

ISSN- 0975-7058

Vol 15, Issue 3, 2023

Original Article

FORMULATION AND OPTIMIZATION OF MUCOADHESIVE BUCCAL FILM FOR NICOTINE REPLACEMENT THERAPY

AADITAYA PRAKASH 🗅, PRAKASH K. SONI 🕩, SURESH K. PASWAN* 🕩, T. R. SAINI 🕩

Industrial Pharmacy Research Lab, Department of Pharmacy, Shri G. S. Institute of Technology and Science, 23, Park Road Indore-452003 (M. P.), India

Email: skpaswan@gmail.com

Received: 24 Jan 2023, Revised and Accepted: 23 Mar 2023

ABSTRACT

Objective: Nicotine replacement therapy (NRT) is a widely used method for reducing the desire to smoke and managing withdrawal symptoms during smoking cessation efforts. The research work aims to prepare and optimize a mucoadhesive nicotine buccal film by solvent casting method to provide the rapid onset and prolonged effects of cigarette smoking.

Methods: Mucoadhesive nicotine buccal films were developed from the polymers Carbopol 934, Eudragit RLPO, and HPMC E15 by solvent casting method. The optimization of the mucoadhesive nicotine buccal film was performed using a three-factor, three-level Box-Behnken design where Carbopol 934, Eudragit RLPO, and HPMC E15 were selected as independent variables, while the swelling index, adhesion time, mucoadhesive strength, and cumulative % drug release were selected as response variables.

Results: The optimized mucoadhesive nicotine buccal film showed uniform thickness and drug content. It had a swelling index of 188.21%, adhesion time of 7 h 45 min, and mucoadhesive strength of 0.23 N. The film showed a burst release followed by a steady release of 76.55 % over 360 min. It exhibited a 2-fold enhancement in buccal mucosal permeation as compared to a lozenge.

Conclusion: The mucoadhesive nicotine buccal film prepared by the solvent casting method provides a rapid onset of action and prolonged effect for an extended period which replicates the effects of cigarette smoking. The findings show that it will reduce the frequency of administration, as a result of decreased nicotine cravings and reduced withdrawal symptoms, compared to currently available NRTs, ultimately helping individuals quit smoking.

Keywords: Mucoadhesive buccal film, Box-behnken design, Nicotine replacement therapy, Rapid onset, Prolonged release

© 2023 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (https://creativecommons.org/licenses/by/4.0/) DOI: https://dx.doi.org/10.22159/ijap.2023v15i3.47412. Journal homepage: https://innovareacademics.in/journals/index.php/ijap

INTRODUCTION

Nicotine is a naturally occurring alkaloid found in tobacco plants (*Nicotiana tabacum*) and tobacco products like cigarettes, cigars, and chewing tobacco. It acts as a stimulant, affecting the central nervous system and increasing the release of various neurotransmitters, including dopamine, norepinephrine, and acetylcholine. The increase in these neurotransmitters results in a pleasurable and euphoric sensation, leading to the development of addiction [1]. Nicotine addiction is a chronic disease caused by the prolonged use of tobacco products containing nicotine. It increases the risk of many health issues, such as cancer, heart disease, and respiratory illnesses. The addicted individuals continue to seek tobacco products despite the detrimental effects on their health. The addictive nature of nicotine makes it challenging to quit tobacco use as it leads to withdrawal symptoms [2].

Nicotine replacement therapy (NRT) is a widely used treatment for nicotine addiction to help people gradually reduce their nicotine cravings and dependence while they work on smoking cessation. NRT products contain lower doses of nicotine than tobacco products and are designed to ease withdrawal symptoms while offering a safer nicotine delivery method [3-5]. Commercially available NRT products include transdermal patches, chewing gums, lozenges, oral inhalers, and nasal sprays [6, 7]. Although NRT products are effective, they often have a slower onset of action and lower drug bioavailability than cigarette smoking. Nicotine absorption is primarily through the buccal mucosa and is excreted through the urine after being metabolized by the liver, lungs, and kidneys [8].

Buccal drug administration is a type of local or systemic drug delivery through the mucosal membrane of the cheek and gums [9]. This route of drug delivery has several advantages over traditional oral and parenteral routes, including quick and direct absorption of the drug into the systemic circulation, bypassing the liver's first-pass metabolism resulting in higher bioavailability and faster onset of the drug's action [10-13]. Furthermore, buccal administration is non-invasive, and the drug is easy to administer, which makes it more convenient for patients having difficulty in swallowing or in cases of

frequent dosing [14]. The buccal route of drug administration is commonly used for drugs with a short half-life, such as nicotine, which is rapidly metabolized and requires frequent administration to maintain therapeutic levels [15].

Buccal films are a recent development in buccal drug delivery and are a preferred option over adhesive tablets due to their flexibility and comfort. Mucoadhesive buccal films, in particular, have the advantage of adhering to the mucosa and providing prolonged drug release, which can extend the duration of the therapeutic effect [16, 17]. These films are composed of a polymeric matrix that can be optimized to control the drug release kinetics and the adhesion properties of the film to the mucosal tissue [18]. As a result, mucoadhesive buccal films may be an effective option for nicotine replacement therapy, which can provide a quick onset of action to alleviate withdrawal symptoms and prolonged release to help reduce cravings over time [19, 20].

In the current study, we aimed to formulate a mucoadhesive nicotine buccal film using a solvent casting method that can provide quick onset of action followed by a prolonged release as a promising NRT. The film was formulated using different mucoadhesive polymers, i.e., Carbopol 934, Eudragit RLPO, and HPMC E15. The optimization of the mucoadhesive nicotine buccal film was carried out by using a statistical approach of the Box-Behnken experimental design [21]. The software suggested quantities of independent variables i.e., mucoadhesive polymers that influence the response variables i.e., swelling index, adhesion time, mucoadhesive strength, and in vitro drug release study at different time intervals. To assess its efficacy in providing prolonged nicotine release and mucoadhesion to the buccal mucosa, an ex-vivo permeation study was performed. This mucoadhesive nicotine buccal film is expected to be a safer alternative for existing NRTs.

MATERIALS AND METHODS

Material

Nicotine was obtained as a gift sample from Gelnova Laboratories Pvt. Ltd. (Navi Mumbai), Carbopol 934, Eudragit RLPO, and HPMC E15 were procured from Lubrizol, Evonik, and Colorcon respectively. Ethanol was purchased from E-Merck. All other chemicals and reagents were of analytical grade.

Preparation of mucoadhesive buccal films

The mucoadhesive nicotine buccal film was prepared by the solvent casting method [22]. A weighed amount of HPMC E15 was dissolved in 10 ml ethanol: water solvent blend (1:1), and the polymeric

solution was slowly added to the ethanolic solution of carbopol 934 and eudragit RLPO (10 ml) under constant stirring, followed by the addition of 0.5 % w/w propylene glycol as a plasticizer. The pH was adjusted by sodium hydroxide, and nicotine (25 mg) was added to the above polymeric solution. A thin film was cast on a glass petri dish with help of the above solution. The film was formed by gradually drying the solvent at room temperature for 12 h. The dried films were stored in aluminium foil for further evaluation.

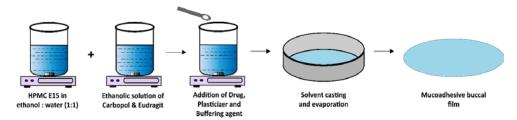


Fig. 1: Schematic diagram of a mucoadhesive buccal film prepared by solvent casting method

Optimization of mucoadhesive nicotine buccal films

The optimization was carried out to minimize the number of experimental trials. The mucoadhesive nicotine buccal films were prepared by solvent casting method. In the current study, a three-factor, three-level Box-Behnken design of response surface methodology was employed [21]. The study included the independent variables i.e., the quantity of Carbopol 934 (A), Eudragit RLPO (B), HPMC E15 (C), and the response variable selected such as swelling index (R1), adhesion time (R2), mucoadhesive strength (R3), cumulative % drug release at 5 min (R4), 15 min (R5), 30 min (R6), 60 min (R7), 120 min (R8), 240 min (R9), 360 min (R10). The

amount of drug (25 mg) and concentration of plasticizer (0.5 % w/w) were kept constant throughout the batches. The variables and their levels are recorded in table 1. The Design-Expert® software (version 12, stat-Ease, Inc., Minneapolis, MN) was used to analyze the experimental data. A 3D response surface was generated to illustrate the independent and response variables. Fifteen experimental runs were planned as per the Box-Behnken design. The ANOVA studies show the significance of the response variable (p-value of 0.05). The correlation coefficients (R²) and adjusted R² were used to assess the model's applicability. The experimental results were compared to the predicted values after the optimized formulation had been prepared.

