

## THE NATURAL AND COMMERCIAL SOURCES OF HYDROXYAPATITE/COLLAGEN COMPOSITES FOR BIOMEDICAL APPLICATIONS: A REVIEW STUDY

MUSTAFA MUDHAFAR<sup>1,\*</sup>, H. A. ALSAILAWI<sup>1</sup>, ISMAIL ZAINOL<sup>2</sup>, MOHAMMED SACHIT HAMZAH<sup>3</sup>, SAHI JAWAD DHAHI<sup>1</sup>, RUAA KADHIM MOHAMMED<sup>1</sup>

<sup>1</sup>Department of Anesthesia and Intensive Care Techniques, Faculty of Al-Tuff Collage, Karbala, 56001, Iraq, <sup>2</sup>Department of Chemistry, Faculty of Science and Mathematics, Universiti Pendidikan Sultan Idris, Tanjung Malim, Perak, Proton, 35900, Malaysia, <sup>3</sup>Medical Laboratory Technique, Kut University College, Al-kut, Wasit, Iraq, 52001  
Email: almosawy2014@gmail.com

Received: 13 Feb 2022, Revised and Accepted: 11 Apr 2022

### ABSTRACT

Bone is considered the core unit that forms the human body's skeleton, consisting primarily of hydroxyapatite (HA) and collagen (Col). The composites of hydroxyapatite/collagen had been prepared through different fabricated techniques and were used in many bone defects as biomaterials for bone tissue engineering. The incorporation of HA and collagen is possible due to the biocompatibility of collagen and the high mechanical properties of the HA. HA/Col composites have been used in many medical and biological fields. Current study have been discussed the synthesis and characterization techniques of HA/Col composites; the study have been included to study the cytotoxicity and cell attachment of the composites, along with their applications, as well as barriers that still remain to their successful development for clinical application.

**Keywords:** Hydroxyapatite, Collagen, HA/Col composites, Biomedical applications

© 2022 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<https://creativecommons.org/licenses/by/4.0/>)  
DOI: <https://dx.doi.org/10.22159/ijap.2022v14i4.44411>. Journal homepage: <https://innovareacademics.in/journals/index.php/ijap>

### INTRODUCTION

Bone defects may occur for several reasons, such as injuries, illnesses, surgical interventions, and accidents, some of which may heal on their own. However, bone defects greater than 1/3 an inch (≈ 8 mm) cannot be healed on their own [1]. Therefore, bone substitutes were used to fill up and enhance bone defects to allow for the rapid healing process [2]. These substitutes provide structural and mechanical support to enhance bone tissue formation or fill gaps to facilitate the healing of bone tissue. Bone substitutes have been widely used in plastic surgery, oral, maxillofacial, dental, and orthopedic surgery, making it one of the most implanted tissues in the medical field [3].

The structure of natural bone is a composite material comprised of organic and inorganic elements [4]. The organic materials are mainly Col fibers containing tropocollagen, which make up most of the organic constituent of bone, and provide strength to the bone [5]. The inorganic materials are mainly calcium (Ca) and phosphorus (P) in the form of HA [6, 7]. However, both of HA and Col are formed the structure of the bone, naturally.

The incorporation of HA and Col is possible due to the biocompatibility of Col and high mechanical properties of the HA, and it is widely utilized as the biomaterial to enhance the healing of bone defect, as well as a replacement material for bone defects. Nonetheless, despite its advantages, several issues of HA/Col composite concern the public. Tampieri *et al.* [3], reported the successful development of HA/Col composite that showed excellent bioactivity properties, which lead to its use as a bone filler for the defect bones. The composite of HA/Col had been prepared through different fabricated techniques and were used in many bone defects as biomaterials for bone tissue engineering. The research were focusing on the years from 2010 into 2021, but there is some old references because some of them as a books and important to add.

#### Preparation methods of HA/Col composites

The HA/Col composites have been fabricated and widely studied for bone engineering purposes. As have been mentioned, pure Col has weak mechanical properties; however, the use of col is limited and to enhance the mechanical properties of Col, the researchers have combined it with HA using different methods.

Significant aim to synthesis HA/Col composites to produce materials that have identical properties of the natural composites. There are

many techniques have been reported to produce HA/Col composites such as freezing-drying [8], 3D printing [9], situ precipitation [10], co-precipitation [11], electrospinning [12], and dehydrothermal [13], fig. 1 shows the most synthesis techniques which have been used to prepared composites of HA/Col. The previous studied have been focused on these two materials due to their mechanical properties and biocompatibility. The researchers were reported to modify the methods of the preparation for the composites of HA/Col. The study of Ficai *et al.*, [14], has been reported to prepared composites of HA/Col by using self-assembled; briefly, 80:20 (HA/Col) were mixed together, the pH of the mixture has been adjusted at 9 via using NaOH as a calibration solution and the temperature kept during the synthesis process of the composites at ~37 °C. The study was reported to new way to calculate the SD for the fibres of the 2D composites. Walsh *et al.*, 2019, have been reported to synthesize composites of HA/Col via using lyophilisation method, the ratio of composites was 70:30 (HA: Col). 1.8 g of Col has been dissolved in 0.05M acetic acid. Col mixture was centrifuged for 90 min 15,000rpm. 3.6g of HA dissolved in 0.05M acetic acid, then added slowly to the Col mixture.

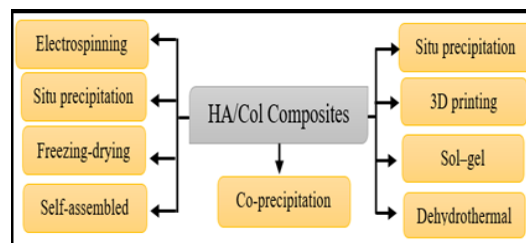


Fig. 1: Synthesis techniques of HA/Col composites (Source: author)

The HA/Col composites were synthesis by using the self-assembling method, the method includes two steps. The first step include to treated Col with the solution of Ca(OH)<sub>2</sub>, then stirred for 24 h. The second step the solution of H<sub>3</sub>PO<sub>4</sub> was added to the mixture. The base of Ca(OH)<sub>2</sub> have been added to adjusted the pH [17, 18]. The freeze-drying method has been used to preparation HA/Col. The

study of Siswanto *et al.*, [19], has been reported to synthesis the composites from natural sources. HA (marine) has been dissolved in phosphoric acid and the Col (bone chicken) have also been dissolved in acetic acid. The solution of HA and Col have been mixed together and stirring them to make a homogeneous solution. The mixture of HA-Col were frozen under  $-80^{\circ}\text{C}$ , for 6 h.

Recently the applications of HA/Col as a Nano sizes have been gotten bulk of attention in materials engineering due to the properties of Nano-materials which could be used in many applications. The ability of the composites can be investigated via analysis of their chemical, physical and biological properties during using many measurements.

**Table 1: Synthesis methods of HA/Col composites**

No	Method	Composites	Application	Ref
1	Freezing-drying	Chitosan/Col/HA	Cartilage tissue engineering	[8]
2	3D printing	HA/Col	Osteochondral regeneration	[9]
3	3D printing	Zinc Silicate/Nano HA/Col	Bone regeneration	[10]
4	Situ precipitation	HA/Col	Bone tissue engineering	[11]
5	Co-precipitation method	HA/Col-magnetite	Bone cancer treatment	[12]
6	Electrospinning	HA/Col	Bone regeneration engineering	[13]
7	Electric field orientation	HA/Col	Cortical bone defect	[14]
8	Dehydrothermal	HA/Col	Bone repair	[16]
9	Self-assembled	HA/Col	Bone regeneration engineering	[15]
10	Sol-gel	HA/Col	Injectable bone substitute	[16]
11	Lyophilisation	HA/Col	Bone defect	[17]

#### The composites of HA/Col incorporate with materials

The HA/Col composites had been prepared through different fabricated techniques and were used in many bone defects as biomaterials for bone tissue engineering. The composite have been incorporated by different sizes (Nano-size and micro-size) with many other materials such as calcium phosphate (CaP) [20], poly(L-lactide) (PLLA) [21], Poly(vinyl alcohol) (PVA) [22], chitosan [23], and metals such as iron (Fe) [24], to develop the properties of the composites, which will possess high compatibility and bio-degradable properties for bone repair.

#### The composites of HA/Col/natural polymers

The composites of HA/Col have to be incorporated with many natural polymer to improve the properties. The previous studies have been reported to prepared the composites of HA/Col with natural polymers such as gelatine [25], cellulose [26], chitosan [27], chitin [28] and starch [29] as it shown in the table 2. Kaviani *et al.*,

[22], was reported to incorporate HA/Col with chitosan by using freezing method. The application of the composites was for cartilage tissue engineering. This study included a cheaper way and environmentally friendly for the preparation of the composite.

The properties of the HA/Col have been improved by incorporated with gelatine. The unidirectional freeze-casting method has been used to synthesize the composites. The results of the availability study against human bone-derived osteoblast, which have been compared with the control cell, showed improved in proliferation, differentiation and adhesion [26, 30]. Study of He *et al.*, [27], reported to prepared HA/Col/cellulose by using in-situ precipitation method for bone tissue engineering. The results showed improved in the swelling ratio of the composites. The mechanical properties and biodegradation have been investigated; the compression strength was increased to 20-40 MPa, which is almost nearby the compression for the natural bone.

