

PHYTOCHEMISTRY AND PHARMACOLOGICAL BIO-ACTIVITIES OF *LANNEA COROMANDELICA*: A REVIEW

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Received: July 16 2022, Revised and Accepted: 29 August 2022

ABSTRACT

Lannea coromandelica is extensively employed as a medicinal herb across the globe. This research work focuses primarily on the phytochemistry, pharmaceuticals, and bio-exertion of *L. coromandelica*. In ethnomedicine, several members of this species function as astringents, aiding in anti-oxidant, anti-inflammatory, and anti-diabetic and are helpful in other physiological functions. In addition, *Lannea* species' therapeutic benefits were acknowledged in numerous scientific investigations. The conclusions derived from the current study corroborate that the therapeutic activities of *L. coromandelica* can be linked to the presence of a broad spectrum of secondary metabolites and bioactive phytochemicals such as phenols, flavonoids, tannins, and proteins, according to the current study. This review highlights phyto-assembly and phyto-activities such as a diuretic, cardiovascular, and bioactivities such as antiepileptic, antimicrobial, and anti-convulsant, and pharmaceutical activities such as anti-cancer, anti-diabetic, and anti-asthmatic properties.

Keywords: *Lannea coromandelica*, Phytochemistry, Ethnobotanical uses, Pharmacological properties, Flavonoids.

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INTRODUCTION

Plants, trees, herbs, shrubs, weeds, etc., are essential supplies of nutrition, protection, and medication for people. Discovering how plants can be used to address ailments is crucial to living a better life. Our predecessors utilized countless botanicals as remedies, and these herbal remedies are scientifically validated. *Lannea coromandelica* is one of the significant species that may be found throughout India. Indian territory undoubtedly has an abundant herbal reservoir that is able to mitigate and avert numerous illnesses [1-5]. At present, these therapeutic remedies are on par with manufactured medical products due to their minimal or no detrimental or harmful effects on humans.

L. coromandelica also termed as Jhingini in Sanskrit and woodier or Indian ash tree in English is a representative of the Anacardiaceae genus that pullulates in the tropical regions of the Asian continent. This species is said to be beneficial in ethnomedicine for local edema, soreness, irritation, inflammation, and redness, as well as for tumor growths, ulceration, sprains, injuries, dermal diseases, melanoma, and diarrhea [6]. In South India, it is addressed as Odiar or Gumphini. Indian tribes have maneuvered this species to target a variety of ailments, including bodily discomforts/aches, toothaches, stomachaches, gastritis, gastritis, and sexual sterility. Several studies on the plant's pharmacological applications have been conducted. Polyphenols, such as bioflavonoids, terpenes, gums, and saccharides, are found in abundance in the plant [7,8].

Herbal plant species contribute a significant role in generating antioxidants due to the prevalence of polyphenols. The secondary metabolites contribute significantly in elevating human health. The ingestion of various phenolic chemicals can result in serious medical conditions such as chronic and degenerative diseases. The reactive oxygen species will cause oxidative stress and injury. Micronutrients can either boost the ROS factors or limit the availability of free radicals [9].

The plant is also known as Bhadi, Jeol, Jialbhandi, Jhingangummi, Jhingam, and Indian Ash Tree in India. Tekeshwar K and Vishal J identified it in arid and humid forests of the Himalayas (Swat to Bhutan), Assam, Burma, Indo-China, Ceylon, Andamans, China,

Thailand, Cambodia, Laos, Vietnam, and Malaysia. The plant's chemical components, polyphenols, and flavonoids have been extracted in large quantities [10,11].

The North and West African communities have utilized these species as food additives and in pharmaceuticals. The slightly acidic fruits are used in culinary to make sweets and puddings. *L. coromandelica* is acknowledged as one of the pivotal herbs in the biosphere because of its anti-inflammatory, anti-oxidant, and anti-diabetic properties [12-16].

L. coromandelica is a versatile tree with a wide range of medicinal and pharmacological benefits. The review mainly focuses on the therapeutic phytochemical and pharmacological properties of *L. coromandelica*. Prior studies have shown that the bark of *L. coromandelica* is effective in the treatment of injuries, lesions, wound infections, scars, bruises, abrasions, ulcerations, gastric ulcers, enterocolitis, leukorrhea, eye infections, peptic ulcer disease, colitis, stomatitis, odontalgia, dislocations, sprains, diarrhea, and constipation, and that the foliage is efficacious in the management of elephantiasis, inflammatory disorders, and neuralgia [17].

