

ANTIBACTERIAL ACTIVITY OF SYNTHESIZED SILVER NANOPARTICLES BY *SIMAROUBAGLAUCA* AGAINST PATHOGENIC BACTERIA

C. P. CHANDRAPPA^{1*}, N. CHANDRASEKAR², M. GOVINDAPPA², CHAITRA SHANBHAG¹, UTTAM KUMAR SINGH¹,
JAYASHRI MASARGHAL¹

¹Department of Biotechnology, Shridevi Institute of Engineering and Technology, Sira Road, Tumkur 572106, Karnataka, India,

²Department of Chemistry, Shridevi Institute of Engineering and Technology, Sira Road, Tumkur 572106, Karnataka, India,

Email: chandrappacp@gmail.com

Received: 27 Jan 2017, Revised and Accepted: 20 Apr 2017

ABSTRACT

Objective: The present study outline the plant-mediated synthesis of silver nanoparticles (AgNPs) using leaf extract *Simaroubaglauca*, which act as both reducing and stabilizing agent.

Methods: Formation of silver nanoparticles was confirmed by primarily by Ultraviolet/visible spectroscopy. X-ray diffraction studies revealed the crystallinity of the nanoparticles. The scanning electron microscopy was carried out to determine the mean particle size, as well as the morphology of the NPs and the composition of elements, was studied with Energy Dispersive X-ray analysis (EDS).

Results: The silver nanoparticles were spherical in shape with a mean size of 23 nm. The EDS showed strong optical absorption peak at 3keV and it was confirmed the formation of AgNPs. The synthesised AgNPs further utilized for the evaluation of antibacterial activity and shown significant antibacterial activity against *Escherichia coli*, *Pseudomonas aeruginosa*, *Enterobacter* and *Klebsiella pneumonia* at 50 µg/ml and 100µg/ml concentrations.

Conclusion: The synthesised silver nanoparticles have been characterised by UV-vis, SEM-EDAX and XRD to determine the sizes and shapes of the silver nanoparticles.

Keywords: *Simaroubaglauca*, Silver nanoparticles, Leaf extract, Bioreductant, Antibacterial

© 2017 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)
DOI: <http://dx.doi.org/10.22159/ijcpr.2017v9i4.20629>

INTRODUCTION

Every person will suffer from either one or other diseases caused by various bacteria at least once in his or her lifetime that shows resistance against one or two existed antibiotics and which leads a severe public health problem [1, 2].

Hence, there is top-priority to develop alternative treatments for bacterial diseases. Silver was being used in the field of medicine for antimicrobial applications such as burn therapy [3, 4], removal of microbes on textile fabrics [5-7], and inhibition of colonization of bacteria on catheters [8-10]. As antimicrobial agents, Nano-silver systems offer many advantages. They own a very high effect towards a broad range of microbes and parasites, even at low doses and does not shows toxicity in humans and relatively inexpensive.

Thus Silver has been suspended within a wide variety of materials, under various forms such as salts, immobilized ions or metallic nanoparticles [11-13]. Plasma membrane, many important enzymes and DNA of the bacteria are important targets for silver ions [14-18].

Simaroubaglauca is one of the important traditional medicinal plants due to the presence alkaloids, flavonoids, carbohydrates, glycosides, a phenolic compound, tannins, terpenoids, cardenolides, saponins, fixed oils which can usually account for their therapeutic action including Antibacterial, antiviral, anti-inflammatory, antiprotozoal and antitumor activities [19]. But never synthesized and characterized silver nanoparticles by the extracts of *Simaroubaglauca*.

Here, we synthesized silver nanoparticles using an aqueous leaf extract obtained from *Simaroubaglauca*. In addition, to consider the biological application of our work, antibacterial evaluation was carried out.

MATERIALS AND METHODS

Preparation of leaf extract

Silver nitrate (AgNO₃) was purchased from sigma-aldrich and fresh *Simaroubaglauca* leaves were collected from Shridevi Institute of Engineering and Technology Campus in Tumakuru of Karnataka, India. The collected leaves were washed thoroughly and cut into small pieces. Finely incised *SIMAROUBAGLAUCA* leaves (20g) were weighed and transferred to 500 ml conical flask containing 100 ml of distilled water and gently mixed and boiled for 5 min.