Factors	Units	Levels		
		Low	Medium	High
A: Carbopol 934	%	1.5	2	2.5
B: Eudragit RLPO	%	1.5	2	2.5
C: HPMC E15	%	3.75	5	6.25
Response variables				
	Units	Co	onstraints	
(R1) Swelling index	%	М	aximize	
(R2) Adhesion time	Н	М	aximize	
(R3) Mucoadhesive strength	N	М	aximize	
(R4) Cumulative % drug release at 5 min	%	М	inimize	
(R5) Cumulative % drug release at 15 min	%	М	inimize	
(R6) Cumulative % drug release at 30 min	%	М	inimize	
(R7) Cumulative % drug release at 60 min	%	М	inimize	
(R8) Cumulative % drug release at 120 min	%	М	inimize	
(R9) Cumulative % drug release at 240 min	%	М	inimize	
(R10) Cumulative % drug release at 360 min	%	Μ	inimize	

Batch No.	Carbopol 934 (%)	Eudragit RLPO (%)	HPMC E 15 (%)
NBF1	2.0	2.5	3.75
NBF2	1.5	2.0	3.75
NBF3	2.0	2.0	5.00
NBF4	2.0	2.0	5.00
NBF5	2.0	2.0	5.00
NBF6	2.5	2.0	3.75
NBF7	1.5	1.5	5.00
NBF8	2.0	1.5	6.25
NBF9	2.5	1.5	5.00
NBF10	1.5	2.5	5.00
NBF11	2.5	2.0	6.25
NBF12	2.0	2.5	6.25
NBF13	2.5	2.5	5.00
NBF14	2.0	1.5	3.75
NBF15	1.5	2.0	6.25

Characterization of mucoadhesive nicotine buccal film

Determination of surface pH

The alkaline or acidic pH of the film may irritate the buccal mucosa. The objective was to keep the surface pH near neutral. The surface pH of the films was measured using a pH meter (Cyberscan 510) by placing a buccal film in a small beaker having 5 ml of water. The observations were recorded in triplicate and their mean value was determined.

Film weight and thickness

The weight and thickness of the film are essential factors in ensuring the content uniformity of the drug and excipients. The drug is very potent, so the uneven thickness and weight variation of the film may result in dose variation. Three equal sections of 2 cm x 2 cm of films were taken, weighed, and their average weight was recorded. The thickness of the film was evaluated using a digital micrometer (Digimatic micrometer, Mitutoyo, Tokyo, Japan). The film was recorded.

Folding endurance

The objective of the folding endurance test is to assess if the film can sustain the mechanical stress that occurs during its handling. The value of folding endurance was determined by repeatedly folding the film at a point, till any crack or break was observed. The mean value of three observations was recorded.

Tensile strength and extensibility

Tensile strength refers to the maximum amount of stress a material can withstand before breaking, while extensibility is the ability of a material to be stretched or extended without breaking. The tensile strength and extensibility were determined to quantify the mechanical stress induced by jaw movement in the buccal cavity. The tensile strength and extensibility of the mucoadhesive nicotine buccal film were evaluated using a texture analyzer (Stable microsystems ltd., Surrey, UK) with probe tensile grips (A/TG). In this method, the buccal film was placed between the mounting cards. The test speed was kept at 0.5 mm/s and the results were recorded.

Swelling index

A mucoadhesive polymer must be hydrated to expand and form an adequate macromolecular mesh, enabling the polymer chains to become more mobile and exposing bioadhesive sites for hydrogen bonding or electrostatic contact between the polymer and the mucosal network, thus facilitating mechanical entanglement. Mucoadhesive nicotine buccal film (2 cm x 2 cm) was weighed and then immersed in phosphate buffer pH 6.8 for a predetermined time, further it was blotted on filter paper, and weighed[23]. The following formula was used to get the percent swelling index.

$$\%$$
 SI = $\frac{W2 - W1}{W2 - W1} \times 100$

Where W1 = weight of the dry film

W2 = weight of the hydrated film

Adhesion time

The adhesion time of the mucoadhesive buccal film is determined by measuring the duration it takes for the film to separate from a stuck surface. Therefore, to increase the bioavailability and extend the retention time of the delivery system, mucoadhesive films should maintain substantial adhesive contact with the membrane. A longer adhesion time is generally desirable, as it can lead to more consistent and prolonged drug delivery. The adhesion time of mucoadhesive nicotine buccal film was determined manually by attaching the film to a moist surface to simulate buccal mucosa and the time of detachment was recorded.

Mucoadhesive strength

The mucoadhesive strength is determined by force required to detach the film from the buccal mucosa. It was measured using a texture analyzer (Stable microsystems ltd., Surrey, UK) and goat buccal mucosa as the substrate. The buccal mucosa was affixed to the stationary stage of the analyzer and a 2 cm x 2 cm film was attached to the probe. The movable probe was gradually brought

down until it made contact with the mucosa, which was maintained for 1 min at the hold position. The probe was released at a prespecified speed (0.5 mm/s) and the force required to detach the film from the mucosa was recorded.

Drug content uniformity

Drug content is an important factor in ensuring the availability and uniformity of drug in a film. A 2 cm x 2 cmpiece of the developed film was cut and placed in a beaker containing 100 ml of phosphate buffer pH 6.8. The film was allowed to dissolve, the resulting solution was filtered and analyzed using a UV spectrophotometer (Shimadzu, 1700) at 260 nm.

 $Drug \ content = \frac{Actual \ amount \ of \ drug \ in \ the \ film}{Theoretical \ amount \ of \ drug \ in \ the \ film} \ X \ 100$

In vitro drug release

The *in vitro* drug release studies of the formulated mucoadhesive nicotine buccal film were carried out using a paddle-over disc (65 mm) dissolution apparatus (Electrolab dissolution apparatus) to determine the amount of nicotine released into the medium. In the dissolution vessel, the mucoadhesive buccal film was placed beneath the disc containing 900 ml phosphate buffer pH 6.8. The test was conducted at 37 °C with a 50-rpm speed, and aliquots of 5 ml were taken out at predetermined intervals for 360 min. After being filtered through a membrane filter (0.45 μ m), an equal volume of fresh, pre-warmed (37±0.5 °C) phosphate buffer media was added to the dissolution vessel. The absorbance of aliquots was measured using a UV spectrophotometer (Shimadzu, 1700) at 260 nm, and the cumulative % drug release was calculated.

Ex-vivo adhesion time

The *ex-vivo* adhesion time of the mucoadhesive nicotine buccal film was determined by evaluating its adhesion to fresh goat buccal mucosa. The mucosa was affixed to a glass slide using adhesive. The film was adhered to buccal mucosa by pressing lightly with a fingertip after it had been moistened on one side with phosphate buffer pH 6.8. The glass slide was placed in a beaker containing 50 ml of phosphate buffer pH 6.8. The retention of the buccal film was observed after two minutes when the contents of the beaker were gently agitated to simulate the conditions of the buccal cavity. The time required for the film to detach from the mucosal surface was used to assess the mucoadhesion time [24].