**Table 2: The HA/Col composites incorporated with the natural polymers**

Natural polymer	Techniques	Ref
Gelatine	Unidirectional freeze-casting	[26]
Cellulose	In-situ precipitation	[27]
	Simple mixing method	[28]
Chitosan	The freeze-gelation process	[22]
	Simple mixing method	[31,32]
Chitin	The freeze-thawing process	[33]
Starch	Simple mixing method	[29]

#### The composites of HA/Col/synthetic polymers

The composites of HA/Col/synthetic polymers were synthesis and has shown unique properties that enable it to be used in many fields. Many synthetic polymers have been incorporated with the composites of HA/Col, such as Poly (L-lactide) (PLLA) [34], Polylactide-co-glycolide (PLGA) [35], Polyvinyl alcohol (PVA) [36], Poly (methyl methacrylate) (PMMA) [28] and Polycaprolactone [37], as shows in table 3. The previous studies were reported to study the ability of the composites of HA/Col for bone tissue engineering through determined the chemical, physical and biological properties of the composites.

Zhou *et al.*, [35], incorporated PLLA with the HA/Col composites by using electrospinning method. The composites were characterized for their biocompatibility against mouse osteoblasts MC3T3-E1. The results have been shown to enhance the spreading, proliferation and adhesion for the cell. The biodegradation study was determined during 80 d, and the composites showed high stable composition and didn't change in the morphology of the composites. Ariesanti *et al.*, [38], were reported to prepared HA/Col/PVA by using a simple mixing method. The cytotoxicity of the composite of HA/Col/PVA showed high cell availability in the MTT assay.

**Table 3: The HA/Col composites incorporated with the synthetic polymers**

Synthetic polymer	Techniques	Ref
Poly (L-lactide) (PLLA)	Optimized sol-gel method.	[34]
Polylactide-co-glycolide (PLGA)	Electrospinning	[35]
Polyvinyl alcohol (PVA)	Stimulate method	[36]
Polycaprolactone	freeze-dried method	[37]
Poly(methyl methacrylate) (PMMA)	Simple mixing method	[28]

### The composites of HA/Col/metals

The parameters of metallic materials have significant properties, which have been shown to have properties higher than ceramics, which gives it a priority to be used in the field of tissue engineering. The biocompatibility of the metallic is lower than ceramics, which leads to allergic reactions in the blood clots [39]. The metallic nanomaterials have unique properties such as anti-microbial activity, high ratio of surface area, and biological, mechanical and physical properties [40, 41]. The composites of HA/Col were incorporated with metals and metals oxide for example, gold (Au) [42], silver (Ag) [43, 44], magnetite (Fe<sub>3</sub>O<sub>4</sub>) [45], graphene oxide (GO) [46] and Iron oxide (Fe<sub>2</sub>O<sub>3</sub>) [11] as it shown in table 4. Characterization of the composites has been shown high ability to be used to inhibition the growth of bacteria [47, 48], in addition, to use in the regeneration of bone tissue engineering [49, 50]. The HA/Col composites don't have ability to inhibition the growth of bacteria; however, recent studies have been focused to incorporated composites of the HA/Col with nanomaterials to use as an antibacterial [51, 52]. The compatibility of the composites HA/Col with metals was investigated and show high compatibility *in vitro* study [54].

Ciobanu *et al.*, [45], reported to synthesis HA/Col with Ti for antibacterial activity. The composite was determined its ability to use as an antibacterial against *staphylococcus aureus* (S. aureus) and *Escherichia coli* (E. coli), the coated composites showed significant

function against both of these bacterial. HA/Col composite was incorporated with Au nanoparticles by using the microwave-assisted green method and investigated for the biocompatibility study against MG-63 cells. The results show high availability of the MG-63 cell after 24 h, and interaction was observed clearly in the SEM image [42]. Song *et al.*, [9], reported to synthesis of composites of HA/Col with zinc silicate by using the hydrothermal method for bone regeneration. Biocompatibility study have been determined for the ZS/HA/Col composites against bone marrow stromal cells (BMSCs) and the availability of the BMSCs after 24 h was very high. According to previous studies the composites of HA/Col with the metals have been shown high biocompatibility which can be used safely in the implants.

### The composites of HA/Col/Drugs

The present studies have been taken to trend to the drug-delivery system. The systems of the drug delivery has the ability to use in the bone pathologies such as osteosarcoma, osteomyelitis, and osteoporosis. The composites of HA/Col were incorporated with wide range of drugs such as paclitaxel [23], cisplatin [55], vancomycin [56], tetracycline [57] and alendronate [58]. The most studies have been focused to incorporate the composites with the drugs to design new antibiotic for inhibition of bacteria growth [59]. The other applications of using drugs in the drug delivery system are anti-cancer [23], biocompatibility [60], and anti-osteoporosis [61].

**Table 4: The HA/Col composites incorporated with metals and metal oxides**

No	Metals and metal oxide	Methods	Applications	Ref
1	Gold (Au)	Microwave-assisted	Tissue engineering and drug delivery	[47]
2	Silver (Ag)	Simulated body fluid (SBF)	Orthopedic	[43]
3	AgNPs	Co-precipitation	Antimicrobial	[44]
4	Titanium (Ti)	Biomimetic method	Bone implants	[42]
5	Zinc (Zn)	Freeze-dryer	Bone regeneration	[9]
6	Magnetite (Fe <sub>3</sub> O <sub>4</sub> )	Co-precipitation	Bone cancer treatment	[11]
7	Graphene oxide (GO)	Electrodeposition	Antibacterial effect	[53]
8	Iron oxide (Fe <sub>2</sub> O <sub>3</sub> )	Co-precipitation	Bone fractures	[23]
9	Titanium (Ti)	Electrochemical deposition	Biocompatibility	[55]

**Table 5: The HA/Col composites incorporated with drugs**

No	Drugs	Methods	Applications	Ref
1	Paclitaxel	Hydrothermal	Anticancer	[23]
2	Cisplatin	Bone Cancer	In situ treatment	[55]
3	Vancomycin	Electrospinning	Antimicrobial Activity	[56]
4	Vancomycin gentamicin	Electrospinning	Antimicrobial Activity	[57]
5	Vancomycin	Freeze-dryer	Biocompatibility	[58]
6	Vancomycin	3D printing	Enhance Osseo integration and antimicrobial activity	[62]
7	Tetracycline	Mineralization	Antimicrobial Activity	[60]
8	Gentamicin	Hydrothermal	Antibacterial	[61]
9	Alendronate	Freeze-dryer	Bone regeneration as a anti-osteoporosis	[62]
10	Alendronate	Freeze-drying	Bone regeneration	[63]

Many studies have been reported to incorporate HA/Col with wide range of antibiotics. The vancomycin has been used widely with composites. The study of suchý *et al.*, [60], reported to loaded of vancomycin with the HA/Col via using the electrospinning method for antimicrobial activity against *Staphylococcus aureus* and *Staphylococcus epidermidis*, the results showed a significant effect against both of the bacterial compared with the HA/Col composites which did not show any inhibition for the growth of the bacteria. The vancomycin/HA/Col composites were modified by incorporated with gentamicin to enhance the ability of the composites against both of the bacteria [65].

These composites have been investigated for the biocompatibility study through tested *in vivo* against wide range of the cells such as human osteoblast-like cell line (SAOS-2 cells) [56, 57], stromal cells (MSCs) [58], MG-63 cells [59], and MC3T3-E1 osteoblastic cells [62]. The composites show high availability for the cells after 24 h; however, the composites are very safely to be used in bone tissue engineering.

### The characterization techniques of HA/Col composites

The prepared HA/Col composites were characterized for their chemical, physical and biological properties. The characterized techniques for the implant composites include two important things biocompatibility and morphology of the composites. Previous studied focused on the *in vivo* study [63]. The biocompatibility test include two parts; firstly, the cytotoxicity study, which give the viability of the cultured cells with the composites after cultured for kwon time. The cytotoxicity studied of HA/Col composites was determined by different techniques such as MTT assay [64], alamar blue assay [66], and flow cytometry analysis [49].

The hydroxyapatite and collagen don't have any toxic itself; moreover, the composites of HA/Col aren't toxic [45], but, there will be cause for concern when combined with other potentially toxic substances, which will lead to the toxicity of the final product. Previous studies have dealt with the study of the toxicity of composites when combined with other materials, as shows in table

5. The study of Popa *et al.*, [49], reported to combined Zn with the composites of HA/Col, the results have been shown viability for the HeLa cell lines more than 95%. The composites of HA/Col were modified for the osteoblast via incorporated with chitosan. The results did not show any toxic against MC3T3-E1 cells for the 7 days.

In the conclusion according to the previous studies, the composites of the HA/Col did not show any toxicity against human cells line; moreover, the composites have been combined with many materials, and the results did not show any toxicity; however, the composites are safe and can be used in the bone tissue engineering.

The cell attachment study of the HA/Col composites was demonstrated in wide range of the previous studies. The composites showed promising attached with the vivo cells. The HA/Col/chitosan was prepared for the restoration of maxillofacial mandible bone. Cell attachment was investigated against mesenchymal stem cells; the results have been shown more than 90% of cells were attached to the composites after 24h of the cultured [67]. The study of Cao *et al.*, [68], reported to prepared HA/Col for the application of bone graft. Study of cell attachment was determined during cultured with murine L929 cells, 75% of the cells attached with the composites after 24h. Table 6 shows summaries for the previous studies which have been mentioned to the study of cell attachment for the composites of HA/Col.

In the conclusion of this part, the composites of HA/Col have been cultured with the human cell line for the known duration time, and the results were demonstrated to high biocompatibility with the human cells line. According to the previous study, the composites of HA/Col is suitable to use in bone tissue engineering due to its high biocompatibility.