PHYTO-ASSEMBLY

The genus *Lannea* is designated after Lannes de Montebello, a French who introduced the plant from Japan to France in 1870, while the species *coromandelica* is titled after Coromandel, a small town on the southern-eastern coast of India. Approximately 30–40 kinds of shrubs and trees belong to the genus *Lannea* which commonly flourishes in Africa e Except for *L. coromandelica* [18], which is grown in tropical and subtropical regions of Asia. It can reach a maximum of 14 m in areas with sufficient sunshine and adequate irrigation, and it has a red blaze. Barks are light-colored with branchlets that congregate at the ends of branches. Starry hairs cover the branchlets in minuscule amounts. Imparipinnate leaves are alternately placed. While juvenile, the leaflets are 5–7 in number, opposite, oval, sharp tip, whole edge, and coated with velvety hairs. Flowers are unisexual and have a green color. They have four broad oval sepals that are roughly 1 mm long [19,20]. Petals are four in number, 2 mm long, rectangular in shape, and greenish-yellow in color, with stamens twice as many as petals and turn yellow

during abscission. The tree blooms during February and April. The fruits are drupes with 3–5 locules and a dull red to pink colors [21].

L. coromandelica has alternative names such as kanbelimarnu and paalkaara, pithmari, pithmari, pithmari, and pithmari, and the plant's twigs are used as tooth sticks by local communities, the bark is used for skin diseases by Toraja people, tender leaves and roots are used for stomach ache by Toraja and Gadaba communities, and past research has revealed that the fruits of this plant are pulverized and combined with water and used as a fish poison. In Oriya, it's known as Raji Mohi, and tests found that the bark constituted flavonoids and terpenoids, but no alkaloids or steroids. The Garo tribes settled in the Madhapur forest region of Bangladesh employed the stem bark of *L. coromandelica*, known as Jiga Bark, for seminal insufficiency and profuse seminal discharges. The tribes of Mt. Yingying, Hainan Island, China, called the Indian ash tree Ziza and used it to cure wounds and hematochezia, the crushed and macerated secondary metabolites of the plant's leaves and bark were administered orally [22-25].

In Mizoram, India, the tree bark is called Tiataasha which is traditionally used as an astringent in ulcers and sores, while the leaf is effective in treating swellings, injuries, bruises, sprains, and body aches. Gumphini was the indigenous name for this plant in Srihari Kota Island, Nellore, India. Conventionally, the juice was used to treat ulcers and toothaches, while the stem bark was blended into a paste to treat body ailments and the wood was used to build agricultural implements. It is utilized spiritually to ward off evil spirits. Locals in Goa refer to it as Moi, and they employ the plant's bark in tanning. It was reported to be administered in medicine as an antidote for necrotic coma patients, as well as to relieve dyspepsia, sprains, and bruises [20,26,27].

The lambada tribes of Nizamabad District, Andhra Pradesh, India, utilize *dumpidi* plant bark as a dressing to cure bone fractures. People in Rajasthan, India, referred to *Lannea* as Jingini. Gelly was traditionally soaked in water and used to treat pain. The interior mass of wood bark was smashed, and the fluids were recovered and were used in the treatment of tetanus [28-32]. The total import value index of plants in the *Mota Magra* Forest in Udaipur, Rajasthan, was reported, and the values of plants growing in other forest zones were given in comparison. According to reports, the plant is as important as Neem, India's sacred tree. *L. coromandelica* hardwood is used to make bleachable pulp and to feed wild silkworms.

Scientists are investigating indigenous medicinal plant extracts on various physicochemical and biological paradigms to explore the bioactive components they possess and to examine their medicinal potential. Lipoma, scurvy, tumors, ulceration, cancer growths, bruises, injuries, dermal problems, dysentery, hematochezia, and constipation are all treated with leaf extracts. Other than that, the leaves are served as livestock fodder [33-35]. Food additives, flavoring agents, dental powders, and impregnating agents in fishnet stockings are the benefits attained from bark and its extracts/powders. Contusions, inflammatory diseases, chronic gastritis, canker sores, scars, skin rash, and other conditions are treated with the bark. Traditional applications, as well as the abovementioned pharmacological findings, support the plant's therapeutic potency [36,37].