Ex-vivo drug permeation

In this study, the goat buccal mucosa was used as a barrier membrane. The buccal mucosa from a freshly sacrificed goat was purchased from a nearby slaughterhouse and rinsed in isotonic phosphate buffer pH 6.8. The goat buccal mucosa (total exposed area 2.54 cm²) was mounted on a diffusion cell between the donor and receptor compartment. The mucoadhesive film then adhered to the goat buccal mucosa. The receptor compartment was filled with phosphate buffer pH 6.8 and maintained at a temperature of 37±0.5 °C. The buffer solution was constantly stirred using a magnetic stirrer set at a low speed of 50±5 rpm. Aliquots of 5 ml were taken at predetermined intervals for 360 min, filtered through a 0.45 µm membrane filter, and analyzed using a UV spectrophotometer (Shimadzu, 1700) at 260 nm. Pre-warmed (37±0.5 °C) dissolution medium was added to the diffusion cell after each sample withdrawal. The experiment was carried out in triplicate (n=3), and the mean value was calculated for the determination of *ex-vivo* drug permeation.

RESULTS AND DISCUSSION

Optimization of mucoadhesive nicotine buccal film

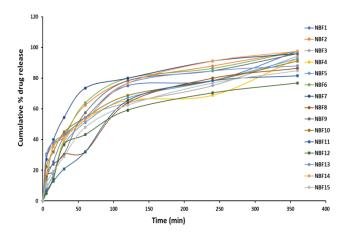
The optimization of mucoadhesive nicotine buccal film was performed using box behnken response surface design with three independent variables such as carbopol 934 (A), Eudragit RLPO (B), and HPMC E15 (C). The ranges of independent variables were selected for optimization were 1.5 %-2.5 % of carbopol 934, 1.5 %-2.5 % of eudragit RLPO and 3.75 %-6.25 % of HPMC E15. The swelling index (R1), adhesion time (R2), mucoadhesive strength (R3), cumulative % drug release at 5 min (R4), 15 min (R5), 30 min (R6), 60 min (R7), 120 min (R8), 240 min (R9), 360 min (R10) of prepared mucoadhesive

nicotine buccal film were rendered as response variables, and the effects of independent variables studied are shown in table 4.

It was found that the swelling index increased with an increase in the concentration of HPMC E15 and carbopol 934 but there was no significant effect of eudragit RLPO as shown in fig. 3 and fig. 4. It is because HPMC E15 and carbopol 934 are hydrophilic polymers that absorb water from the surrounding environment, increasing the size and volume of the film which results in slower drug release [25]. It was found that the adhesion time increased with an increase in the concentration of HPMC E15 and carbopol 934 (fig. 5 and fig. 6). The HPMC E15 forms a gel in an aqueous environment that adher to mucosal membrane for a longer period, whereas carbopol 934 adhere to the mucosal membrane due to increased surface charge of the film, the concentration of eudragit RLPO showed no substantial effect on adhesion of film [26]. It was observed that the mucoadhesive strength increased with an increase in the concentration of HPMC E15 and concentration of HPMC E15 and carbopol 934 (fig. 7 and fig. 8). The HPMC E15 increases the

mechanical strength and flexibility of the film, while carbopol 934 shows better interaction with mucosal tissue due to increased surface charge creating a stronger bond, [27]. It was observed that the concentration of carbopol 934, eudragit RLPO, and HPMC E15 affected the cumulative % drug release (R4-R10) at 5 min, 15 min, 30 min, 60 min, 120 min, 240 min, and 360 min respectively. The drug release decreased with an increase in the concentration of the polymers (fig. 9 to fig. 22). The HPMC E15 forms a matrix with the drug, prolonging its release [25, 27]. The carbopol 934 and eudragit RLPO are pH-sensitive polymers due which the release of nicotine was slightly decreased with an increase in their concentration [26, 28]. The initial burst release of the mucoadhesive buccal film varied depending on the viscosity and solubility of the polymers, while the prolonged release was solely determined by the quantity of polymers used [29]. Therefore, the optimization process helped in identifying the optimal concentration of polymers that provide the desired properties of the mucoadhesive buccal film as shown in table 6.

Response variables	Polynomial equation
Swelling index (R1)	182.18-2.94A-1.21B+21.05C-0.4075AB+7.82AC+0.9150BC+0.0663A ² -2.55B ² -10.77C ²
Adhesion time (R2)	7.30+0.1250A+0.0937B+0.4063C
Mucoadhesive strength (R3)	0.1669+0.0080A+0.0012B+0.0631C+0.0012AB+0.0000AC+0.0012BC-
	$0.0013A^2 + 0.0024B^2 + 0.0061C^2$
Cumulative % drug release at 5 min (R4)	6.73-2.79A-0.7958B-3.98C+4.38AB+2.79AC-1.99BC+9.35A ² +8.55B ² -1.79C ²
Cumulative % drug release at 15 min (R5)	15.95-2.00A-0.9993B-8.58C+5.59AB-1.58AC+1.18BC+10.20A ² +7.41B ² +0.1890C ²
Cumulative % drug release at 30 min (R6)	38.28-4.39A+1.19B-6.41C+5.60AB-1.60AC+1.60BC+0.4540A ² +4.42B ² -5.17C ²
Cumulative % drug release at 60 min (R7)	52.72-3.82A-1.19B-9.77C+4.01AB+2.40BC+6.56A ² +0.6321B ² -7.80C ²
Cumulative % drug release at 120 min (R8)	65.54+1.37A-2.39B-7.41C+2.40AB+0.3737AC-0.7825BC+6.60A ² +3.79B ² -0.6428C ²
Cumulative % drug release at 240 min (R9)	74.10+0.0080A-4.39B-4.82C+3.20AB+0.7980AC+0.7915BC+6.53A ² +4.13B ² +1.72C ²
Cumulative % drug release at 360 min (R10)	93.25-0.7959A-2.81B-7.59C-0.7777AB-0.7915AC-2.38BC-0.1688A ² -0.6405B ² -3.04C ²



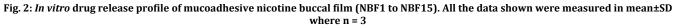
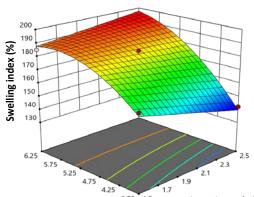


Table 4: Observed response of the optimization batches of mucoadhesive nicotine buccal film produced by Box-Behnken design

Batch	Swelling	Adhesion	Mucoadhesive	Cumulative 9	% drug release					
No.	index (R1)	time (R2)	strength (R3)	5 min (R4)	15 min	30 min	60 min	120 min	240 min	360 min
					(R5)	(R6)	(R7)	(R8)	(R9)	(R10)
NBF1	144.11±0.94	7.00±0.23	0.11±0.01	14.32±0.12	25.54±0.19	41.52±0.17	54.34±0.09	75.10±0.10	78.40±0.08	97.51±0.10
NBF2	163.69±1.41	6.75±0.13	0.09±0.01	27.05±0.06	36.75±0.09	43.17±0.09	62.30±0.18	78.32±0.20	87.96±0.12	97.56±0.10
NBF3	181.82±1.89	7.25±0.15	0.16±0.01	4.77±0.10	15.94±0.14	38.28±0.18	54.32±0.34	63.96±0.32	75.15±0.44	94.31±0.30
NBF4	180.17±1.41	7.25±0.09	0.16±0.01	6.36±0.23	14.35±0.27	39.86±0.36	52.74±0.23	65.54±0.39	68.79±0.59	92.68±0.38
NBF5	184.55±1.38	7.25±0.13	0.16±0.02	7.95±0.73	17.55±0.43	36.70±0.56	51.13±0.63	67.12±0.56	78.35±0.56	92.74±0.55
NBF6	142.22±1.82	7.00±0.09	0.11±0.01	11.14±0.69	36.66±0.71	41.58±0.53	63.80±0.68	79.92±0.80	86.38±0.51	97.56±0.65
NBF7	184.40±0.84	7.25±0.22	0.16±0.01	27.05±0.65	39.38±0.71	54.33±0.49	73.51±0.57	79.98±0.61	91.15±0.44	95.99±0.64
NBF8	191.79±1.22	7.50±0.30	0.23±0.02	15.19±0.70	19.18±0.61	30.34±0.56	31.99±0.86	63.83±0.71	79.92±0.77	86.38±0.72
NBF9	179.30±0.85	7.50±0.35	0.17±0.01	17.50±0.59	23.97±0.73	30.37±0.60	57.46±0.62	78.30±0.73	84.78±0.68	95.95±0.71
NBF10	180.91±1.39	7.25±0.25	0.16±0.02	22.28±0.82	31.95±0.93	44.73±0.85	54.35±0.74	68.76±0.64	78.36±0.80	91.14±0.77
NBF11	194.91±1.15	7.75±0.23	0.24±0.02	6.36±0.37	12.76±0.66	20.76±0.63	31.94±0.62	65.42±0.65	78.34±0.65	81.60±0.76
NBF12	193.08±1.66	8.00±0.15	0.24±0.03	4.87±0.42	14.35±1.07	36.68±0.83	43.17±0.70	59.12±0.88	70.35±0.78	76.78±0.66
NBF13	174.18±1.27	7.50±0.53	0.17±0.01	30.23±1.01	38.36±0.81	43.18±0.80	54.35±0.57	76.69±0.40	84.77±0.78	88.00±0.66
NBF14	146.48±1.29	6.75±0.23	0.11±0.01	17.50±0.62	35.11±0.73	41.57±1.22	52.75±0.56	76.68±0.57	91.14±0.79	97.58±0.64
NBF15	185.10±1.30	7.50±0.23	0.22±0.02	11.14±0.66	19.16±0.72	28.75±0.70	47.90±0.72	62.33±0.48	76.73±0.80	84.77±0.56