The second part of the characterization techniques are chemical, physical and biological scan for the composites. Many techniques have been used to confirm this part, such as SEM, EDX, XRD, XPS, FTIR, AFM, antibacterial, anticancer and swelling ratio, etc., as shows in fig. 2. The chemical composition of the HA/Col composites has

been confirmed by using XRD, EDX, XPS, and FTIR.

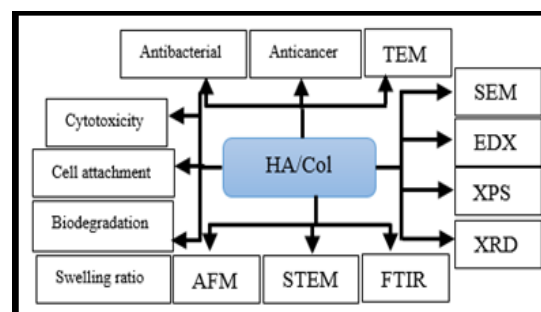


Fig. 2: Characterization techniques of HA/Col composites (Source: author)

The characterization techniques of HA/Col composites aimed to identify the chemical composition, surface morphology, mechanical properties, and biological ability for the prepared composites. The chemical composition was conformed by using XRD [69, 70], FTIR [39], EDS [71] and XPS. The XRD was used to determine crystal, size of the particle and shape. The XPS and EDS aimed to confirm the elements that possible to be present. The FTIR analysis has been used to investigate the chemical groups which is constituted the structure of the composites [74, 75], as shows in table 7.

The morphology of the HA/Col composites was determined by using TEM, AFM and SEM. These instruments were used to identify the homogeneous of the surface sizes and shapes of the particles. The biological properties of the composites were determined by using many assays such as antibacterial, anticancer, degradation, swelling ratio and cell attachment. The results of the previous studies showed good ability for the composites to be used as osteoconductivity and biocompatibility [69, 72]; table 7 shows more details about them.

Table 6: Cytotoxicity and cell attachment of HA/Col composites

No	Composites	Test	Cells type	Results	Application	Ref.
1	HA/Col/Zn	Flow cytometry analysis	HeLa cell lines	More than 95%.	Bone regeneration	[49]
2	Nano-HA/Col	Cytotoxicity (MTT)	Fibroblast L929 cell line	85-100 %	Cytocompatibility	[66]
3	HA/Col	Cell attachment, <i>In vitro</i> cytotoxicity	Murine L929 cells	75% of the cells attached with the composites.	Bone graft	[67]
4	Nano-HA/Col	Cell Counting Kit-8 (CCK8), flow cytometry, Cell adhesion	MC3T3-E1	80-100 % cell viability after 3 d.	Osteogenesis	[68]
5	HA/Col/chitosan	Cytotoxicity tests, attachment cells	Mesenchymal stem cells	The concentration less than 1 mg doesn't has any toxic. More than 90% of cells were attached to the composites.	Restoration of maxillofacial mandible bone	[69]
6	Nano-HA/Col/chitosan	Cytotoxicity (CCK-8 assay)	MC3T3-E1	The MC3T3-E1 cells were grow normally during the 7 d that gave advantage to be non-toxic materials.	Biocompatibility and Osteoblast	[70]
7	HA/Col/calcium phosphate (CaP)	MTT	hMSCs	The MTT assay for the composites didn't show any toxic. However, the present of CaP did not effect on the final products.	Tissue engineering	[71]
8	HA/col/pectin	MTT assay	MEF-WT cells	The viability of the MEF-WT cells after 7 d were from 83-93%, the results have been shown there is not any toxic for the prepared composites	Bone replacement	[72]
9	HA/Col	Cell attachment	MG-63 cells	The attachment study for the HA/Col composites was investigated for duration time from 1-7 d, the results have been shown good attached to the cell in the SEM image.	Bone tissue engineering	[73]
10	HA/Col	Cell attachment	Mesenchymal stem cells	SEM image shown excellent attachment for the cells with the components of the composite	Bone regeneration	[74]

Table 7: Characterization techniques of HA/Col composites

No	Composites	Chemical composition	Morphology	Biological/Medical/Application	Ref.
1	HA/Col	EDS and XRD were used to determine the chemical composition of the composites. The EDS shows to present of an element, Ca, P, O, Cu, and Al. The ratio of Ca/P was 1.5. XRD analysis conform to present all standard peaks of functional groups for HA and Col.	Study of the morphology has been done by using FESEM, the SEM image showed a homogeneous surface with a rod-like shape for the composites.	Bone tissue engineering	[75]
2	HA/Col	XRD and FTIR were used to investigate the chemical composition of HA/Col composites. XRD was conformed to present of functional groups of HA and col. The FTIR shows major peaks at 3000 cm <sup>-1</sup> belong to Amide I group. The Po <sub>3</sub> and OH were conformed to present as well.	This study did not mentioned for the morphology tests.	MTT assay have been used to determine the cytotoxicity of the prepared composites; results showed 100% percentage of the cell to present after 24h cultured. Osteoconductivity And Biocompatibility have been confirmed.	[42]
3	HA/Col	XRD and FTIR were used to conform all functional groups of HA and Col. The results showed to present for all major groups that was correspond to HA and Col.	SEM has used to determine the morphology. The SEM image showed very well distributing for HA in the matrix of Col. The shape of composites have been confirmed as a plate-like shape.	In the biology part the author mentioned to study of swelling and degradation ratios. The results show swelling ratios were from 250% to 650%, and the stability of the composites after 7 d was 80%, while it's degraded to 50% after 28 d.	[76]
4	HA/Col	XRD showed crystal structure for the composite according to sharp peaks of XRD spectrum. FTIR showed all functional groups of HA and Col.	TEM and SEM were used to conform the morphology of the composites. Results shows nano size from 200-400 nm with the a homogeneous surface	Degradation study of the prepared composites showed slow degraded.	[77]
5	HA/Col	FTIR, EDS, and XRD were used to analysis the chemical structure of the composites. The results have been shown the chemical groups of the HA and Col in the FTIR, while the XRD showed the major peaks corresponding for both of them.	The SEM image showed spherical shape for the HA particles and very well distribution in the matrix of Col.	The cell attachment with MC3T3-E1 cells, the SEM images showed very well attached to the cell with composites.	[78]
6	HA/Col/chitosan/carbon	XRD, FTIR, and EDS aimed to analysis the chemical structure of the composites.	SEM image showed good pore size with high porosity (98 ± 0.15 to 95.7 ± 0.1%).	The MTT assay used to analysis the cytotoxicity of prepared composites. The results showed non-toxic for the composites.	[79]

### The applications of HA/Col composites

The micro and Nano HA/col scaffold developed for the applications of bone tissue engineering due to the pure Col showed unwanted foreign body reactions [79, 80, 81]. HA has been developed from tri-calcium phosphate (TCP) cement by modifying the particle size of the starting cement powder, which is then precipitated in the solution of Col. The emulsification technique has been used to fabricate the micro-carriers

of the ceramic slurry. The material has shown a synergistic effect that leads to enhanced differentiation and proliferation of cells. The scaffold of HA/Col that has been developed to be used is coated with titanium (Ti), then implants to improve the osseointegration [82]. The group that was coated had been created (Ti-6Al-4V) using plasma technique spraying with HA, and was dropped into the collagen solution. The Co/HA scaffold has been implanted in the muscles of rabbits and has shown high improved osteogenesis [83, 84].

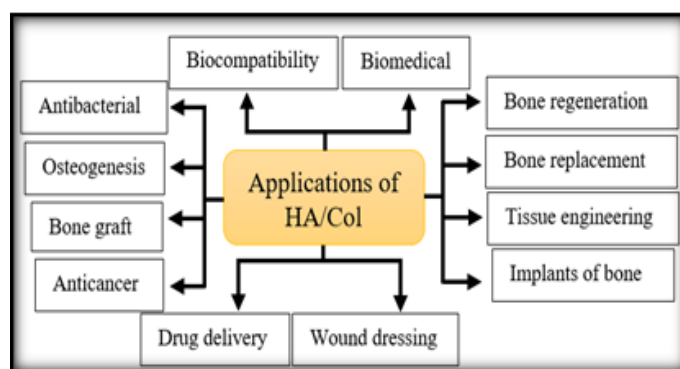


Fig. 3: Applications of HA/Col composites (Source: author)

Fig. 3 shows examples for the applications which have been mentioned in the previous studies. Among them, there were many applications that have been mentioned, such as bone regeneration [49], bone graft [67], osteogenesis [70] and biocompatibility [58], etc. in conclusion, the HA/Col composites possesses unique properties that enable it to be used in wide fields such as medical, biological and industrial fields.

### Sources of HA/Col composites

The composites of HA/Col were fabricated for the defects of bone, and Col was obtained from different sources such as porcine dermal [85], bovine serosa [86], calf hide [87], rat tails [89], bovine skin, horse tendon [88, 90], while the most of HA was from natural [65] and synthetic [91] sources.