PHYTO-CHEMISTRY AND CONSTITUENTS

The biochemical contents of all plant parts, including gums, stems, leaves, flowers, and bark, were examined. Carbohydrates, gums, proteins, glycosides, terpenoids, and polyphenols are among the major constituents found in the plant. Terpenoids and flavonoids were discovered in the qualitative phytochemical investigation of the barks. β -Sitosterol and physicochemical were found in the petroleum ether extract of stem bark. The subsequent hot chloroform extract of the preceding contained Physcionanthranol-B [38]. All the phytochemicals are predominantly present in the mother liquor. The ether extracts yielded β -sitosterol (H), while the acetone extract yielded leucocyanidin. Phenolics such as tannins, flavonoids, flavanols, flavanones, isoflavones,

and flavanol glycosides, have been found in abundance in the flowers. Tannins, such as ellagic acid and a flavonoid glycoside, Quercetin-3-arabinoside, were isolated from fresh flowers using ether and ethyl acetate fractioned alcoholic extract [39,40]. Isoquercitin was found in a crude alcoholic extract of flowers (g). The plant's leaves have been found to contain β -Sitosterol, Ellagic acid, Quercetin (a), Quercetin-3-arabinoside, leucocyanidin (c), and leucodelphinidin (d). Morin (e), a flavonoid aglycone, was also extracted from the plant, with the structure 3,5,7,2',4'-OH-flavone reported as the probable structures [41].

The qualitative examinations of the leaves indicated the presence of carbohydrates, alkaloids, glycosides, tannins, flavonoids, polyphenols, saponins, amino acids, gums, and mucilages in the water extract while triterpenoids were present only in the methanol extract. Whereas steroids, volatile oils, fats, and fixed oils were completely absent [42,43].

PHARMACOLOGICAL EFFECTS ON SECONDARY METABOLITES

Steroids, tannins, flavonoids, alkaloids, saponins, triterpenoids, and saponins were found in the phytochemical screening of extracts of *L. humilis* and *L. barberi*. Flavonoids, sugar, protein, triterpenoids, alkaloids, tannins, and phenols were found in *Lannea coromandelica* bark extracts. Chemical analysis revealed that *Lannea welwitschia* consists of oil, sugar metabolites, alkaloids, saponins, and tannins. Only flavonoids and tannins were extracted from the plant leaves of *Lannea kerstingii* using ethyl acetate [44,45]. The petroleum crude ether extract solely includes steroids and triterpenes. Terpenes, carbohydrates, glycosides, alkaloids, flavonoids, fats, oils, waxes, and tannins were found in the leaf's extracts of *L. coromandelica* including 4'-methoxymyricetin 3-O- α -L-rhamnopyranoside, myricetin (3-O- α -L-rh), amnopyranoside, myricetin 3-O- β -D-glucopyranoside, vitexin, isovitexin, gallic acid, and epicatechin have been considered as major components leaf extract of *Lannea macrocarpa* [46-48].

PHYSICO-CHEMICAL ENTERPRISE

ANTIOXIDANT ACTIVITY

Phytochemical screening of odina woodier leaf and *in vitro* antioxidant study of DPPH radical scavenging at different concentrations in methanol and ethyl acetate extract of odina woodier leaf was measured. With the increase in concentration, the radical scavenging effect was discovered to increase. Using DPPH, the previous research described the varied quantities of ethanolic extract of bark in terms of radical scavenging activity. The IC50 value for ethanolic extract of bark was 83.28 μ g/mL [57].

The leaves and barks of *L. coromandelica* are rich in flavonoids and other phenolic compounds. The solubility of phenolic compounds validates their polar nature. The DPPH assay showed maximum absorption at 517 nm which subsequently declined with the presence of anti-oxidants in the medium [58]. The ethyl acetate fractions of bark exhibited the most effective free radical scavenging properties with the least IC50 value corresponding to 3.8 μ g/ml. The antioxidant capacity of the ethyl acetate fractions of the leaves and barks was reported to be 46.67 mg and 33.78 mg of L-ascorbic acid/dried extract as per the results of a study conducted [58].