All the data shown were measured in mean±SD where n = 3.



C: HPMC E15 (%) ^{3.75} ^{1.5} A: Carbopol 934 (%) Fig. 3: 3D plot showing the effect of carbopol 934 and HPMC E15 on the swelling index

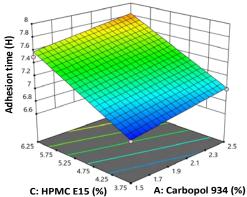


Fig. 5: 3D plot showing the effect of carbopol 934 and HPMC E15 on

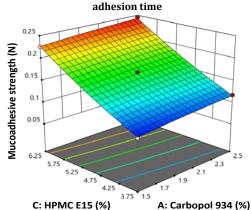
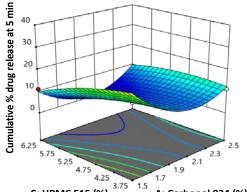
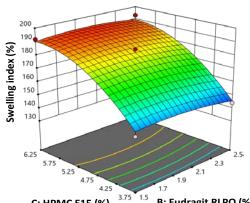


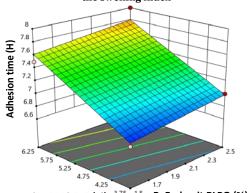
Fig. 7: 3D plot showing the effect of carbopol 934 and HPMC E15 on mucoadhesive strength



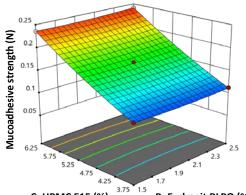
C: HPMC E15 (%) ^{3.73} ^{1.3} A: Carbopol 934 (%) Fig. 9: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 5 min



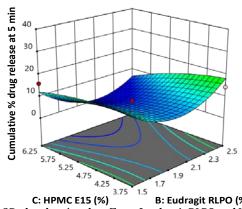
C: HPMC E15 (%) ^{3.75 1.5} B: Eudragit RLPO (%) Fig. 4: 3D plot showing the effect of eudragit RLPO and HPMC E15 on the swelling index



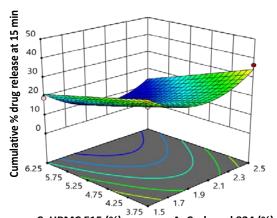
C: HPMC E15 (%) ^{3.75} ^{1.5} B: Eudragit RLPO (%) Fig. 6: 3D plot showing the effect of eudragit RLPO and HPMC E15 on adhesion time



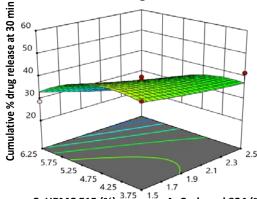
C: HPMC E15 (%) ^{3.73} B: Eudragit RLPO (%) Fig. 8: 3D plot showing the effect of eudragit RLPO and HPMC E15 on mucoadhesive strength



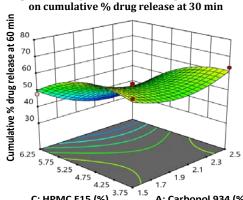
C: HPMC E15 (%) B: Eudragit RLPO (%) Fig. 10: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 5 min



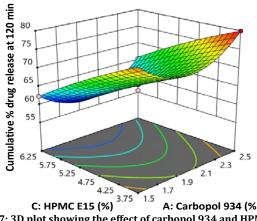
C: HPMC E15 (%) A: Carbopol 934 (%) Fig. 11: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 15 min



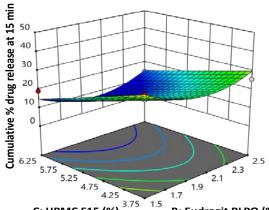
C: HPMC E15 (%) ^{3.75} ^{1.5} A: Carbopol 934 (%) Fig. 13: 3D plot showing the effect of carbopol 934 and HPMC E15



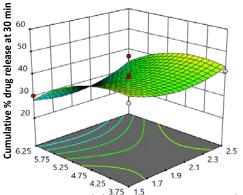
C: HPMC E15 (%) 3.75 1.5 A: Carbopol 934 (%) Fig. 15: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 60 min



C: HPMC E15 (%) ^{3.7} A: Carbopol 934 (%) Fig. 17: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 120 min



C: HPMC E15 (%) ^{3.73}^{1.3} B: Eudragit RLPO (%) Fig. 12: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 15 min



C: HPMC E15 (%) ^{3.75} ^{1.7} B: Eudragit RLPO (%) Fig. 14: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 30 min

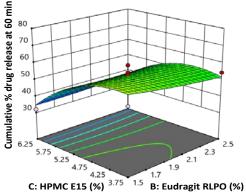
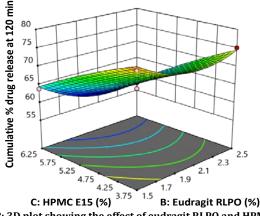
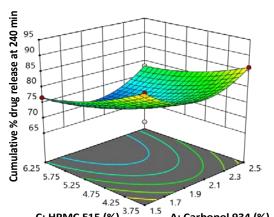


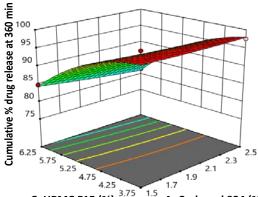
Fig. 16: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 60 min



C: HPMC E15 (%) ^{3.73} ^{1.3} B: Eudragit RLPO (%) Fig. 18: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 120 min



C: HPMC E15 (%) ^{3.75}^{1.5} A: Carbopol 934 (%) Fig. 19: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 240 min



C: HPMC E15 (%) ^{3.75}^{1.5} A: Carbopol 934 (%) Fig. 21: 3D plot showing the effect of carbopol 934 and HPMC E15 on cumulative % drug release at 360 min

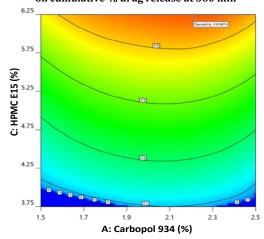
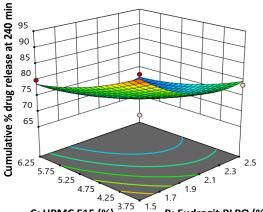
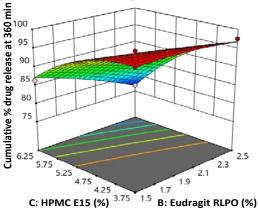


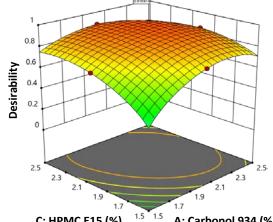
Fig. 23: Contour plot showing the maximum desirability of mucoadhesive nicotine buccal film