**Table 8: Shows the sources of Col and HA from different composite formulation**

The scaffolds	HA sources	Collagen sources	Form of scaffolds	Ref
HA/Col nanocomposite	Synthetics	porcine dermal	Paste	[92]
HA/Col	Synthetics	bovine serosa	Solid	[93]
HA/Col	Synthetics	bovine femur	Spongy	[94]
HA/Col	Synthetics	calf hide	Fiber	[95]
HA/Col	Synthetics	bovine skin	Solid	[96]
HA/Col	Synthetics	calf skin	Solid	[97]
HA/Col	Synthetics	mineralized blend	cylindrical shape	[98]
HA/Col	Synthetics	bovine tendon	Solid	[99]
HA/Col	Synthetics	Bovine	Solid	[100]
Col/HA/pectin	Synthetics	Rabbit skin	cylindrical	[101]
Col/HA Nano composites	Synthetics	calf skin	Fiber	[102]
Col/HA	Synthetics	rat tails	Hydrogel	[103]
Col/HA	Synthetics	bovine tendon	lyophilized	[105]
Col/HA	Synthetics	rat tail tendons	Solid	[106]
Col/HA	Synthetics	rat tail tendons	Solid	[107]
HA/Col/Calcium phosphate	Synthetics	bovine skin	Solid	[108]
HA/Col/polycaprolactone	Synthetics	Bovine	Fibers	[109]
HA/Col/Fe	Synthetics	horse tendon	Solid	[110]
Han/Col	Nano-powder	tail tendons rats	lyophilized	[111]
HA/Col	Micro-powder	Rat	Gel	[112]
HA/Col	Synthetics	tail tendon	Solid	[113]
Col/HA/Cisplatin	Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O, and NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	calf hide	Solid	[114]
Col/HA	Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O, and NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	calf hide	Solid	[115]
Col/HA	Synthetics	bovine tendon	Solid	[116]
Col/HA/PLCL	Sigma-Aldrich	Sigma-Aldrich	Solid	[117]
PVA-Col-HA	Berkeley, CA, USA	rat tail	Fibers	[118]
Col/HA Nanocomposite	Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O, and NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	Porcine	Solid	[119]
Col/HA	Synthetics	calf skin	lyophilized	[120]
Col/HA/PVA	Synthetics	rat-tail	Hydrogel	[121]
Col/HA/hyaluronic acid	Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O, and NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	bovine Achilles tendon	Solid	[122]
Col/HA	Synthetics	Calf Hides	Solid	[123]
Col/HA/Chitosan	Synthetics	bovine tendon	Solid	[124]
silica/Col/HA	Synthetics	Bovine	hydrogels	[125]
Col/HA/PLCL	Synthetics	Purchase from Fisher Scientific	Solid	[126]
Col/nHA/PVA	Purchase from aap Implantate AG, Germany	Rat tail	Fibers	[127]
Col\HA\ PLLA	purchased from PURAC	bovine tendon	Solid	[128]
Col/HA nano/Chitosan composite	Synthetics	bovine dermis	Fibers	[129]
Col/HA	Plasma Biota Limited	Fetal calf skin	Solid	[130]
Col/HA	Synthesized	calf hide	Solid	[131]
Col/HA	Sigma-Aldrich	tail tendons of rats	Spongy	[132]
Col/HA	Synthesized	calf hides	Solid	[133]
Col/HA	purchased from Merck	bull skin	Gel	[34]

### CONCLUSION

HA/Col composites were investigated as a functional biomaterial, which was used in many medical applications; the composites that were mentioned in this section were fabricated from various sources. The previous studies demonstrated for many sources for the HA and Col; the wide source for hydroxyapatite was synthetic, while the col was isolated from an animal source. In conclusion, Accordingly, previous studies have been documented to prepare composites of hydroxyapatite/collagen (HA/Col) from synthetic and natural sources. The majority of these studies were using hydroxyapatite from synthetic sources of chemicals that may cause toxicity when applied in the future. Col was used from natural sources such as porcine dermal, bovine skin, rat tails and bovine tendon. However, the recent problem of animal diseases such as Hyaline Membrane Disease (HMD) and mad cow disease made animal-based HA not a good alternative of HA. A lot of HA/Col

composites were used in tissue engineering applications due to the higher properties that can be obtained from them. Despite the exceptional and harmonic properties of these substances and the way they are synthesized as compounds for the medical and biological fields of many prosthetic processes to enhance the growth and cohesion of the bones, many operations have failed due to bacterial infection.

### FUNDING

Nil

### AUTHORS CONTRIBUTIONS

All the authors have contributed equally.

### CONFLICT OF INTERESTS

Declared none

## REFERENCES

- Gupta A, Thussbas C, Koch M, Seebauer L. Management of glenoid bone defects with reverse shoulder arthroplasty-surgical technique and clinical outcomes. *J Shoulder Elbow Surg.* 2018;27(5):853-62. doi: 10.1016/j.jse.2017.10.004, PMID 29306665.
- Qian Y, Zhou X, Sun H, Yang J, Chen Y, Li C, Wang H, Xing T, Zhang F, Gu N. Biomimetic domain-active electrospun scaffolds facilitating bone regeneration synergistically with antibacterial efficacy for bone defects. *ACS Appl Mater Interfaces.* 2018;10(4):3248-59. doi: 10.1021/acsami.7b14524, PMID 29172421.
- Tampieri A, Iafisco M, Sandri M, Panseri S, Cunha C, Sprio S, Savini E, Uhlarz M, Herrmannsdörfer T. Magnetic bioinspired hybrid nanostructured collagen-hydroxyapatite scaffolds supporting cell proliferation and tuning regenerative process. *ACS Appl Mater Interfaces.* 2014;6(18):15697-707. doi: 10.1021/am5050967, PMID 25188781.
- Zhu L, Luo D, Liu Y. Effect of the Nano/microscale structure of biomaterial scaffolds on bone regeneration. *Int J Oral Sci.* 2020;12(1):6. doi: 10.1038/s41368-020-0073-y, PMID 32024822.
- Filippi M, Born G, Chaaban M, Scherberich A. Natural polymeric scaffolds in bone regeneration. *Front Bioeng Biotechnol.* 2020;8(8):474. doi: 10.3389/fbioe.2020.00474, PMID 32509754.
- Hasan MR, Mohd Yasin NS, Mohd Ghazali MS, Mohtar NF. Proximate and morphological characteristics of nano hydroxyapatite (Nano HAP) extracted from the fish bone. *J Sustain Sci Manage.* 2020;15(8):9-21. doi: 10.46754/jssm.2020.12.002.
- Majhool AA, Zainol I, Jaafar CN, Ha A, Hassan MZ, Mudhafar M, Majhool AA, Asaad A. A brief review on biomedical applications of hydroxyapatite use as fillers in polymer. *J Chem* 2019;13:112-9.
- Parisi C, Salvatore L, Veschini L, Serra MP, Hobbs C, Madaghiele M, Sannino A, Di Silvio L. Biomimetic gradient scaffold of collagen-hydroxyapatite for osteochondral regeneration. *J Tissue Eng.* 2020;11:2041731419896068. doi: 10.1177/2041731419896068, PMID 35003613.
- Song Y, Wu H, Gao Y, Li J, Lin K, Liu B, Lei X, Cheng P, Zhang S, Wang Y, Sun J, Bi L, Pei G. Zinc silicate/nano-hydroxyapatite/collagen scaffolds promote angiogenesis and bone regeneration via the p38 MAPK pathway in activated monocytes. *ACS Appl Mater Interfaces.* 2020;12(14):16058-75. doi: 10.1021/acsami.0c00470, PMID 32182418.
- Chen L, Hu J, Ran J, Shen X, Tong H. Synthesis and cytocompatibility of collagen/hydroxyapatite nanocomposite scaffold for bone tissue engineering. *Polym Compos.* 2016;37(1):81-90. doi: 10.1002/pc.23157.
- Andronesu E, Ficaï M, Voicu G, Ficaï D, Maganu M, Ficaï A. Synthesis and characterization of collagen/hydroxyapatite: magnetite composite material for bone cancer treatment. *J Mater Sci Mater Med.* 2010;21(7):2237-42. doi: 10.1007/s10856-010-4076-7, PMID 20372983.
- Teng SH, Lee EJ, Wang P, Kim HE. Collagen/hydroxyapatite composite nanofibers by electrospinning. *Mater Lett.* 2008;62(17-18):3055-8. doi: 10.1016/j.matlet.2008.01.104.
- Mudhafar M, Zainol I, Jaafar CN, Alsailawi HA, Majhool AA. Microwave-assisted green synthesis of Ag nanoparticles using leaves of *Melia dubia* (Neem) and its antibacterial activities. *J Adv Res Fluid Mech Therm Sci.* 2020;65(1):121-9.
- Zhang Z, Ma Z, Zhang Y, Chen F, Zhou Y, An Q. Dehydrothermally crosslinked collagen/hydroxyapatite composite for enhanced *in vivo* bone repair. *Colloids Surf B Biointerfaces.* 2018;163:394-401. doi: 10.1016/j.colsurfb.2018.01.011, PMID 29366982.
- Ficaï A, Andronesu E, Trandafir V, Ghitulica C, Voicu G. Collagen/hydroxyapatite composite obtained by electric field orientation. *Mater Lett.* 2010;64(4):541-4. doi: 10.1016/j.matlet.2009.11.070.
- Ficaï A, Andronesu E, Voicu G, Ghitulica C, Vasile BS, Ficaï D, Trandafir V. Self-assembled collagen/hydroxyapatite composite materials. *Chem Eng J.* 2010;160(2):794-800. doi: 10.1016/j.cej.2010.03.088.
- Pelin IM, Maier SS, Chitanu GC, Bulacovschi V. Preparation and characterization of a hydroxyapatite-collagen composite as a component for injectable bone substitute. *Mater Sci Eng C.* 2009;29(7):2188-94. doi: 10.1016/j.msec.2009.04.021.
- Chang SH, Hsu YM, Wang YJ, Tsao YP, Tung KY, Wang TY. Fabrication of pre-determined shape of bone segment with collagen-hydroxyapatite scaffold and autogenous platelet-rich plasma. *Journal of Materials Science: Materials in Medicine.* 2009;20(1):23-31. doi: 10.1007/s10856-008-3507-1, PMID 18651114.
- Walsh DP, Raftery RM, Chen G, Heise A, O'Brien FJ, Cryan SA. Rapid healing of a ~~skeletal~~ bone defect using a collagen-hydroxyapatite scaffold to facilitate low dose, combinatorial growth factor delivery. *J Tissue Eng Regen Med.* 2019;13(10):1843-53. doi: 10.1002/term.2934, PMID 31306563.
- Zhou G, Liu S, Ma Y, Xu W, Meng W, Lin X, Wang W, Wang S, Zhang J. Innovative biodegradable poly (L-lactide)/collagen/hydroxyapatite composite fibrous scaffolds promote osteoblastic proliferation and differentiation. *Int J Nanomedicine.* 2017;12:7577-88. doi: 10.2147/IJN.S146679, PMID 29075116.
- Ariesanti Y, Poedjiastoeti W, Sriyanto GA, Angraini Y. Increase of fibroblast proliferation by composite membrane (polyvinyl alcohol-collagen-hydroxyapatite). In: *IEEE International Conference on Health, Instrumentation and Measurement, and Natural Sciences (InHeNce).* Vol. 14; 2021. p. 1-5.
- Thongtham N, Chai-in P, Unger O, Boonrungsiman S, Suwanton O. Fabrication of chitosan/collagen/hydroxyapatite scaffolds with encapsulated *Cissus quadrangularis* extract. *Polym Adv Technol.* 2020;31(7):1496-507. doi: 10.1002/pat.4879.
- Wei X, Zhang X, Yang Z, Li L, Sui H. Osteoinductive potential and antibacterial characteristics of collagen-coated iron oxide nanosphere containing strontium and hydroxyapatite in long term bone fractures. *Arab J Chem.* 2021;14(3). doi: 10.1016/j.arabj.2020.102984, PMID 102984.
- Pottathara YB, Vuherer T, Maver U, Kokol V. Morphological, mechanical, and *in vitro* bioactivity of gelatine/collagen/hydroxyapatite based scaffolds prepared by unidirectional freeze-casting. *Polym Test.* 2021;102. doi: 10.1016/j.polymertesting.2021.107308, PMID 107308.
- Xichan He, Xialian Fan, Wenpo Feng, Yifei Chen, Ting Guo, F Wang, Jie Liu, K Tang. Incorporation of micro fibrillated cellulose into a collagen-hydroxyapatite scaffold for bone tissue engineering. *Int J Biol Macromol.* 2018;115:385-92. doi: 10.1016/j.ijbiomac.2018.04.085, PMID 29673955.
- He X, Tang K, Li X, Wang F, Liu J, Zou F, Yang M, Li M. A porous collagen-carboxymethyl cellulose/hydroxyapatite composite for bone tissue engineering by bi-molecular template method. *Int J Biol Macromol.* 2019;137:45-53. doi: 10.1016/j.ijbiomac.2019.06.098, PMID 31220495.
- He X, Tang K, Li X, Wang F, Liu J, Zou F, Yang M, Li M. A porous collagen-carboxymethyl cellulose/hydroxyapatite composite for bone tissue engineering by bi-molecular template method. *Int J Biol Macromol.* 2019;137:45-53. doi: 10.1016/j.ijbiomac.2019.06.098, PMID 31220495.
- Kaviani A, Zebarjad SM, Javadpour S, Ayatollahi M, Bazargan-Lari R. Fabrication and characterization of low-cost freeze-gelated chitosan/collagen/hydroxyapatite hydrogel nanocomposite scaffold. *Int J Polym Anal Char.* 2019;24(3):191-203. doi: 10.1080/1023666X.2018.1562477.
- Iqbal B, Sarfaraz Z, Muhammad N, Ahmad P, Iqbal J, Khan ZUH, Gonfa G, Iqbal F, Jamal A, Rahim A. Ionic liquid as a potential solvent for preparation of collagen-alginate-hydroxyapatite beads as bone filler. *J Biomater Sci Polym Ed.* 2018;29(10):1168-84. doi: 10.1080/09205063.2018.1443604, PMID 29460709.
- Tang Y, Zhang H, Wei Q, Tang X, Zhuang W. Biocompatible chitosan-collagen-hydroxyapatite nanofibers coated with platelet-rich plasma for regenerative engineering of the rotator cuff of the shoulder. *RSC Adv.* 2019;9(46):27013-20. doi: 10.1039/C9RA03972D.
- Xing F, Chi Z, Yang R, Xu D, Cui J, Huang Y, Zhou C, Liu C. Chitin-hydroxyapatite-collagen composite scaffolds for bone



