprapti *et al.* examined the effect of the extract of *P. macrocarpa* leaf on colon inflammation in mice. From these studies, it was concluded that the extract of the *P. macrocarpa* leaf can reduce colonic tissue damage, as well as prevents an increase in COX-2, NOS and β -catenin expression [20]. Studies by Maharani *et al.* showed that 300 mg can inhibit a decrease in the number of goblet cells in the colon [16].

The increased amount of foci of inflammation in UC is caused by the role of neutrophils, which are the first immune cells that respond during the inflammation process. Uncontrolled activation of neutrophils with UC causes damage to the colonic epithelial tissue and allows infiltration into the crypts [21]. Showed the antiinflammatory effect of *P. macrocarpa* leaf extract by suppressing the amount of inflammatory focus in DSS-induced UC [16].

In this study, the administration of *P. macrocarpa* leaf extract did not show a significant difference (p>0.05). The amount of inflammatory focus in the control group did not differ significantly from the group given extract. In addition, the amount of inflammatory focus in the group that was given the highest dose of *P. macrocarpa* leaf extract showed the highest number of inflammatory focus. Differences in results from previous studies may be due to the longer induction period of this trial (20 w), which lead to apoptosis or carcinogenesis.

In UC, continuous ulceration is followed by tissue regeneration. This increases the need for oxygen and nutrient supply to the colon tissue. Under physiological conditions, the process of angiogenesis occurs due to a balance between pro-and anti-angiogenic factors; but with UC, the process of angiogenesis becomes uncontrollable due to chronic inflammation, or what is referred to as 'immune-driven angiogenesis' [22–25]. Administration of DSS does not increase the number of blood vessels in the distal colonic tissue. In addition, administration of acetosal or extracts of the *P. macrocarpa* leaf did not show a decrease in the number of angiogenesis.

The ethanol extract of the *P. macrocarpa* leaves is able to prevent the inflammation process in the distal intestine by preventing a decrease in the number of goblet cells. However, P. macrocarpa leaf extract cannot reduce the amount of angiogenesis or inflammatory focus.

CONCLUSION

The ethanol extract of the Mahkota Dewa leaves (*P. macrocarpa*) is able to prevent the inflammatory process in the distal intestine by

preventing a decrease in the number of goblet cells, but it cannot reduce the amount of angiogenesis and inflammatory focus.

ACKNOWLEDGEMENT

We wish to extend our sincere thanks to Mrs. Tati Suprapti, our supervisor of this research and pathological anatomy laboratory staff. We also acknowledge the Directorate of Research and Innovation, Universitas Indonesia.

FUNDING

This research was granted by Hibah PITTA B 2019.

AUTHORS CONTRIBUTIONS

All the authors have contributed equally.

CONFLICT OF INTERESTS

The authors declare no competing interest.