C: HPMC E15 (%) ^{3.75} ^{1.5} B: Eudragit RLPO (%) Fig. 20: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 240 min



C: HPMC E15 (%) ^{3.75} ^{1.5} B: Eudragit RLPO (%) Fig. 22: 3D plot showing the effect of eudragit RLPO and HPMC E15 on cumulative % drug release at 360 min



C: HPMC E15 (%) ^{1.5} ^{1.5} A: Carbopol 934 (%) Fig. 24: 3D-response surface curve showing the maximum desirability of mucoadhesive nicotine buccal film

Response variable	Model	Sequential p-value	Lack of fit <i>p</i> -value	Adjusted R ² value	Predicted R ² value
Swelling index (R1)	Quadratic	0.0124	0.2086	0.9567	0.7833
Adhesion time (R2)	Linear	< 0.0001	-	0.8963	0.8301
Mucoadhesive strength (R3)	Quadratic	0.0354	0.3972	0.9964	0.9845
Cumulative % drug release at 5 min (R4)	Quadratic	0.0243	0.0606	0.6617	-0.8651
Cumulative % drug release at 15 min (R5)	Quadratic	0.0130	0.0873	0.8283	0.0686
Cumulative % drug release at 30 min (R6)	Quadratic	0.0969	0.0882	0.7419	-0.3992
Cumulative % drug release at 60 min (R7)	Quadratic	0.0938	0.0411	0.6881	-0.7400
Cumulative % drug release at 120 min (R8)	Quadratic	0.0011	0.5979	0.9587	0.8563
Cumulative % drug release at 240 min (R9)	Quadratic	0.0416	0.9153	0.7490	0.5606
Cumulative % drug release at 360 min (R10)	Quadratic	0.0071	0.5434	0.9803	0.9267

Prediction of optimized formulation of mucoadhesive nicotine buccal film

Statistical analysis was performed using the Design Expert software. The software provided a final optimized formula for the mucoadhesive nicotine buccal film with a desirability of 0.917. This formula met all the requirements for the properties of the final batch, such as maximum swelling index, adhesion time, mucoadhesive strength, and prolonged release of the drug. The mucoadhesive nicotine buccal films were prepared using predicted values and then evaluated. The results

of this evaluation are shown in table 6. The graph of cumulative % drug release between the predicted and the observed responses showed that the predicted and observed responses were in close agreement with each other. The drug release profile of the predicted and observed batch of optimized mucoadhesive nicotine buccal film is shown in fig. 25. A linear regression plot between the predicted and observed drug release profile was also plotted for studying the correlation between them, as shown in fig. 26. The regression coefficient (R²) was calculated as 0.9982 and the correlation coefficient was 0.9989, both of which were close to 1.

Table 6: Software predicted and experimentally observed variable response data of optimized mucoadhesive nicotine buccal film

Optimized formulation composition		Response					
Component	Quantity	Evaluation parameter	Software	Experimentally	Relative error		
			predicted	observed	(%)		
A = Carbopol 934	2.21 %	Swelling index	193.95	188.21±1.93	2.96		
B = Eudragit RLPO	2.19 %	Adhesion time	7.79	7.75±0.22	0.59		
C = HPMC E 15	6.25 %	Mucoadhesive strength	0.24	0.23±0.01	2.32		
		Cumulative % drug release at 5 min	3.20	3.34±0.10	4.40		
		Cumulative % drug release at 15 min	10.00	9.83±0.75	1.67		
		Cumulative % drug release at 30 min	26.96	28.57±0.90	5.97		
		Cumulative % drug release at 60 min	34.11	36.14±0.97	5.94		
		Cumulative % drug release at 120 min	59.12	54.10±1.09	8.49		
		Cumulative % drug release at 240 min	72.25	70.11±0.99	2.96		
		Cumulative % drug release at 360 min	79.72	76.55±0.99	3.97		

All the data shown were measured in mean±SD where n = 3

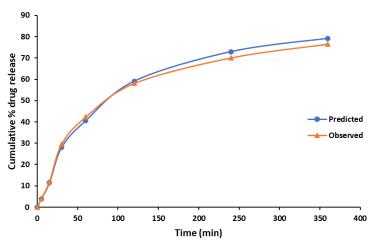


Fig. 25: Predicted and observed drug release profile of optimized mucoadhesive nicotine buccal film. All the data shown were measured in mean±SD where n = 3

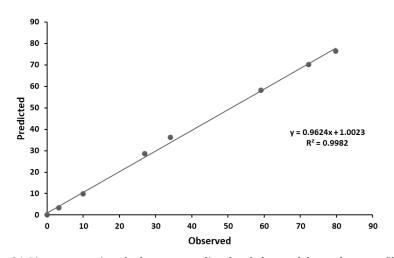


Fig. 26: Linear regression plot between predicted and observed drug release profile

Surface pH

The surface pH of the film was measured to determine the effect of pH on the buccal mucosa. It was found that the pH of the optimized buccal film was 7.54±0.10, which is neutral and suitable for the buccal mucosa, as it does not irritate. This indicates that the pH of the film is well-suited for the buccal mucosa and does not cause any discomfort [30].

Film weight and thickness

A mucoadhesive buccal film with consistent thickness and weight ensures a uniform drug concentration throughout the film. The optimized mucoadhesive nicotine buccal film was found to have a thickness of 0.18 ± 0.01 mm, which is an optimal thickness for a buccal film because a film that is too thick may not dissolve as intended, resulting in slow drug release, while films that are too thin may dissolve too quickly and may not deliver an adequate amount of drug to be effective [25]. The weight of the optimized mucoadhesive nicotine buccal film (2 cm x 2 cm) was found to be 106.28 ± 0.37 mg, which is uniform for buccal mucosa application and homogeneity [31].

Folding endurance

High folding endurance is a desirable characteristic in films as it ensures that they do not easily displace from the application site or break while being administered. The optimized mucoadhesive nicotine buccal film was found to have a folding endurance of 540folds, indicating that it has high mechanical strength and resistance to breaking [32].

Tensile strength and extensibility

Soft and durable inserts that can withstand the continuous mechanical stress caused by jaw movement in the buccal cavity are preferred for buccal application. The tensile strength of the optimized mucoadhesive nicotine buccal film was observed to be 87.20 N and its extensibility was observed to be 23.69 mm as seen in fig. 27. The formulation exhibited high tensile strength and optimal elongation due to the high concentration of HPMC E15 and the

presence of carbopol 934, respectively [33]. As reported in the literature for oral thin films of nicotine [34], the mucoadhesive buccal film showed higher mechanical strength, as indicated by its higher tensile strength and greater flexibility and elongation without breaking, as shown by its higher extensibility. These properties suggest that the film has better integrity and mechanical properties, making it a more durable and robust option for buccal drug delivery.

Swelling index

The swelling index is an important factor in drug delivery through mucoadhesive buccal film due to its ability to affect the rate of drug release. This is because water absorption-induced swelling allows initially twisted, stretched, or entangled bioadhesive polymers to relax, resulting in the rapid disentanglement of individual polymer chains and the formation of a specific-size macromolecular network that increases the porosity of the film and initiates drug release [35]. The swelling index of all the batches was found to be in the range of 142.22 % to 194.91 %, as recorded in table 4. The 3D response surface plots in fig. 3 and fig. 4 show the effect of independent variables on the swelling index of all batches. It was observed that the swelling index increased with an increase in the concentration of HPMC E15 and carbopol 934, as they are hydrophilic polymers that absorb water from the surrounding environment, increasing the size and volume of the film, but there was no significant effect of eudragit RLPO on the swelling index of the film. The swelling index of the optimized mucoadhesive nicotine buccal film was found to be 188.21±1.93 %, indicating a slow drug release profile, as a high swelling index is generally associated with a slower drug release, while a low swelling index is associated with a more rapid release. The results indicate that the developed mucoadhesive nicotine buccal film has a higher swelling index than nicotine thin films as reported in the literature [36]. The higher swelling index leads to a slower drug release profile and stronger mucoadhesion, suggesting that the mucoadhesive nicotine buccal film has better water uptake and film integrity. In contrast, nicotine thin films tend to disintegrate quickly and have a faster release rate.