The obtained extract was collected filtered through Whatman No.1 filter paper and the filtrate was collected in 250 ml Erlenmeyer flask and stored at 4 °C for further use.



Fig. 1: *Simaroubaglauca*

Synthesis of silver nanoparticles using leaf extract

Silver nanoparticles were synthesized by mixing 5 ml of *Simarouba glauca* leaf extract with 50 ml of silver nitrate aqueous solution (1 mmol) and stirred for 10 min at 30 °C. Reduction rapidly occurs as indicated by a pale bluish colour after 30 min indicating the formation of silver nanoparticles. The obtained silver nanoparticles were purified by centrifuging in a Remi cooling centrifuge at 10,000rpm for 30 min. The pellets obtained were dispersed in deionized three times to remove water solubles.

Characterization of synthesized silver nanoparticles

UV-Vis spectral analysis

Biological mediated reductions of silver salts were monitored by UV-Vis spectroscopy. Absorption spectra were obtained using Shimadzu company model-UV3600 UV-Vis spectrophotometer using quartz cuvette and distilled water as a reference. Spectral readings of UV-Vis were recorded between 350 and 550 nm for synthesized silver nanoparticles.

Scanning electron microscopy (SEM) and EDAX

Size, shape and morphology of silver nanoparticles were resolved by scanning electron microscopy by using ZEISS Ultra 55 SEM machine operated 5 kV at Center for Nanoscience and Engineering, Indian Institute of Science, Bengaluru. A thin film of the sample was prepared on a carbon coated copper grid by just keeping the very small amount of the sample on the grid. The readings and photographic scan were taken at 50.00, 75.00 and 100.00 magnification with constant voltage and at different angles.

X-ray diffraction measurements (XRD)

Crystalline metallic properties of AgNPs were examined by XRD (Rigaku, SmartLab X-ray Diffractometer) at Center for Nanoscience and Engineering, Indian Institute of Science, Bengaluru at a voltage of 40 keV and a current of 30 mA with Cu K α radiation, step size-0.02, speed-5 °/min with a wavelength of 1.5418 Å and at 2 θ angle.

Antimicrobial activity of silver nanoparticles

Microbial cultures: *Escherichia coli*, *Pseudomonas aeruginosa*, *Enterobacter* and *Klebsiella pneumonia* were collected from Department of Microbiology of Shridevi Institute of Medical Sciences and Research Hospital, Tumakuru, Karnataka of India and were maintained in nutrient broth.

The antimicrobial assay was carried out by Agar well diffusion method. Overnight samples were swabbed on the plates containing Muller Hinton Agar (MHA) medium (Hi-Media). Wells were prepared in the medium using sterile gel puncture. 50 μ g/ml and 100 μ g/ml of synthesized silver nanoparticle solution was added to wells. Sterile distilled water and antibiotic ampicillin were added as negative and positive control respectively. Petri plates were kept for incubation for 24h at 37 °C.

Statistical analysis

All the experiments were performed in triplicate, and the data were expressed as mean.

RESULTS AND DISCUSSION

The extracellular green synthesis of silver nanoparticles occurred during the exposure of the leaf extract to 1 mmol aqueous silver nitrate solution. The complete reduction of silver ions was observed 2-3h. The color change of the reaction was observed during the incubation period because the formation of silver nanoparticles is able to produce a specific color of the reaction mixtures because of their unique properties. The appearance of bluish color is a conclusive indication of the formation of silver nanoparticles in the reaction mixture (fig. 2). Due to the presence of coherent excitation of all the 'free' electrons within the conduction band, the color could be exhibited by the metal particles and leading to an in-phase oscillation which is known as surface plasmon resonance (SPR) [20].