The extracts' free-radical scavenging ability toward stable DPPH was measured using a spectrophotometer at 515 nm. The ethyl acetate extract exceeded the methyl alcohol extract and aqueous fractions in terms of DPPH scavenging activity. The DPPH scavenging efficacy of both standards and plant extracts decreased at a dosage of 100 mg/mL. The reducing power assay is frequently used to assess an antioxidant's ability to give electrons. Ascorbic acid was used as a reference to equate the reducing potential of the leaf extraction [59-61]. With the increase in concentration, the reducing power of all samples increased significantly. The most frequent free radical produced is the superoxide anion. Superoxide radical-scavenging capabilities of *L. coromandelica*

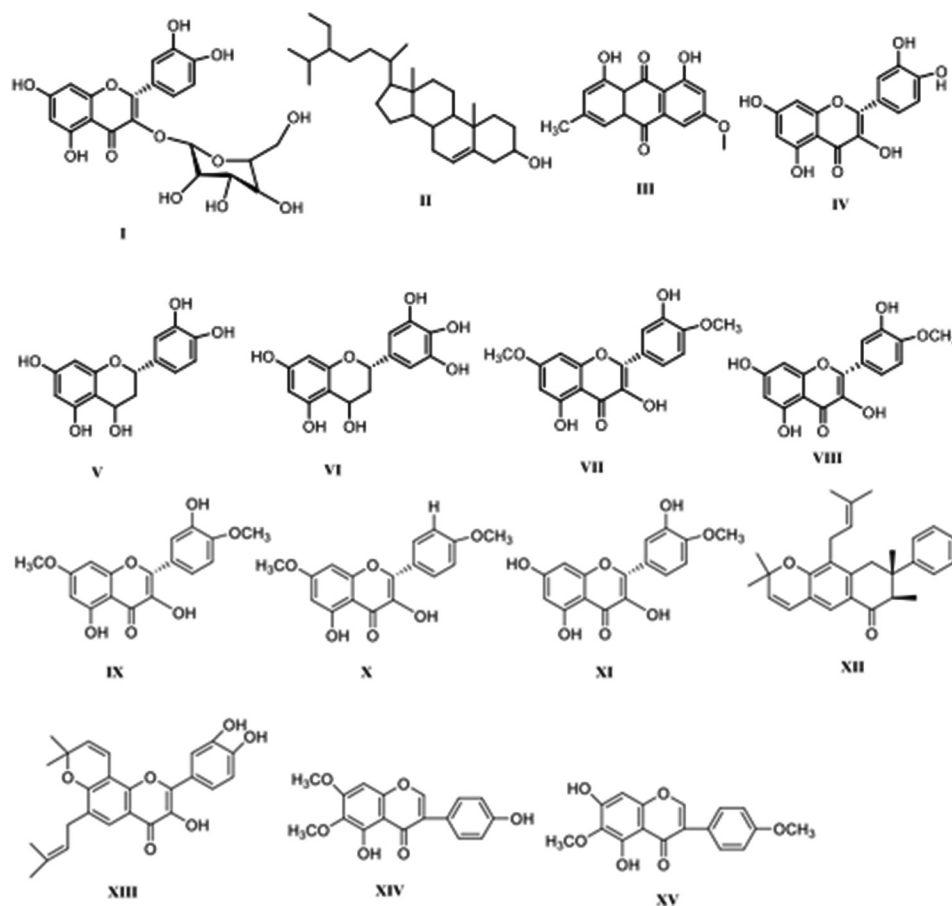


Fig. 1: Chemical structures of phytoconstituents isolated from *Lannea* species [54]

Table 1: Traditional and Medicinal uses of *L. coromandelica* [3,10,49-52]

Region	Local name	Plant part used	Medicinal uses
Khoraput district of Orissa	Kondh tribe-Kanbeli	Twigs	Tooth sticks
	Poraja tribe-paalkaara, pithmari	Bark	Skin diseases
	Gadaba tribe-pithmari	Leaves and roots	Stomach ache
Gadchiroli district of Maharashtra	Gond tribes- gopid	Fruits	Fish poison
Madhapur forest region of Bangladesh	Garos tribes-Jiga	Bark	Seminal weakness and excessive seminal emissions.
Tingeing, Hainan Island, China.	Ziza	Decoction and macerations of leaves and bark	Injuries and Hematochezia
Mizoram, India	Tawitawsuak	Bark	Astringent, in ulcers and sore, leaf is used in swellings, sprains and Pain of the body
Srihari Kota island, Nellore dist., India	Gumphini	Leaf juice	Ulcers, applied to treat tooth ache
Goa, India	Moi	Stem bark paste	Body pains
		Wood	To make agricultural implements
		Bark	Tanning, as antidote for coma patient caused by narcotics, and to treat dyspepsia, general debility, gout, Dysentery, sore eyes, leprosy, sprains and bruises.
Nizamabad District, Andhra Pradesh, India	Lambada tribes- Dumpidi	Bark	Bandage to treat bone fracture, impotency
Rajasthan, India	Jingini	Gum- soaked in water, rubbed on stone and applied locally	Pain
		The inner bark of stem- crushed and make suspension juice is applied over cuts	Stops bleeding and to prevent tetanus