- regeneration. *Int J Biol Macromol.* 2021;184:170-80. doi: 10.1016/j.ijbiomac.2021.05.019, PMID 34052273.
32. Castro Cesena AB, Camacho Villegas TA, Lugo Fabres PH, Novitskaya EE, McKittrick J, Licea Navarro A. Effect of starch on the mechanical and *in vitro* properties of collagen-hydroxyapatite sponges for applications in dentistry. *Carbohydr Polym.* 2016;148:78-85. doi: 10.1016/j.carbpol.2016.04.056, PMID 27185118.
  33. Kwon GW, Gupta KC, Jung KH, Kang IK. Lamination of microfibrillar PLGA fabric by electrospinning a layer of collagen-hydroxyapatite composite nanofibers for bone tissue engineering. *Biomater Res.* 2017;21(1):11. doi: 10.1186/s40824-017-0097-3, PMID 28620549.
  34. Song W, Markel DC, Jin X, Shi T, Ren W. Poly(vinyl alcohol)/collagen/hydroxyapatite hydrogel: properties and *in vitro* cellular response. *J Biomed Mater Res A.* 2012;100(11):3071-9. doi: 10.1002/jbm.a.34240, PMID 22733675.
  35. Zhao H, Tang J, Zhou D, Weng Y, Qin W, Liu C, Lv S, Wang W, Zhao X. Electrospun icariin-loaded core-shell collagen, polycaprolactone, hydroxyapatite composite scaffolds for the repair of rabbit tibia bone defects. *Int J Nanomedicine.* 2020;15:3039-56. doi: 10.2147/IJN.S238800, PMID 32431500.
  36. Kozłowska J, Jundzill A, Bajek A, Bodnar M, Marszałek A, Witmanowski H, Sionkowska A. Preliminary *in vitro* and *in vivo* assessment of modified collagen/hydroxyapatite composite. *Mater Lett.* 2018;221:74-6. doi: 10.1016/j.matlet.2018.03.122.
  37. Ariesanti Y, Poedjiastoeti W, Sriyanto GA, Angraini Y. Increase of fibroblast proliferation by composite membrane (polyvinyl alcohol-collagen-hydroxyapatite). In: *IEEE International Conference on Health, Instrumentation and Measurement, and Natural Sciences (InHeNce)*. Vol. 14; 2021. p. 1-5.
  38. Ficai A, Andronescu E, Voicu G, Ficai D. *Advances in collagen/hydroxyapatite composite materials [Chapter]*; 2011.
  39. Hikmawati D, Kulsum U, Rudyardjo DI, Apsari R. Biocompatibility and osteoconductivity of scaffold porous composite collagen-hydroxyapatite based coral for bone regeneration. *Open Chem.* 2020;18(1):584-90. doi: 10.1515/chem-2020-0080.
  40. Zhou J, Xu C, Wu G, Cao X, Zhang L, Zhai Z, Zheng Z, Chen X, Wang Y. *In vitro* generation of osteochondral differentiation of human marrow mesenchymal stem cells in novel collagen-hydroxyapatite layered scaffolds. *Acta Biomaterialia.* 2011;7(11):3999-4006. doi: 10.1016/j.actbio.2011.06.040, PMID 21757035.
  41. Chen LY, Cui YW, Zhang LC. Recent development in beta titanium alloys for biomedical applications. *Metals.* 2020;10(9):1139. doi: 10.3390/met10091139.
  42. Perse M, Veceric Haler Z. Cisplatin-induced rodent model of kidney injury: characteristics and challenges. *BioMed Res Int.* 2018;2018:1462802. doi: 10.1155/2018/1462802, PMID 30276200.
  43. Heinemann S, Heinemann C, Jager M, Neunzehn J, Wiesmann HP, Hanke T. Effect of silica and hydroxyapatite mineralization on the mechanical properties and the biocompatibility of nanocomposite collagen scaffolds. *ACS applied materials and interfaces.* 2011;3(11):4323-31.
  44. Ciobanu G, Harja M. Cerium-doped hydroxyapatite/collagen coatings on titanium for bone implants. *Ceram Int.* 2019;45(2):2852-7. doi: 10.1016/j.ceramint.2018.07.290.
  45. Popa CL, Bartha CM, Albu M, Guegan R, Motelica Heino M, Chifiriuc CM, Bleotu C, Badea ML, Antohe S. Synthesis, characterization and cytotoxicity evaluation on zinc doped hydroxyapatite in collagen matrix. *Dig J Nanomater Biostruct.* 2015;10:681-91.
  46. Mondal S, Hoang G, Manivasagan P, Moorthy MS, Vy Phan TT, Kim HH, Nguyen TP, Oh J. Rapid microwave-assisted synthesis of gold-loaded hydroxyapatite collagen nano-bio materials for drug delivery and tissue engineering application. *Ceram Int.* 2019;45(3):2977-88. doi: 10.1016/j.ceramint.2018.10.016.
  47. Rasool I, Singh A. *In vitro* studies of biomaterial device" hydroxyapatite" prepared from different routes for biomedical applications *in vitro*. *Asian J Pharm Clin Res.* 2018;11(10):493-7. doi: 10.22159/ajpcr.2018.v11i10.27452.
  48. Predoi D, Iconaru SL, Albu M, Petre CC, Jiga G. Physicochemical and antimicrobial properties of silver-doped hydroxyapatite collagen biocomposite. *Polym Eng Sci.* 2017;57(6):537-45. doi: 10.1002/pen.24553.
  49. Sionkowska A, Kozłowska J. Characterization of collagen/hydroxyapatite composite sponges as a potential bone substitute. *International Journal of Biological Macromolecules.* 2010;47(4):483-7. doi: 10.1016/j.ijbiomac.2010.07.002, PMID 20637799.
  50. Socrates R, Sakthivel N, Rajaram A, Ramamoorthy U, Kalkura SN. Novel fibrillar collagen-hydroxyapatite matrices loaded with silver nanoparticles for orthopedic application. *Mater Lett.* 2015;161:759-62. doi: 10.1016/j.matlet.2015.09.089.
  51. Gleeson JP, Plunkett NA, O'Brien FJ. Addition of hydroxyapatite improves the stiffness, interconnectivity and osteogenic potential of a highly porous collagen-based scaffold for bone tissue regeneration. *Eur Cell Mater.* 2010;20(218):30218-30. doi: 10.22203/ecm.v020a18, PMID 20922667.
  52. Yılmaz E, Cakıroglu B, Gokce A, Findik F, Gulsoy HO, Gulsoy N, Mutlu O, Ozacar M. Novel hydroxyapatite/graphene oxide/collagen bioactive composite coating on Ti16Nb alloys by electrodeposition. *Mater Sci Eng C Mater Biol Appl.* 2019;101:292-305. doi: 10.1016/j.msec.2019.03.078, PMID 31029323.
  53. Zhang Y, Reddy VJ, Wong SY, Li X, Su B, Ramakrishna S, Lim CT. Enhanced biomineralization in osteoblasts on a novel electrospun biocomposite nanofibrous substrate of hydroxyapatite/collagen/chitosan. *Tissue Engineering Part A.* 2010;16(6):1949-60. doi: 10.1089/ten.TEA.2009.0221, PMID 20088700.
  54. Wang QQ, Ma N, Jiang B, Gu ZW, Yang BC. Preparation of a HA/collagen film on a bioactive titanium surface by the electrochemical deposition method. *Biomed Mater.* 2011;6(5):055009. doi: 10.1088/1748-6041/6/5/055009.
  55. Watanabe K, Nishio Y, Makiura R, Nakahira A, Kojima C. Paclitaxel-loaded hydroxyapatite/collagen hybrid gels as drug delivery systems for metastatic cancer cells. *Int J Pharm.* 2013;446(1-2):81-6. doi: 10.1016/j.ijpharm.2013.02.002, PMID 23402979.
  56. Suchy T, Supova M, Sauerova P, Hubalek Kalbacova MH, Klapkova E, Pokorny M, Horny L, Zavora J, Ballay R, Denk F, Sojka M, Vistejnova L. Evaluation of collagen/hydroxyapatite electrospun layers loaded with vancomycin, gentamicin and their combination: comparison of release kinetics, antimicrobial activity and cytocompatibility. *European Journal of Pharmaceutics and Biopharmaceutics.* 2019;140:50-9. doi: 10.1016/j.ejpb.2019.04.021, PMID 31055065.
  57. Mudhafar M, Alsailawi H, Abdulrasool M, Jawad Rk, Mmays A. Mini-review of phytochemicals of ten ficus species. *International Journal Of Chemistry Research.* 2021;1:7-18.
  58. Andronescu E, Ficai A, Albu MG, Mitran V, Sonmez M, Ficai D, Ion R, Cimpean A. Collagen-hydroxyapatite/cisplatin drug delivery systems for locoregional treatment of bone cancer. *Technology in Cancer Research and Treatment.* 2013;12(4):275-84.
  59. Suchy T, Supova M, Klapkova E, Adamkova V, Zavora J, Zaloudkova M, Ryglova S, Ballay R, Denk F, Pokorny M, Sauerova P, Hubalek Kalbacova M, Horny L, Vesely J, Vonavkova T, Prusa R. The release kinetics, antimicrobial activity and cytocompatibility of differently prepared collagen/hydroxyapatite/vancomycin layers: microstructure vs. nanostructure. *Eur J Pharm Sci.* 2017;100:219-29. doi: 10.1016/j.ejps.2017.01.032, PMID 28132822.
  60. Lian X, Liu H, Wang X, Xu S, Cui F, Bai X. Antibacterial and biocompatible properties of vancomycin-loaded nano-hydroxyapatite/collagen/poly (lactic acid) bone substitute. *Progress in Natural Science: Materials International.* 2013;23(6):549-56. doi: 10.1016/j.pnsc.2013.11.003.
  61. Mudhafar M, Zainol I, Jaafar CN, Alsailawi HA, Desa S. A review study on synthesis methods of Ag Nanoparticles, considering antibacterial property and cytotoxicity. *International Journal of Drug Delivery Technology.* 2021;11(2):635-48.
  62. Majhooll AA, Zainol I, Jaafar CN, Mudhafar M, Ha A, Asaad A, Mezaal FW. Preparation of fish scales hydroxyapatite (FShAp) for potential use as fillers in polymer. *J Chem.* 2019;13:97-104.