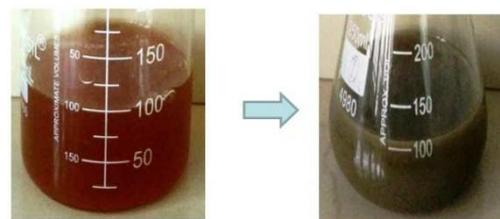


Fig. 2: *Simarouba glauca* leaf extract, before and after treating with silver nitrate

UV-vis spectroscopy technique was applied to monitor the SPR absorbance band of silver nanoparticles synthesized using leaf extract of *Simarouba glauca* and it was centered at 440 nm and relentlessly increments in intensity as a function of the time of reaction without any shift in the peak wavelength (fig. 3) [21]. The frequency and width of the surface plasmon absorption relies on the size and shape of the metal nanoparticles and in addition on the dielectric constant of the metal itself and the encompassing medium.

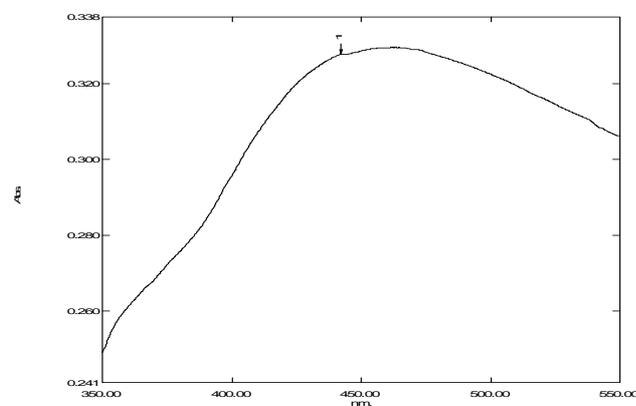


Fig. 3: UV-vis spectrum of silver nanoparticles synthesized using leaf extract *Simarouba glauca*

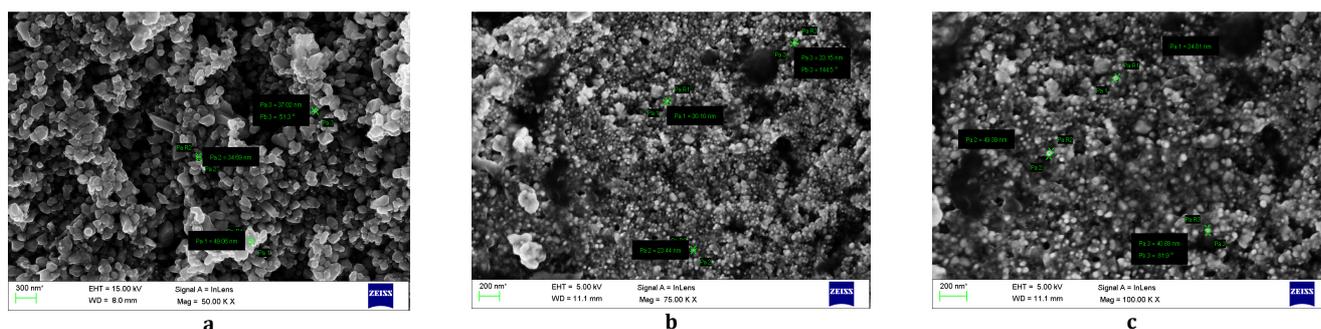


Fig. 4: SEM photographs of silver nanoparticles obtained using leaf extract *Simarouba glauca*, a. 50.00, b. 75.00, c. 100.00

The microstructural qualities of the synthesized silver nanoparticles were studied by SEM (fig. 4). Little round shaped outgrowths of silver nanoparticles agglomerates with homogenous dissemination. The silver nanoparticles were generally circular, and the size was evaluated to be in the range in the vicinity of 23 and 49 nm. The EDX investigation of silver nanoparticles affirms that the nanoparticles are in fact silver nanoparticles (fig. 5). It demonstrates that silver nanoparticles were upheld on the natural grid from the *Simarouba glauca*. The peak of Ag component is most likely from the silver nitrate.