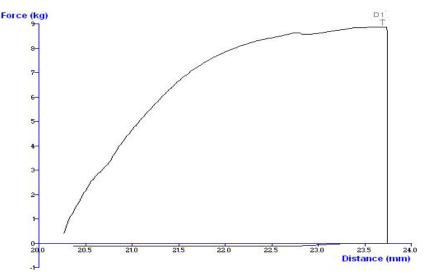


Fig. 27: Tensile strength and extensibility of optimized mucoadhesive nicotine buccal film

Adhesion time

Adhesion time is a measure of the effectiveness of mucoadhesive properties of film and can impact the rate and duration of drug delivery. A longer adhesion time allows for a slower release of the drug, while a shorter adhesion time results in a rapid release [37]. The adhesion time of all the batches was found to be in the range of 6 h to 8 h, as recorded in table 4. The 3D response surface plots in fig. 5 and fig. 6 show the effect of independent variables on the adhesion time of all batches. The results showed that the adhesion time increased with an increase in the concentration of HPMC E15 and carbopol 934 because HPMC E15 formed a gel that adhered for a longer period, whereas carbopol 934 increased the surface charge of the film, allowing it to adhere better. However, the varied concentration of eudragit RLPO had no substantial effect. The adhesion time of the optimized mucoadhesive nicotine buccal film was found to be 8 h approximately, indicating that the mucoadhesive film sustained a strong bond for an extended period, leading to a prolonged release of the drug. The developed mucoadhesive nicotine buccal film exhibits a higher adhesion time compared to 2-3 h adhesion time of nicotine mucoadhesive tablets and other mucoadhesive buccal films as reported in various literatures [16, 38]. This improved adhesion leads to enhanced efficacy, patient compliance, and convenience of nicotine replacement therapy by reducing the frequency of drug administration.

Mucoadhesive strength

Mucoadhesive strength is a measure of the adhesive force between a mucoadhesive buccal film and a mucous membrane. The mucoadhesive strength of all the batches was found to be in the range of 0.09 N to 0.24 N, as recorded in table 4. The 3D response surface plots in fig. 7 and fig. 8 show the effect of independent variables on the mucoadhesive strength of all batches. It was observed that the mucoadhesive strength increased with an increase in the concentration of HPMC E15 and carbopol 934. This is because HPMC E15 increases the mechanical strength and flexibility of the film, while carbopol 934 increases the surface charge on the film, thus allowing better interaction with the negatively charged mucosal tissue and creating a stronger bond [39]. However, the concentration of eudragit RLPO did not have a significant impact on the mucoadhesive strength. The mucoadhesive strength of the optimized mucoadhesive nicotine buccal film was found to be 0.23±0.01 N, which is higher than nicotine wafer and film formulations as reported in literature [40]. This allows it to maintain high drug concentration and prevent displacement, resulting in more effective nicotine delivery, as well as enhancing patient compliance and convenience by reducing the frequency of drug administration.

Drug content uniformity

The uniformity of drug content is an important aspect of pharmaceutical quality control to ensure that the drug is distributed evenly throughout the formulation. The drug content of the optimized mucoadhesive nicotine buccal film was found to be 96.13 ± 1.71 %. The result indicates that the nicotine was dispersed

uniformly throughout the buccal film, with a very low and acceptable standard deviation [35].

In vitro drug release studies

The in vitro drug release studies of formulated batches were carried out in dissolution apparatus using phosphate buffer pH 6.8 as the dissolution medium. The cumulative % drug release of all batches ranged from 4.77 % to 30.23 % at 5 min and 76.78 % to 97.58 % at 360 min, as recorded in table 4 and shown graphically in fig. 2. All formulated batches showed an initial burst release of nicotine, which was followed by a prolonged release. The initial burst effect varied depending on the viscosity of the polymers. The 3D response surface plots from fig. 9 to fig. 22 show the effect of the independent variables on the cumulative % drug release for different time intervals for all batches. It was observed that the drug release decreased with an increase in the concentration of HPMC E15, this is because the matrix formed by the polymer entrapped the drug and prolonged its release [25]. However, the drug release slightly decreased with an increase in the concentration of carbopol 934 and eudragit RLPO. It is because carbopol 934 absorbs water and swells, which affects the release of the drug from the film, and eudragit RLPO alters the release of nicotine as it is a pH-sensitive polymer. The in vitro drug release study of the optimized batch of mucoadhesive nicotine buccal film showed a cumulative % drug release of 76.55±0.99 % at 360 min as shown in fig. 28. This infers that it has an initial burst release followed by a prolonged release. The in vitro drug release studies of nicotine lozenges and transdermal patches have been reported in various literature [41, 42]. However, the developed mucoadhesive nicotine buccal film exhibited both rapid onset of action and prolonged release, setting it apart from the lozenge and transdermal patch formulations which provide only either fast or prolonged action.

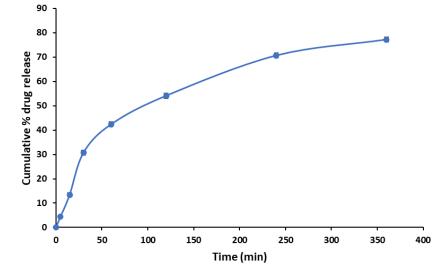


Fig. 28: *In vitro* drug release profile of optimized mucoadhesive nicotine buccal film. All the data shown were measured in mean±SD where n = 3

Ex-vivo adhesion time

The *ex-vivo* adhesion time of the optimized mucoadhesive nicotine buccal film was measured to be 6 h approximately, which was found to be in concurrence with the previously reported studies on mucoadhesive buccal films [29]. This is significant as prolonged adhesion time is desirable in nicotine replacement therapy, allowing for consistent delivery of nicotine over an extended period which can increase its effectiveness in aiding smoking cessation. This is due to the properties of HPMC E15, which is a hydrophilic polymer that swells significantly upon absorbing water, and is less affected by changes in hydration, thereby maintaining its structure well and prolonging the release of the entrapped drug. The increased viscosity of HPMC also resulted in the formation of a surface gel that

remained in place for a long time, contributing to the extended adhesion time of the film. Further, higher carbopol content increases the surface charge on the film, allowing it to interact better with the negatively charged mucosal tissue, creating a stronger bond [27, 43].

Ex-vivo permeation

Ex-vivo permeation studies were conducted to determine the kinetics of drug absorption through a biological membrane. The result indicates that the optimized mucoadhesive nicotine buccal film had a 2-fold higher drug permeation rate (0.56 mg or 37.38 %) compared to the lozenge (0.28 mg or 19.29 %) over 360 min. The optimized film also permeated a higher percentage of the drug (95.09 %) compared to the lozenge (49.07 %) over the same

period. The swelling properties of HPMC E15, the surface charge of carbopol, and the pH-sensitive nature of eudragit RLPO contributed to the higher drug permeation of the optimized film, as shown in fig. 29. The *ex-vivo* permeation study of marketed

lozenge is reported in various literature [44]. However, the use of mucoadhesive nicotine buccal film has an advantage over the marketed lozenge in terms of higher drug delivery efficiency through the biological membrane.