63. Joy DP, Kumar KK, Kumar SK, KSS. Collagen from squid and its biological activity. *Int J Curr Pharm Res.* 2017;9(3):24-6.
64. Rusu LC, Nedelcu IA, Georgiana Albu M, Sonmez M, Voicu G, Radulescu M, Ficai D, Ficai A, Negrutiu ML, Sinescu C. Tetracycline loaded collagen/hydroxyapatite composite materials for biomedical applications. *Journal of Nanomaterials.* 2015;2015:1-5. doi: 10.1155/2015/361969.
65. Oshima S, Sato T, Honda M, Suetsugu Y, Ozeki K, Kikuchi M. Fabrication of gentamicin-loaded hydroxyapatite/collagen bone-like nanocomposite for anti-infection bone void fillers. *International J Molecular Sciences.* 2020;21(2):551. doi: 10.3390/ijms21020551, PMID 31952242.
66. Mudhafar M, Zainol I, Alsailawi HA, Aiza Jaafar CN. Synthesis and characterization of fish scales of hydroxyapatite/collagen-silver nanoparticles composites for the applications of bone filler. *Journal of the Korean Ceramic Society.* 2022;59(2):229-39. doi: 10.1007/s43207-021-00154-0.
67. Cao HD, Tan YF, Lin XY, Fan HS, Zhang XD. Cytocompatibility testing of hydroxyapatite/collagen composite cross-linked with glutaraldehyde. In *Key Eng Mater.* 2005;288:223-6.
68. Wenpo F, Gaofeng L, Shuying F, Yuanming Q, Keyong T. Preparation and characterization of collagen-hydroxyapatite/pectin composite. *International Journal of Biological Macromolecules.* 2015;74:218-23. doi: 10.1016/j.ijbiomac.2014.11.031, PMID 25485944.
69. Ficai A, Andronesu E, Voicu G, Ghitulica C, Ficai D. The influence of collagen support and ionic species on the morphology of collagen/hydroxyapatite composite materials. *Materials Characterization.* 2010 Apr 1;61(4):402-7. doi: 10.1016/j.matchar.2010.01.003.
70. Tenkumo T, Vanegas Saenz JR, Takada Y, Takahashi M, Rotan O, Sokolova V, Epple M, Sasaki K. Gene transfection of human mesenchymal stem cells with a nNano-hydroxyapatite-collagen scaffold containing DNA-functionalized calcium phosphate nanoparticles. *Genes to Cells.* 2016;21(7):682-95. doi: 10.1111/gtc.12374, PMID 27238217.
71. Yoshida T, Kikuchi M, Koyama Y, Takakuda K. Osteogenic activity of MG63 cells on bone-like hydroxyapatite/collagen nanocomposite sponges. *Journal of Materials Science: Materials in Medicine.* 2010;21(4):1263-72. doi: 10.1007/s10856-009-3938-3, PMID 19924517.
72. Zhang Y, Reddy VJ, Wong SY, Li X, Su B, Ramakrishna S, Lim CT. Enhanced biomineralization in osteoblasts on a novel electrospun biocomposite nanofibrous substrate of hydroxyapatite/collagen/chitosan. *Tissue Engineering Part A.* 2010;16(6):1949-60. doi: 10.1089/ten.TEA.2009.0221, PMID 20088700.
73. Lei X, Gao J, Xing F, Zhang Y, Ma Y, Zhang G. Comparative evaluation of the physicochemical properties of nano-hydroxyapatite/collagen and natural bone ceramic/collagen scaffolds and their osteogenesis-promoting effect on MC3T3-E1 cells. *Regenerative Biomaterials.* 2019;6(6):361-71. doi: 10.1093/rb/rbz026, PMID 31827888.
74. Huang Z, Yu B, Feng Q, Li S, Chen Y, Luo L. In situ-forming chitosan/nano-hydroxyapatite/collagen gel for the delivery of bone marrow mesenchymal stem cells. *Carbohydrate Polymers.* 2011;85(1):261-7. doi: 10.1016/j.carbpol.2011.02.029.
75. Chen Y, Huang Z, Li X, Li S, Zhou Z, Zhang Y, Yu BFeng QI, Yu B. *In vitro* biocompatibility and osteoblast differentiation of an injectable chitosan/nano-hydroxyapatite/collagen scaffold. *Journal of Nanomaterials.* 2012;2012:1-6. doi: 10.1155/2012/401084.
76. Sun TW, Zhu YJ, Chen F, Zhang YG. Ultralong hydroxyapatite nanowire/collagen bio paper with high flexibility, improved mechanical properties and excellent cellular attachment. *Chemistry-An Asian Journal.* 2017;12(6):655-64. doi: 10.1002/asia.201601592, PMID 28133927.
77. Montalbano G, Molino G, Fiorilli S, Vitale-Brovarone C. Synthesis and incorporation of rod-like nano-hydroxyapatite into type I collagen matrix: A hybrid formulation for 3D printing of bone scaffolds. *Journal of the European Ceramic Society.* 2020;40(11):3689-97. doi: 10.1016/j.jeurceramsoc.2020.02.018.
78. Takallu S, Mirzaei E, Azadi A, Karimizade A, Tavakol S. Plate-shape carbonated hydroxyapatite/collagen nanocomposite hydrogel via in situ mineralization of hydroxyapatite concurrent with gelation of collagen at pH = 7.4 and 37°C. *Journal of Biomedical Materials Research Part B: Applied Biomaterials.* 2019;107(6):1920-9. doi: 10.1002/jbm.b.34284, PMID 30467948.
79. Sionkowska A, Kozłowska J. Characterization of collagen/hydroxyapatite composite sponges as a potential bone substitute. *International Journal of Biological Macromolecules.* 2010;47(4):483-7. doi: 10.1016/j.ijbiomac.2010.07.002, PMID 20637799.
80. Marchianti ACN, Prameswari MC, Sakinah EN, Ulfa EU. The enhancement of collagen synthesis process on the diabetic wound by *Merremia mammosa* (Lour.) extract fraction. *Int J Pharm Pharm Sci.* 2019;11(2):47-50. doi: 10.22159/ijpps.2019v11i2.30170.
81. Ying RL, Sun RX, Li QQ, Fu CN, Chen KZ. Synthesis of ultralong hydroxyapatite micro/nanoribbons and their application as reinforcement in collagen scaffolds for bone regeneration. *Ceramics International.* 2019;45(5):5914-21. doi: 10.1016/j.ceramint.2018.12.059.
82. Zhao X, Li H, Xu Z, Li K, Cao S, Jiang G. Selective preparation and characterization of nano-hydroxyapatite/collagen coatings with three-dimensional network structure. *Surf Coatings Technol.* 2017;322:227-37. doi: 10.1016/j.surfcoat.2017.05.042.
83. Jones GL, Walton R, Czernuszka J, Griffiths SL, El Haj AJ, Cartmell SH. Primary human osteoblast culture on 3D porous collagen-hydroxyapatite scaffolds. *Journal of Biomedical Materials Research Part A.* 2010;94(4):1244-50. doi: 10.1002/jbm.a.32805, PMID 20694991.
84. Türk S, Altınsoy I, Celebi Efe GC, İlpek M, Ozacar M, Bindal C. 3D porous collagen/functionalized multiwalled carbon nanotube/chitosan/hydroxyapatite composite scaffolds for bone tissue engineering. *Materials Science and Engineering: C Mater Biol Appl.* 2018;92:757-68. doi: 10.1016/j.msec.2018.07.020, PMID 30184804.
85. Perez RA, Ginebra MP. Injectable collagen/ $\alpha$ -tricalcium phosphate cement: collagen-mineral phase interactions and cell response. *Journal of Materials Science: Materials in Medicine.* 2013;24(2):381-93. doi: 10.1007/s10856-012-4799-8, PMID 23104087.
86. Uezono M, Takakuda K, Kikuchi M, Suzuki S, Moriyama K. Hydroxyapatite/collagen nanocomposite-coated titanium rod for achieving rapid osseointegration onto bone surface. *Journal of Biomedical Materials Research Part B: Applied Biomaterials.* 2013;101(6):1031-8. doi: 10.1002/jbm.b.32913, PMID 23554303.
87. Huang Y, He J, Gan L, Liu X, Wu Y, Wu F, Gu ZW. Osteoconductivity and osteoinductivity of porous hydroxyapatite coatings deposited by liquid precursor plasma spraying: *in vivo* biological response study. *Biomedical Materials.* 2014;9(6):065007:065007. doi: 10.1088/1748-6041/9/6/065007.
88. Zhu L, Xie Y, Wen B, Ye M, Liu Y, Imam KMSU, Cai H, Zhang C, Wang F, Xin F. Porcine bone collagen peptides promote osteoblast proliferation and differentiation by activating the PI3K/Akt signaling pathway. *Journal of Functional Foods.* 2020;64. doi: 10.1016/j.jff.2019.103697, PMID 103697.
89. Santhanam R, Rameli MAP, Al Jeffri AA, Ismail WIW. Bovine-based collagen dressings in wound care management. *Journal of Pharmaceutical Research International.* 2020;9:48-63. doi: 10.9734/jpri/2020/v32i3330949.
90. Noorzai S, Verbeek CJR, Lay MC, Swan J. Collagen extraction from various waste bovine hide sources. *Waste Biomass Valor.* 2020;11(11):5687-98. doi: 10.1007/s12649-019-00843-2.
91. Salvatore L, Gallo N, Aiello D, Lunetti P, Barca A, Blasi L, Madaghiele M, Bettini S, Giancane G, Hasan M, Borovkov V, Natali ML, Campa L, Valli L, Capobianco L, Napoli A, Sannino A. An insight on type I collagen from horse tendon for the manufacture of implantable devices. *International J Biological Macromolecules.* 2020;154:291-306. doi: 10.1016/j.ijbiomac.2020.03.082, PMID 32173436.