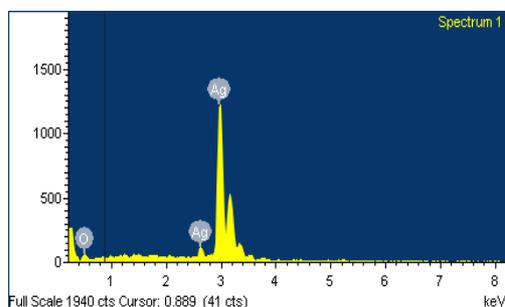


Fig. 5: EDAX spectrum of silver nanoparticles

Powder X-ray diffraction studies of silver nanoparticles

X-ray diffraction studies have provided an excellent result that confirms the biosynthesis of silver nanoparticles (AgNPs) from leaf extract of *Simarouba glauca*. The bioreductive process has generated

the cubic crystalline structure of silver nanoparticles. The XRD pattern shows distinct peaks of diffraction at 2θ . A number of Bragg reflections with 2θ values of 27.60° , 32.10° , 38.00° , 44.17° , 46.08° (222), 64.42° (400), 77.32° (420) and 81.40° , corresponded to the (111), (200), (220), (311), (222), (400), (420) and (422) face centered cubic structures of silver and compared with the data of JCPDS (Joint Committee on Powder Diffraction Standards), File No. 85-1355. The average particle sizes were calculated according to Debye-Scherrer equation [22]. According to the equation, average silver nanoparticle sizes of *Simarouba glauca* were found to be 2.310-19.929 nm range at different θ values (fig. 6). Earlier, the similar results have been reported for silver nanoparticles [23].

Antibacterial activities of Au and Ag nanoparticles

In our investigation, the synthesized silver nanoparticles have been evaluated for their antibacterial activities against human pathogens such as Gram-negative *Escherichia coli*, *Pseudomonas aeruginosa*, *Enterobacter* and *Klebsiella pneumonia* at (50 $\mu\text{g/ml}$ and 100 $\mu\text{g/ml}$) different concentration. Silver nanoparticles shown zones of inhibition 30 mm and 32 mm, 5 mm and 8 mm, 4 mm and 6 mm and 3 mm and 5 mm against *Escherichia coli*, *Pseudomonas aeruginosa*, *Enterobacter* and *Klebsiella pneumonia* respectively at the concentration of 50 $\mu\text{g/ml}$ and 100 $\mu\text{g/ml}$ (fig. 7). Both the concentration of silver nanoparticles have indicated practically comparative outcomes have appeared by standard ampicillin against *Escherichia coli* whereas other pathogens against which have failed to show significant results. Silver nanoparticles have shown their effect against *Escherichia coli* Gram-negative bacteria and it was suggested that the effect would be size and dose dependent [24].

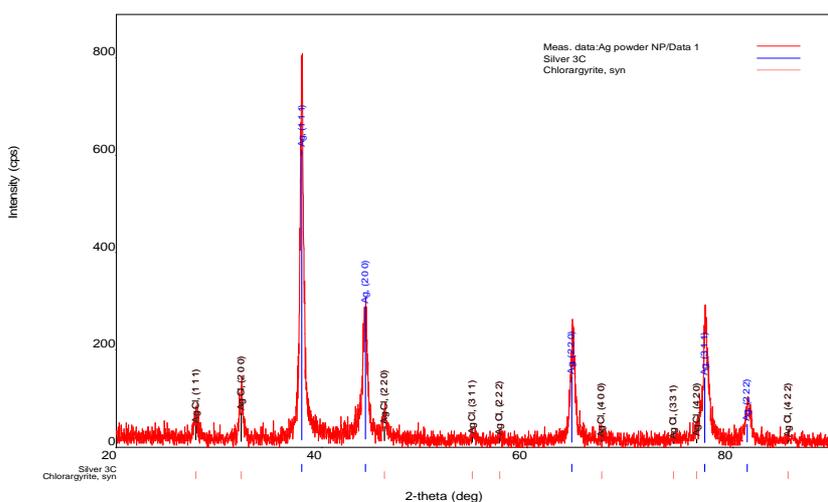


Fig. 6: X-ray diffraction crystallographic studies of silver nanoparticles from leaf extract of *Simarouba glauca*