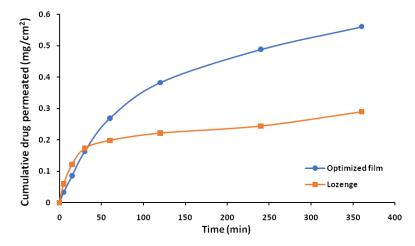


Fig. 29: Comparison of cumulative drug permeated from optimized mucoadhesive nicotine buccal film and lozenge. All the data shown were measured in mean±SD where n = 3

Kinetic release assessment

The *in vitro* drug release data for the optimized mucoadhesive nicotine buccal film were analyzed using different mathematical models such as zero order, first order, Higuchi, Korsmeyer-peppas, and Hixson-crowell. The best fit in the drug release kinetic model was selected based on the highest regression coefficient (R^2) value. It was found that *in vitro* drug release kinetic followed the Higuchi model with the highest regression coefficient (R^2) of 0.9679 as recorded in table 7 and plotted in fig. 30. This implies that the drug

release mechanism from the film follows a diffusion-controlled process, where the rate of drug release is directly proportional to the surface area available for dissolution, providing a rapid onset of drug release followed by a prolonged release [45]. The results were found in agreement with previously reported work done on nicotine fast-dissolving film and nicotine mucoadhesive tablets which respectively provided either a fast onset of action or prolonged release [38, 46]. However, the developed mucoadhesive nicotine buccal film combines the benefits of both fast onset and prolonged action, making it a better option in terms of drug release kinetics.

Table 7: Drug release kinetic data and model fitting of optimized mucoadhesive nicotine buccal film

S. No.	Drug release kinetic model	Equation	К	\mathbb{R}^2
1.	Zero-order	$Qo-Q_t = k_0 t$	2.09 x 10 ⁻¹ mol/min	0.8452
2.	First order	$\log Q = \log Q_0 - kt/2.303$	-1.80 x 10 ⁻³ mol/min	0.9494
3.	Higuchi	$Q_0 = Q_t = kt^{1/2}$	4.43 x 10 ° mol/min	0.9679
4.	Korsmeyer-Peppas	$\log (Q_0 - Q_t) = \log k - n \log t$	7.72 x 10 ⁻¹ mol/min	0.9640
5.	Hixson-Crowell	$Q_0^{1/3} - Q_t^{1/3} = kt$	5.0 x 10 ⁻³ mol/min	0.9196

 Q_0 is initial drug concentration; Q_t is the amount of drug remaining at a specific time; k is rate constant; t is time.

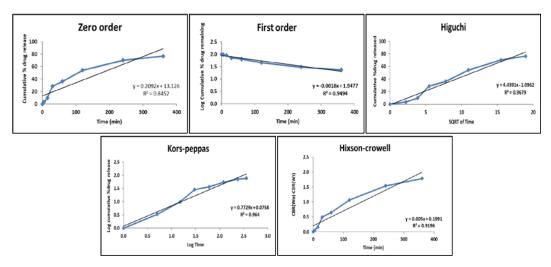


Fig. 30: Kinetic graphs of in vitro drug release of optimized mucoadhesive nicotine buccal film

CONCLUSION

In the present study, the mucoadhesive nicotine buccal film was formulated by solvent casting method and optimized by boxbehnken experimental design. The film was evaluated for its swelling index, adhesion time, mucoadhesive strength, and cumulative % drug release. The effect of independent variables on response variables was determined using 3D surface plots and polynomial equations. The optimized formulation was prepared based on the evaluation data and desired constraints of response variables, resulting in the highest desirability. The optimized film demonstrated a prolonged adhesion time and better contact with the buccal mucosa, resulting in a rapid onset of action followed by prolonged release. The in vitro drug release study showed a release of 76.55 % in 6 h. The ex-vivo study of developed mucoadhesive nicotine buccal film revealed a 2-fold enhancement in buccal mucosal permeation compared to a lozenge. This inferred that the optimized mucoadhesive nicotine buccal film can be an effective tool for smokers seeking to quit, increasing the likelihood of achieving and maintaining a smoke-free lifestyle.

ACKNOWLEDGEMENT

The authors acknowledge Gelnova Laboratories Pvt. Ltd. (Navi Mumbai), for the drug sample and Lubrizol, Evonik, and Colorcon for polymers.

AUTHORS CONTRIBUTIONS

All the authors contributed equally.

CONFLICT OF INTERESTS

The authors hereby declare that there is no conflict of interest and other support.

REFERENCES

- 1. West R. Tobacco smoking: health impact, prevalence, correlates and interventions. Psychol Health. 2017;32(8):1018-36. doi: 10.1080/08870446.2017.1325890, PMID 28553727.
- Benowitz NL. Nicotine addiction. N Engl J Med. 2010;362(24):2295-303. doi: 10.1056/NEJMra0809890.
- Hartmann Boyce J, Chepkin SC, Ye W, Bullen C, Lancaster T. Nicotine replacement therapy versus control for smoking cessation. Cochrane Database Syst Rev. 2018;5(5):CD000146. doi: 10.1002/14651858.CD000146.pub5. PMID 29852054.
- Thompson GH, Hunter DA. Nicotine replacement therapy. Ann Pharmacother. 1998;32(10):1067-75. doi: 10.1345/aph.17382, PMID 9793600.
- Kamala K, Sankethguddad S, Sujith S. An update on nicotine replacement therapy. J Oral Res Rev. 2019;11(2):41-7. doi: 10.4103/jorr.jorr_9_18.
- Sachdev R, Garg K. Nicotine replacement therapy: effective tool in smoking cessation: a review. Glob J Res Anal. 2018;7:456-7.
- Wadgave U, Nagesh L. Nicotine replacement therapy: an overview. Int J Health Sci (Qassim). 2016;10(3):425-35. doi: 10.12816/0048737, PMID 27610066.
- Herman AI, Sofuoglu M. Comparison of available treatments for tobacco addiction. Curr Psychiatry Rep. 2010;12(5):433-40. doi: 10.1007/s11920-010-0134-6, PMID 20623259.
- Abd El Azim H, Nafee N, Ramadan A, Khalafallah N. Liposomal buccal mucoadhesive film for improved delivery and permeation of water-soluble vitamins. Int J Pharm. 2015;488(1-2):78-85. doi: 10.1016/j.ijpharm.2015.04.052, PMID 25899288.
- Adrover A, Di Muzio L, Trilli J, Brandelli C, Paolicelli P, Petralito S. Enhanced loading efficiency and mucoadhesion properties of gellan gum thin films by complexation with hydroxypropyl-βcyclodextrin. Pharmaceutics. 2020;12(9):3-21. doi: 10.3390/pharmaceutics12090819, PMID 32872207.
- Fonseca Santos B, Chorilli M. An overview of polymeric dosage forms in buccal drug delivery: state of art, design of formulations and their *in vivo* performance evaluation. Mater Sci Eng C Mater Biol Appl. 2018;86:129-43. doi: 10.1016/j.msec.2017.12.022, PMID 29525088.
- 12. Kontogiannidou E, Ferrari M, Deligianni AD, Bouropoulos N, Andreadis DA, Sorrenti M. *In vitro* and ex vivo evaluation of tablets containing piroxicam-cyclodextrin complexes for buccal

delivery. Pharmaceutics. 2019;11(8). doi: 10.3390/pharmaceutics11080398, PMID 31398833.