L. coromandelica: *Lannea coromandelica*

extracts were in a dosage-dependent manner. The elimination of Fe³⁺ is a crucial component of transferring electrons and a fundamental step in exerting antioxidant properties. In the extracts of *L. coromandelica*, the (Ferric Reducing Antioxidant Assay) test indicated a favorable connection between reducing power and the total concentration of phenols. The antioxidant capacity index for polyphenolic compounds, flavonoid, ascorbic acid, and tocopherol is frequently measured using chromogenic oxidizing agents. According to the findings, the cupric reducing power of the ethanolic fraction was the highest, while the aqueous fraction had the lowest concentration [62].

ANTIMICROBIAL ACTIVITY AND WOUND HEALING

Gouranga das and Durbadal Ojha examined the antimicrobial activity of various extracts of cassia toralinn plant leaves in 2010. The antibacterial and wound healing activities of the bark of *L. coromandelica* were determined by the disc diffusion method by [63-65]. On mice, it demonstrates substantial wound healing activity.

The isolates were evaluated for antibacterial activity (ABA) against 11 bacterial species (*Bacillus subtilis*, *Bacillus cereus*, *Sarcina lutea*, *Staphylococcus aureus*, *Shigella boydii*, *Vibrio mimicus*, *Shigella dysentery*, *Escherichia coli*, *Salmonella typhi*, *Salmonella paratyphi*, and *Pseudomonas aeruginosa*) and contrasted to the reference antibiotic cephadrine disc by measuring the zone of inhibition (in mm). Antifungal activity was also evaluated against two fungi (*Candida albicans* and *Saccharomyces cerevisiae*) [45,66], and the results were comparable to nystatin. In comparison to the reference, dichloromethane portions of both bark and leaves demonstrated better ABA against both Gram +/- bacteria. Furthermore, leaf portions exerted stronger anti-bacterial properties than bark fractions. The dichloromethane soluble fractions of leaves demonstrated the highest ABA against *S. paratyphi*. Most of the other fractions exhibited minimal or no antifungal properties [67].

Table 2: List of the constituents and phytochemicals

Phytochemicals	References
4-O- (a-D-galactopyranosyluronic acid)-D-galactose	[20]
6-O- (β-Dglucopyranosyluronic acid)-D-galactose	[20]
6-O-(4-O-methyl-Dglucopyranosyluronic acid)- D-galactose	
3-O-β-L-arabinofuranosyl-L-arabinose	[53]
3-a-D-galactopyranosyl-L-arabinose	[53]
3-O-β-D-galactopyranosyl-D-galactose	[53]
6-O-β-Dgalactopyranosyl-D-galactose	[33]
4-O-methyl derivatives namely	[33]
2,3,4-tri-O-methyl-Lrhamnose	
2,3,5- and 2,3,4- tri- and 2,5-di-O-methyl-L-arabinose	[54]

Aspergillus flavus, *Aspergillus niger*, *Colletotrichum gloeosporioides*, and *Alternaria alternata* were investigated for their effectiveness in the test dermatophytic fungus *C. albicans* and *Malassezia pachydermic* with ethyl ethanoate fractions of fungal endophytes derived from *L. coromandelica*. *A. flavus* has a maximal inhibitory region of 26.22 mm toward *C. albicans* and 16.72 mm toward *M. pachydermic*. *C. albicans* showed substantial suppression, while *A. niger* and *A. alternata* showed no significant inhibition. A considerable inhibitory zone was found in *M. pachydermic* and *C. gloeosporioides* [68-71].

ANTIDIABETIC AND ANALGESIC ACTIVITY

Past investigations have shown that an ethyl alcohol extract of *L. coromandelica* bark has an anti-hyperglycemic effect in mouse models and demonstrates significant analgesic properties in heat-induced animals. The ability of the methanolic isolates of *L. coromandelica* bark to tolerate hyperglycemia was investigated. The mice were orally fed with 2 g glucose/kg of body weight, blood samples were taken after 2 h to test blood glucose levels by glucose oxidase methodology. Glibenclamide, a standard antidiabetic medication, was given to the control groups at a concentration of 10 mg/body weight [72]. At a concentration of 200 and 400 mg/kg body weight, the glucose-loaded mice treated with the bark extracts of *L. coromandelica* displayed significant antidiabetic action [73].