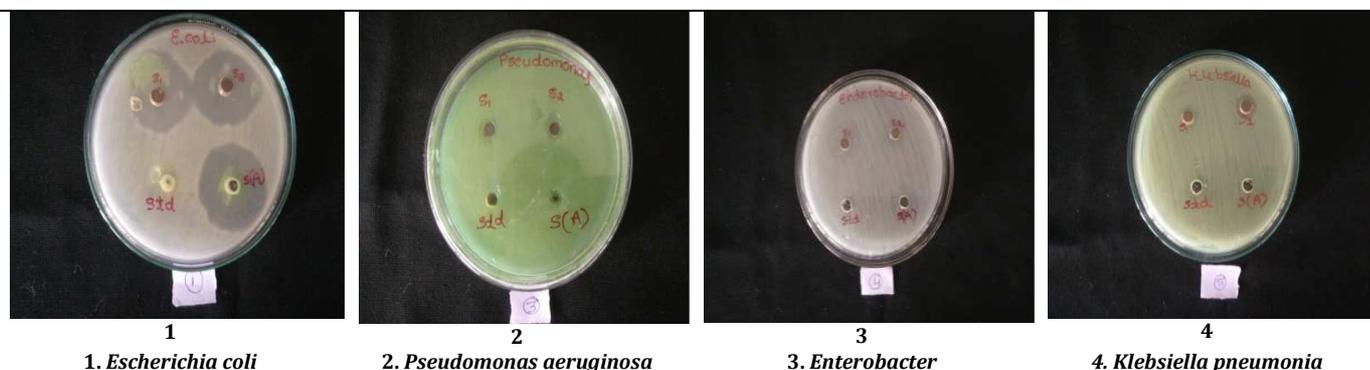


Fig. 7: Antibacterial activity of synthesized silver nanoparticles, S1-50 $\mu\text{g/ml}$. S2-100 $\mu\text{g/ml}$. Std-Distilled water. S(A)-Ampicillin

CONCLUSION

The medicinally significant aqueous leaf extract of *Simarouba glauca* was found to behave as a reducing agent for the production of silver nanoparticles. The synthesized silver nanoparticles have been characterized by UV-vis, SEM-EDAX and XRD to determine the sizes and shapes of the silver nanoparticles. The developed method is one of the excellent methods to produce silver nanoparticles in absence of toxic reducing chemicals. Our outcomes suggest that the biologically synthesized silver nanoparticles are most prominent against human pathogens.