- Salehi S, Boddohi S. Design and optimization of kollicoat
 IR based mucoadhesive buccal film for co-delivery of rizatriptan benzoate and propranolol hydrochloride. Mater Sci Eng C Mater Biol Appl. 2019;97:230-44. doi: 10.1016/j.msec.2018.12.036, PMID 30678908.
- Abruzzo A, Bigucci F, Cerchiara T, Cruciani F, Vitali B, Luppi B. Mucoadhesive chitosan/gelatin films for buccal delivery of propranolol hydrochloride. Carbohydr Polym. 2012;87(1):581-8. doi: 10.1016/j.carbpol.2011.08.024, PMID 34663007.
- Hearnden V, Sankar V, Hull K, Juras DV, Greenberg M, Kerr AR. New developments and opportunities in oral mucosal drug delivery for local and systemic disease. Adv Drug Deliv Rev. 2012;64(1):16-28. doi: 10.1016/j.addr.2011.02.008, PMID 21371513.
- Abdella S, Afinjuomo F, Song Y, Upton R, Garg S. Mucoadhesive buccal film of estradiol for hormonal replacement therapy: development and *in vivo* performance prediction. Pharmaceutics. 2022;14(3):1-24. doi: 10.3390/pharmaceutics14030542, PMID 35335917.
- 17. Sindhu GD, Nallaguntla L, Kulkarni GS. Buccal mucoadhesive films. Int J Curr Pharm. 2021;13(4):14-20.
- Tejada G, Piccirilli GN, Sortino M, Salomón CJ, Lamas MC, Leonardi D. Formulation and *in vitro* efficacy of antifungal mucoadhesive polymeric matrices for the delivery of miconazole nitrate. Mater Sci Eng C Mater Biol Appl. 2017;79:140-50. doi: 10.1016/j.msec.2017.05.034, PMID 28629000.
- Abruzzo A, Nicoletta FP, Dalena F, Cerchiara T, Luppi B, Bigucci F. Bilayered buccal films as child-appropriate dosage form for systemic administration of propranolol. Int J Pharm. 2017;531(1):257-65. doi: 10.1016/j.ijpharm.2017.08.070, PMID 28811117.
- Krampe R, Visser JC, Frijlink HW, Breitkreutz J, Woerdenbag HJ, Preis M. Oromucosal film preparations: points to consider for patient centricity and manufacturing processes. Expert Opin Drug Deliv. 2016;13(4):493-506. doi: 10.1517/17425247.2016.1118048, PMID 26559519.
- Ferreira SLC, Bruns RE, Ferreira HS, Matos GD, David JM, Brandão GC. Box-behnken design: an alternative for the optimization of analytical methods. Anal Chim Acta. 2007;597(2):179-86. doi: 10.1016/j.aca.2007.07.011, PMID 17683728.
- Haju SS, Yadav S. Formulation and evaluation of cilnidipine mucoadhesive buccal film by solvent casting technique for the treatment of hypertension. Int J Pharm Pharm Sci. 2021;13(9):34-43. doi: 10.22159/ijpps.2021v13i9.42641.
- Abouhussein DMN, El El-bary AA, Shalaby SH, El Nabarawi MAE. Chitosan mucoadhesive buccal films: effect of different casting solvents on their physicochemical properties. Int J Pharm Pharm Sci. 2016;8(9):206-13. doi: 10.22159/ijpps.2016.v8i9.12999.
- 24. Haju S, Yadav S, Baig R, Sawant G. Buccal film: a novel approach for oral mucosal drug delivery system. Asian J Pharm Clin Res. 2021;14(1):27-35. doi: 10.22159/ajpcr.2021.v14i1.39687.
- Karki S, Kim H, Na S, Shin D, Jo K, Lee J. Thin films as an emerging platform for drug delivery. Asian Journal of Pharmaceutical Sciences. 2016;11(5):559-74. doi: 10.1016/j.ajps.2016.05.004.
- 26. Yasmin Begum M, Alqahtani A, Ghazwani M. Preparation of carbopol 934 based ketorolac tromethamine buccal mucoadhesive film: *in vitro*, ex vivo, and *in vivo* assessments. Int J Polym Sci. 2021;2021:1-11.
- 27. Roy Biswas G. Hydroxypropyl methylcellulose: different aspects in drug delivery. J Pharm Pharmacol. 2016;4:381-5.
- Patra CN, Priya R, Swain S, Kumar Jena G, Panigrahi KC, Ghose D. Pharmaceutical significance of eudragit: a review. Future J Pharm Sci. 2017;3(1):33-45. doi: 10.1016/j.fjps.2017.02.001.
- Ikram M, Gilhotra N, Gilhotra RM. Formulation and optimization of mucoadhesive buccal patches of losartan potassium by using response surface methodology. Adv Biomed Res. 2015;4:239. doi: 10.4103/2277-9175.168606, PMID 26682205.

- Aframian DJ, Davidowitz T, Benoliel R. The distribution of oral mucosal pH values in healthy saliva secretors. Oral Dis. 2006;12(4):420-3. doi: 10.1111/j.1601-0825.2005.01217.x, PMID 16792729.
- Dixit GR, Chavhan JI, Upadhye PK, Misra S. Formulation and characterization of mucoadhesive buccal film of ranitidine hydrochloride using sterculla foetida gum as polymer. Asian J Pharm Clin Res. 2015;8(3):68-71.
- Semalty A, Semalty M, Nautiyal U. Formulation and evaluation of mucoadhesive buccal films of enalapril maleate. Indian J Pharm Sci. 2010;72(5):571-5. doi: 10.4103/0250-474X.78522, PMID 21694987.
- Bodini RB, Guimaraes J, Monaco Lourenço CA, Aparecida de Carvalho R. Effect of starch and hydroxypropyl methylcellulose polymers on the properties of orally disintegrating films. J Drug Deliv Sci Technol. 2019;51:403-10. doi: 10.1016/j.jddst.2019.03.028.
- 34. Khan MA, Ameen S, Zia A, Ijaz U. Formulation of fast dissolving oral films of nicotine. Contemp Pharm Pract. 2018;2(1):1-7.
- Kumar A, Bali V, Kumar M, Pathak K. Comparative evaluation of porous versus nonporous mucoadhesive films as buccal delivery system of glibenclamide. AAPS PharmSciTech. 2013;14(4):1321-32. doi: 10.1208/s12249-013-0014-6, PMID 23990119.
- Konda M, Chinnala KM, Samineni L, Naznin Y, Avirneni J, Kondamalla KC. Formulation and characterization of nicotine thin films for smoking cessation through buccal delivery. Intern Jour Contemp Microbiol 2016;9(12):2079. doi: 10.5958/0974-360X.2016.00424.8.
- Shaikh R, Raj Singh TR, Garland MJ, Woolfson AD, Donnelly RF. Mucoadhesive drug delivery systems. J Pharm Bioallied Sci. 2011;3(1):89-100. doi: 10.4103/0975-7406.76478, PMID 21430958.
- 38. Bahri Najafi R, Rezaei Z, Peykanpour M, Shabab L, Solooki R, Akbari P. Formulation of nicotine mucoadhesive tablet for

smoking cessation and evaluation of its pharmaceuticals properties. Adv Biomed Res. 2013;2:88. doi: 10.4103/2277-9175.122515, PMID 24524034.

- Jacob S, Nair AB, Boddu SHS, Gorain B, Sreeharsha N, Shah J. An updated overview of the emerging role of patch and film-based buccal delivery systems. Pharmaceutics. 2021;13(8):1-39. doi: 10.3390/pharmaceutics13081206, PMID 34452167.
- Boateng J, Okeke O. Evaluation of clay-functionalized wafers and films for nicotine replacement therapy via buccal mucosa. Pharmaceutics. 2019;11(3):104. doi: 10.3390/pharmaceutics11030104, PMID 30832244.
- 41. Pothu R, Shayeda YM. Development and *in vitro* evaluation of nicotine troches for smoking cessation. Asian J Pharm Clin Res. 2014;7(2):68-75.
- 42. Maheswara Reddy D, Kumar V, Harshini P, Lahari P. Formulation and evaluation of nicotine transdermal patches by the using of hydrophilic and hydrophobic polymers. World J Pharm Res. 2018;7(16):1101-15.
- Surini S, Gotalia F, Putri KSS. Formulation of mucoadhesive buccal films using pregelatinized cassava starch phthalate as a film-forming polymer. Int J App Pharm. 2018;10(1):225-9. doi: 10.22159/ijap.2018.v10s1.50.
- Ngono F, Willart JF, Cuello GJ, Jimenez Ruiz M, Yelles CH, Affouard F. Impact of amorphization methods on the physicochemical properties of amorphous lactulose. Mol Pharm. 2020;17(1):1-9. doi: 10.1021/acs.molpharmaceut.9b00740. PMID 31647674.
- Higuchi T. Mechanism of sustained-action medication. Theoretical analysis of rate of release of solid drugs dispersed in solid matrices. J Pharm Sci. 1963;52(12):1145-9. doi: 10.1002/jps.2600521210, PMID 14088963.
- 46. Kheawfu K, Panraksa P, Jantrawut P. Formulation and characterization of nicotine microemulsion-loaded fastdissolving films for smoking cessation. Molecules. 2022;27(10). doi: 10.3390/molecules27103166, PMID 35630643.