92. Rittie L. Type I collagen purification from rat tail tendons. In: *Fibrosis Methods Mol Biol* 2017 .p. 287-308. doi: 10.1007/978-1-4939-7113-8\_19, PMID 28836209.
93. Carvalho AM, Marques AP, Silva TH, Reis RL. Evaluation of the potential of collagen from codfish skin as a biomaterial for biomedical applications. *Marine Drugs*. 2018;16(12):495. doi: 10.3390/md16120495, PMID 30544788.
94. Kolmas J, Krukowski S, Laskus A, Jurkitewicz M. Synthetic hydroxyapatite in pharmaceutical applications. *Ceramics International*. 2016;42(2):2472-87. doi: 10.1016/j.ceramint.2015.10.048.
95. Sato T, Kochi A, Shirotsaki Y, Hayakawa S, Aizawa M, Osaka A, Kikuchi M. Preparation of injectable hydroxyapatite/collagen paste using sodium alginate and influence of additives. *J Ceram Soc Japan*. 2013;121(1417):775-81. doi: 10.2109/jcersj2.121.775.
96. Silva CC, Thomazini D, Pinheiro AG, Aranha N, Figueiroo SD, Goes JC, Sombra ASB. Collagen-hydroxyapatite films: piezoelectric properties. *Materials Science and Engineering: B*. 2001;86(3):210-8. doi: 10.1016/S0921-5107(01)00674-2.
97. Castro Cesena AB, Camacho Villegas TA, Lugo Fabres PH, Novitskaya EE, McKittrick J, Licea Navarro A. Effect of starch on the mechanical and *in vitro* properties of collagen-hydroxyapatite sponges for applications in dentistry. *Carbohydrate Polymers*. 2016;148:78-85. doi: 10.1016/j.carbpol.2016.04.056, PMID 27185118.
98. Mudhafar M, Zainol I, Alsailawi HA, Jaafar CN, Mohammed RK, Dhahi SJ. Preparation, repair and characterization of beads of fish scales hydroxyapatite/collagen/silver nanoparticles by using infiltration method. *Malaysian Journal of Microscopy*. 2021;17(2):239-50.
99. Mederle N, Marin S, Marin MM, Danila E, Mederle O, Albu Kaya MG, Ghica MV. Innovative biomaterials based on collagen-hydroxyapatite and doxycycline for bone regeneration. *Advances in Materials Science and Engineering*. 2016;2016:1-5. doi: 10.1155/2016/3452171.
100. Liu Z, Yin X, Ye Q, He W, Ge M, Zhou X, Hu J, Zou S. Periodontal regeneration with stem cells-seeded collagen-hydroxyapatite scaffold. *Journal of Biomaterials Applications*. 2016;31(1):121-31. doi: 10.1177/0885328216637978, PMID 27009932.
101. Tapsir Z, Saidin S. Synthesis and characterization of collagen-hydroxyapatite immobilized on polydopamine grafted stainless steel. *Surf Coatings Technol*. 2016;285:11-6. doi: 10.1016/j.surfcoat.2015.11.024.
102. Calabrese G, Giuffrida R, Fabbri C, Figallo E, Lo Furno D, Gulino R, Colarossi C, Fullone F, Giuffrida R, Parenti R, Memeo L, Forte S. Collagen-hydroxyapatite scaffolds induce human adipose derived stem cells osteogenic differentiation *in vitro*. *PLoS ONE*. 2016;11(3):e0151181. doi: 10.1371/journal.pone.0151181, PMID 26982592.
103. Kane RJ, Weiss-Bilka HE, Meagher MJ, Liu Y, Gargac JA, Niebur GL, Wagner DR, Roeder RK. Hydroxyapatite reinforced collagen scaffolds with improved architecture and mechanical properties. *Acta Biomaterialia*. 2015;17:16-25. doi: 10.1016/j.actbio.2015.01.031, PMID 25644451.
104. Quinlan E, Lopez Noriega A, Thompson E, Kelly HM, Cryan SA, O'Brien FJ. Development of collagen-hydroxyapatite scaffolds incorporating PLGA and alginate microparticles for the controlled delivery of rhBMP-2 for bone tissue engineering. *Journal of Controlled Release*. 2015;198:71-9. doi: 10.1016/j.jconrel.2014.11.021, PMID 25481441.
105. Kane RJ, Weiss Bilka HE, Meagher MJ, Liu Y, Gargac JA, Niebur GL, Wagner DR, Roeder RK. Hydroxyapatite reinforced collagen scaffolds with improved architecture and mechanical properties. *Acta Biomaterialia*. 2015;17:16-25. doi: 10.1016/j.actbio.2015.01.031, PMID 25644451.
106. Zhou Y, Yao H, Wang J, Wang D, Liu Q, Li Z. Greener synthesis of electrospun collagen/hydroxyapatite composite fibers with an excellent microstructure for bone tissue engineering. *Int Journal of Nanomedicine*. 2015;10:3203-15. doi: 10.2147/IJN.S79241, PMID 25995630.
107. Mudhafar M, Alsailawi HA, Raheem HA, Mohammed RK. A review study on the biomaterials and bone grafts for bone defects applications. *European Journal of Biomedical*. 2022;9(4):63-73.
108. Bendtsen ST, Wei M. Synthesis and characterization of a novel injectable alginate-collagen-hydroxyapatite hydrogel for bone tissue regeneration. *J Mater Chem B*. 2015;3(15):3081-90. doi: 10.1039/c5tb00072f, PMID 32262508.
109. Majhool AA, Zainol I, Azziz SS, Jaafar CN, Jahil MM. Mechanical properties improvement of epoxy composites by natural hydroxyapatite from fish scales as a filler. 2018;10(2):1424-9.
110. Villa MM, Wang L, Huang J, Rowe DW, Wei M. Bone tissue engineering with collagen-hydroxyapatite scaffold and culture expanded bone marrow stromal cells. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2015;103(2):243-53. doi: 10.1002/jbm.b.33225, PMID 24909953.
111. Mudhafar M, Zainol I, Alsailawi HA, Aiza Jaafar CN. Green synthesis of silver nanoparticles using neem and collagen of fish scales as a reducing and stabilizer agents. *Jordan Journal of Biological Sciences*. 2021;14(5):899-903. doi: 10.54319/jjbs/140503.
112. Villa MM, Wang L, Rowe DW, Wei M. Effects of cell-attachment and extracellular matrix on bone formation *in vivo* in collagen-hydroxyapatite scaffolds. *PLoS ONE*. 2014;9(10):e109568. doi: 10.1371/journal.pone.0109568, PMID 25329879.
113. Prosecka E, Rampichova M, Litvinec A, Tonar Z, Kralickova M, Vojtova L, Kochova P, Plencner M, Buzgo M, Mickova A, Jancar J, Amler E. Collagen/hydroxyapatite scaffold enriched with polycaprolactone nanofibers, thrombocyte-rich solution and mesenchymal stem cells promotes regeneration in large bone defect *in vivo*. *Journal of Biomedical Materials Research Part A*. 2015;103(2):671-82. doi: 10.1002/jbm.a.35216, PMID 24838634.
114. Mudhafar M, Zainol I, Aiza Jaafar CN, Alsailawi HA, Majhool AA. Two green synthesis methods to prepared nanoparticles of Ag: two sizes and shapes via using extract of *N. dubia* leaves. *J Comput Theor Nanosci*. 2020 Jul 1;17(7):2882-9. doi: 10.1166/jctn.2020.8954.
115. Sionkowska A, Kozłowska J. Properties and modification of porous 3-D collagen/hydroxyapatite composites. *International Journal Biological Macromolecules*. 2013;52:250-9. doi: 10.1016/j.ijbiomac.2012.10.002, PMID 23063427.
116. Laydi F, Rahouadj R, Cauchois G, Stoltz JF, De Isla N. Hydroxyapatite incorporated into collagen gels for mesenchymal stem cell culture. *Biomed Mater Eng*. 2013;23(4):311-5. doi: 10.3233/BME-130755, PMID 23798652.
117. Mudhafar MU, Zainol IS. Medical values, antimicrobial, and anti-fungal activities of polyalthia genus. *International Journal of Pharmaceutical Research*. 2019;11(1):90-6.
118. Panda NN, Jonnalagadda S, Pramanik K. Development and evaluation of cross-linked collagen-hydroxyapatite scaffolds for tissue engineering. *Journal of Biomaterials Science, Polymer Edition*. 2013;24(18):2031-44. doi: 10.1080/09205063.2013.822247, PMID 23905722.
119. Antebi B, Cheng X, Harris JN, Gower LB, Chen XD, Ling J. Biomimetic collagen-hydroxyapatite composite fabricated via a novel perfusion-flow mineralization technique. *Tissue Engineering Part C: Methods*. 2013;19(7):487-96. doi: 10.1089/ten.TEC.2012.0452, PMID 23157544.
120. Alsailawi HA, Misnan R, Mudhafar M, A Lsailawi. Major and minor allergen ige reactivity of purple mud crab (*Scylla tranquebarica*) against a cross-reactive allergen in crustacean and molluscs in patients with a seafood allergy. *Research Journal of Pharmacy and Technology*. 2021;14(1):239-44. doi: 10.5958/0974-360X.2021.00042.1.
121. Akkouch A, Zhang Z, Rouabhia M. Engineering bone tissue using human dental pulp stem cells and an osteogenic collagen-hydroxyapatite-poly (L-lactide-co-ε-caprolactone) scaffold. *Journal of Biomaterials Applications*. 2014;28(6):922-36. doi: 10.1177/0885328213486705, PMID 23640860.
122. Zainurin MA, Zainol I, Mudhafar M. Biogenic synthesis of silver nanoparticles using neem leaf extract as reducing agent and hydrolyzed collagen as stabilizing agent. *Malaysian Journal of Microscopy*. 2022;18(1):215-25.