CONFLICT OF INTERESTS

Declare none

REFERENCES

- Benjamin LO, Francesco S. Antibacterial activity of silver nanoparticles: a surface science insight. *Nano Today* 2015;10:339-54.
- Swarnali M, Deepak K, Gadadhar B, Sudip KG, Jayasree KL. Antimicrobial activities of silver nanoparticles synthesized from *Lycopersicon esculentum* extract. *J Anal Sci Technol* 2014;5:40:1-7.
- Parikh DV, Fink T, Rajasekharan K, Sachinvala ND, Sawhney APS, Calamari TA, *et al.* Antimicrobial silver/sodium carboxymethyl cotton dressings for burn wounds. *Text Res J* 2005;75:134-8.
- Ulkur E, Oncul O, Karagoz H, Yeniz E, Celikoz B. Comparison of silver-coated dressing (Acticoat™), chlorhexidine acetate 0.5% (Bactigrass®), and fusidic acid 2% (Fucidin®) for topical antibacterial effect in methicillin-resistant staphylococci-contaminated, full-skin-thickness rat burn wounds. *Burns* 2005;31:874-7.
- Jeong SH, Yeo SY, Yi SC. The effect of filler particle size on the antibacterial properties of compounded polymer/silver fibres. *J Mater Sci* 2005;40:5407-11.
- Lee HY, Park HK, Lee YM, Kim K, Park SB. A practical procedure for producing silver nanocoated fabric and its antibacterial evaluation for biomedical applications. *Chem Commun* 2007;28:2959-61.
- Yuranova T, Rincon AG, Bozzi A, Parra S, Pulgarin C, Albers P, *et al.* Antibacterial textiles prepared by RF-plasma and vacuum-UV mediated deposition of silver. *Photochem Photobiol* 2003;161:27-34.
- Samuel U, Guggenbichler JP. Prevention of catheter-related infections: the potential of a new nanosilver-impregnated catheter. *Int J Antimicrob Agents* 2004;23:75-8.
- Alt V, Bechert T, Streinrucke P, Wagener M, Seidel P, Dingeldein E, *et al.* Domanne E, schnettler r *an in vitro* assessment of the antibacterial properties and cytotoxicity of nanoparticulate silver bone cement. *Biomaterials* 2004;25:4383-91.
- Rupp ME, Fitzgerald T, Marion N, Helget V, Puumala S, Anderson JR, *et al.* Effect of silver-coated urinary catheters: efficacy, cost-effectiveness antimicrobial resistance. *Am J Infect Control* 2004;32:445-50.
- Chenousova S, Epple M. Silver as an antibacterial agent: ion, nanoparticle, and metal. *Angew Chem Int Ed Engl* 2013; 4:1636-53.
- Kalsen HJ. Historical review of the use of silver in the treatment of burns. I. Early uses. *Burns* 2000;26:2:117-30.
- Fuhrmann GF, Rothstein A. The mechanism of the partial inhibition of fermentation in yeast by nickel ions. *Biochim Biophys Acta* 1968;163:331-8.
- Miller LP, Mc Callan SEA. The toxic action of metal ions to fungus spores. *J Agric Food Chem* 1957;5:116-22.
- Rayman MK, LoT C, Sanwal BD. Transport of succinate in *Escherichia coli*. II. Characteristics of uptake and energy coupling with transport in membrane preparations. *J Biol Chem* 1972;247:6332-9.
- Schreurs WJ, Rosenberg H. Effect of silver ions on transport and retention of phosphate by *Escherichia coli*. *J Bacteriol* 1982;152:7-13.
- Joseph JD, Jeffrey TB, Jude CO, Peter MD, Oliver NM, Ibrahim M. Antimicrobial efficacy of biosynthesized silver nanoparticles from different solvent extracts of *Waltheria americana* root. *J Anal Sci Technol* 2016;7:23.
- Santhana LK, Sangeetha D, Sivamani S, Tamilarasan M, Rajesh TP, Anandraj B. *In vitro* antibacterial, antioxidant, haemolytic, thrombolytic activities and phytochemical analysis of *Simarouba glauca* leaves extracts. *Int J Pharm Sci Res* 2014;5:432-7.
- Nagaraj B, Agnieszka Sobczak-Kupiec, Rebecca IF, Salman D. Bioreduction of chloroaurate ions using fruit extract *Punicagranatum* (Pomegranate) for synthesis of highly stable gold nanoparticles and assessment of its antibacterial activity. *Micro Nano Lett* 2013;8:400-4.
- Nithya R, Ragunathan R. Synthesis of the silver nanoparticle using pleurotus *Sajorcaju* and its antimicrobial study. *Digest J Nanomaterials Biostructures* 2009;4:623-9.
- Chandrappa CP, Govindappa M, Chandrasekar N, Sonia S, Sepuri O, Channabasava R. Endophytic synthesis of silver chloride nanoparticles from *Penicillium* sp. of *Calophyllum apetalum*. *Adv Nat Sci: Nanosci Nanotechnol* 2016;7:1-5.
- Nagaraj B, Akber I, Yong RL. Preparation of Au and Ag nanoparticles using *Artemisia annua* and their *in vitro* antibacterial and tyrosinase inhibitory activities. *Mater Sci Eng Carbon* 2014;43:58-64.
- Baker RA, Tatum JH. Novel anthraquinones from stationary cultures of *Fusarium oxysporum*. *J Ferment Bioeng* 1998; 85:359-61.
- Mritunjai S, Shinjini S, Prasad S, Gambhir IS. Nanotechnology in medicine and antibacterial effect of silver nanoparticles. *Digest J Nanomater Biostructures* 2007;3:115-22.

How to cite this article

- CP Chandrappa, N Chandrasekar, M Govindappa, Chaitra Shanbhag, Uttam Kumar Singh, Jayashri Masarghal. Antibacterial activity of synthesized silver nanoparticles by *Simarouba glauca* against pathogenic bacteria. *Int J Curr Pharm Res* 2017;9(4):19-22.