REFERENCES

- 1. Conrad K, Roggenbuck D, Laass MW. Diagnosis and classification of ulcerative colitis. Autoimmun Rev 2014;13:463–6.
- Danese S, Fiocchi C. Ulcerative colitis. N Engl J Med 2011;365:1713–25.
- Zhang Z, Kennedy H. Ulcerative colitis: current medical therapy and strategies for improving medication adherence. Eur J Gastroenterol Hepatol 2009;21:1–8.
- Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. Nat Rev Immunol 2009;9:313–23.
- 5. Blumberg RS. Inflammation in the intestinal tract: pathogenesis and treatment. Dig Dis 2009;27:455–64.
- Hauso Ø, Martinsen TC, Waldum H. 5-aminosalicylic acid, a specific drug for ulcerative colitis. Scand J Gastroenterol 2015;50:933–41.
- Cottone M, Renna S, Modesto I, Orlando A. Is 5-ASA still the treatment of choice for ulcerative colitis? Curr Drug Targets 2011;12:1396–405.
- Altaf R, Asmawi MZB, Dewa A, Sadikun A, Umar MI. Phytochemistry and medicinal properties of *Phaleria macrocarpa* (Scheff.) Boerl. extracts. Pharmacogn Rev 2013;7:73–80.
- 9. Dharmani P, Srivastava V, Kissoon Singh V, Chadee K. Role of intestinal mucins in innate host defense mechanisms against pathogens. J Innate Immun 2009;1:123–35.
- McGuckin MA, Eri R, Simms LA, Florin TH, Radford Smith G. Intestinal barrier dysfunction in inflammatory bowel diseases. Inflamm Bowel Dis 2009;15:100–13.
- 11. Shirazi T, Longman R, Corfield A, Probert C. Mucins and inflammatory bowel disease. Postgrad Med J 2000;76:473–8.
- Longman RJ, Poulsom R, Corfield AP, Warren BF, Wright NA, Thomas MG. Alterations in the composition of the supramucosal defense barrier in relation to disease severity of ulcerative colitis. J Histochem Cytochem 2006;54:1335–48.
- Dorofeyev AE, Vasilenko IV, Rassokhina OA, Kondratiuk RB. Mucosal barrier in ulcerative colitis and Crohn's disease. Gastroenterol Res Pract 2013. DOI:10.1155/2013/431231
- 14. Gersemann M, Becker S, Kübler I, Koslowski M, Wang G, Herrlinger KR, *et al.* Differences in goblet cell differentiation between Crohn's disease and ulcerative colitis. Differentiation 2009;77:84–94.
- Erben U, Loddenkemper C, Doerfel K, Spieckermann S, Haller D, Heimesaat MM, *et al.* A guide to the histomorphological evaluation of intestinal inflammation in mouse models. Int J Clin Exp Pathol 2014;7:4557–76.
- Maharani R, Kusmardi, Elya B. Inhibitory activity goblet depletion and focal inflammatory *Phaleria macrocarpha* leaves ethanol extract on crypta mouse after dextran sodium sulphate induction. Int J Pharmtech Res 2018;12:37–48.
- 17. Greenson JK, Stern RA, Carpenter SL, Barnett JL. The clinical significance of focal active colitis. Hum Pathol 1997;28:729–33.
- Osmond A, Ashok D, Francoeur CA, Miller M, Walsh JC. Is focal active colitis of greater clinical significance in pediatric patients? a retrospective review of 68 cases with clinical correlation. Hum Pathol 2018;74:164–9.

4th International Conference and Exhibition on Indonesian Medical Education and Research Institute 2019

- Bressenot A, Salleron J, Bastien C, Danese S, Boulagnon Rombi C, Peyrin-Biroulet L. Comparing histological activity indexes in UC. Gut 2015;64:1412–8.
- 20. Suprapti T, Louisa M, Tedjo A, Kusmardi, Fadilah, Handjari DR, et al. Antiinflammatory effect of mahkota dewa (*Phaleria* macrocarpa (Scheff.) Boerl.) leaves extract on colon carcinogenesis induced by azoxymethane and dextran sodium sulphate: focus on the iNOS, β-catenin and COX-2 expression. Asian J Appl Sci 2014;2:511–27.
- 21. Wera O, Lancellotti P, Oury C. The dual role of neutrophils in inflammatory bowel diseases. J Clin Med 2016;5:118.
- 22. Pousa ID, Mate J, Gisbert JP. Angiogenesis in inflammatory bowel disease. Eur J Clin Invest 2008;38:73–81.
- Alkim C, Alkim H, Koksal AR, Boga S, Sen I. Angiogenesis in inflammatory bowel disease. Int J Inflam 2015. DOI:10.1155/2015/970890
- Jerkic M, Peter M, Ardelean D, Fine M, Konerding MA, Letarte M. Dextran sulfate sodium leads to chronic colitis and pathological angiogenesis in Endoglin heterozygous mice. Inflamm Bowel Dis 2010;16:1859–70.
- Chidlow JH, Shukla D, Grisham MB, Kevil CG. Pathogenic angiogenesis in IBD and experimental colitis: New ideas and therapeutic avenues. Am J Physiol Gastrointest Liver Physiol 2007;293:G5–G18.

 $4^{
m th}$ International Conference and Exhibition on Indonesian Medical Education and Research Institute 2019