123. Al Sailawi HA, Misnan R, Yazdir ZH, Abdullah N, Bakhtiar F, Arip M, Mudhafar M, Ateshan HM. Effects of different salting and drying methods on allergenicity of purple mud crab (*Scylla tranquebarica*). *Indian Journal of Ecology*. 2020;47(4):1173-9.
124. Chang SH, Hsu YM, Wang YJ, Tsao YP, Tung KY, Wang TY. Fabrication of pre-determined shape of bone segment with collagen-hydroxyapatite scaffold and autogenous platelet-rich plasma. *Journal of Materials Science: Materials in Medicine*. 2009;20(1):23-31. doi: 10.1007/s10856-008-3507-1, PMID 18651114.
125. Zhou J, Xu C, Wu G, Cao X, Zhang L, Zhai Z, Zheng Z, Chen X, Wang Y. *In vitro* generation of osteochondral differentiation of human marrow mesenchymal stem cells in novel collagen-hydroxyapatite layered scaffolds. *Acta Biomaterialia*. 2011;7(11):3999-4006. doi: 10.1016/j.actbio.2011.06.040, PMID 21757035.
126. Mudhafar M, Zainol I, Jaafar CN, Alsailawi HA, Majhool AA, Alsaady M. Phytochemical screening and characterization of meliadubia leaves extract for antimicrobial activity against *Escherichia coli* and *Staphylococcus aureus*. *Indian Journal of Ecology*. 2020;47(2):493-6.
127. Heinemann S, Heinemann C, Jager M, Neunzehn J, Wiesmann HP, Hanke T. Effect of silica and hydroxyapatite mineralization on the mechanical properties and the biocompatibility of nanocomposite collagen scaffolds. *ACS Applied Materials and Interfaces*. 2011;3(11):4323-31.
128. Akkouch A, Zhang Z, Rouabhia M. A novel collagen/hydroxyapatite/poly (lactide-co- $\epsilon$ -caprolactone) biodegradable and bioactive 3D porous scaffold for bone regeneration. *Journal of Biomed Materials Research Part A*. 2011;96(4):693-704. doi: 10.1002/jbma.33033, PMID 21284080.
129. Asran AS, Henning S, Michler GH. Polyvinyl alcohol-collagen-hydroxyapatite biocomposite nanofibrous scaffold: mimicking the key features of natural bone at the nanoscale level. *Polymer*. 2010;51(4):868-76. doi: 10.1016/j.polymer.2009.12.046.
130. Gleeson JP, Plunkett NA, O'Brien FJ. Addition of hydroxyapatite improves stiffness, interconnectivity and osteogenic potential of a highly porous collagen-based scaffold for bone tissue regeneration. *Eur Cell Mater*. 2010;20(218):30218-30. doi: 10.22203/ecm.v020a18, PMID 20922667.
131. Zhang Y, Reddy VJ, Wong SY, Li X, Su B, Ramakrishna S, Lim CT. Enhanced biomineralization in osteoblasts on a novel electrospun biocomposite nanofibrous substrate of hydroxyapatite/collagen/chitosan. *Tissue Engineering Part A*. 2010;16(6):1949-60. doi: 10.1089/ten.TEA.2009.0221, PMID 20088700.
132. Majhool AA, Zainol I, Azziz SS, Jaafar CN, Jahil MM. Mechanical properties improvement of epoxy composites by natural hydroxyapatite from fish scales as a fillers. *International Journal of Research in Pharmaceutical Sciences* 2018;10(2):1424-9. doi: 10.26452/ijrps.v10i2.708
133. Jones GL, Walton R, Czernuszka J, Griffiths SL, El Haj AJ, Cartmell SH. Primary human osteoblast culture on 3D porous collagen-hydroxyapatite scaffolds. *Journal of Biomedical Materials Research Part A*. 2010;94(4):1244-50. doi: 10.1002/jbma.32805, PMID 20694991.