On 3 weeks of oral ingestion of *L. coromandelica* at concentrations of 250 and 500 mg/kg, hyperglycemic mice showed a notable reduction in blood sugar levels both in acute and long-term - term studies, and a remarkable reduction in total cholesterol, total triglycerides, low-density lipoprotein cholesterol, and an insubstantial improvement in high-density lipoprotein cholesterol levels were noticed [74]. In comparison to the treated diabetic control group, oral consumption of *L. coromandelica* dramatically lowered creatinine levels, alanine aminotransferase, and aspartate transaminase. These findings revealed a significant enhancement from the detrimental impact of alloxan on the pancreatic islet cells [75].

The extracts have been reported to effectively block glucagon-assisted cAMP synthesis, and are ineffective in lowering forskolin-induced cAMP production. The availability of numerous bioactive elements such as flavonoids, glycosides, and alkaloids in the ethyl alcohol leaf fractions of *L. coromandelica* resulted in a substantial hypolipidemic impact at a concentration of 200 mg/kg body weight [76].

OTHER THERAPEUTIC PROPERTIES

Antinociceptive effects

Chemicals and temperature-induced pain paradigms, such as acetic acid-induced writhing, hot plate, tail immersion, formalin, and glutamate testing's, were used to assess the antinociceptive effect of an

Table 3: Bio-exertion of biological activities

Hepatoprotective	Ethanolic extract of <i>M. oleifera</i> leaves as well as alcoholic extract of seed in perspective of hepatoprotective (<i>in vivo</i>) was estimated contrary to the rifampicin and pyrazinamide evoked liver damage ^[5] and also hepatoprotective effect contrary to antitubercular medical pills and alloxan evoked liver damage noted in diabetic rats
Antifungal Activity	Antimicrobial activity of the ethanolic and aqueous extracts of <i>L. coromandelica</i> on three microbes (two bacteria and one fungus) by agar well diffusion method. On the basis of previous studies, we can conclude that <i>L. coromandelica</i> bark extract having antimicrobial property and antifungal property against the female RTI which caused by some pathogenic microorganism. Extracted material shows the relevant stronger activity against all tested bacterial stains. However, aqueous extract demonstrated higher antifungal activity [55]. Therefore, <i>L. coromandelica</i> has been used for further analysis.
CNS enterprise	Leaves extract of <i>M. oleifera</i> restores the capability of amine levels of the central nervous system which will be helpful in Alzheimer's sickness and <i>in vitro</i> anticonvulsant enterprise from the liquid extract of <i>Moringa oleifera</i> roots [8].
Antidiabetic activity	The study showed that Tropolpropanaltosylhydrazone could be used as a potential lead compound in the treatment of diabetes. Further exploration of the function of the compound will facilitate a better understanding towards development of Tropolpropanaltosylhydrazone ^[50] as an antidiabetic agent.
Hypolipidemic Activity	In terms of hypolipidemic activities, the indigenous products can be inter-related with the secondary metabolites such as flavonoids, tannins, carbohydrates, and so on. It acts as inhibitor for biosynthesis and absorption of lipogenic and lipolytic enzymes due to lipid enzymes [56].

ethyl alcohol extract of *L. coromandelica* leaves. In the chemical/heat-induced nociception in murine models, the ethyl alcohol fractions of *L. coromandelica* showed strong dose-dependent antinociceptive action, confirming the leaf's longstanding usage in the healing of many painful disorders. Another scientific study found that the ethanolic extract of *L. coromandelica* produced a substantial dose-dependent increase in hot plate latencies and tail retraction duration. It also decreased the number of stomach constraints and paw lickings caused by acetic acid [77-79].

Anti-arthritis effects

In terms of ABA, preliminary phytochemical research, *in vitro* antioxidant, and anti-arthritis tests reveal that the ethanolic extract of *L. coromandelica* leaves has increased anti-arthritis potency [80].

HYPERTENSIVE REACTIVITY

It showed a reduction in arterial blood pressure so can be used as a hypotensive agent. It was found no effect in vagotomy and eviscerated dogs, but spinal preparation showed a slight increase in hypotension [81].

WOUND HEALING ACTIVITY

Ethyl alcohol and propanone fractions of *L. coromandelica* (Houtt) Merr bark hinder remediation activity in excision and incision form when treated to male rats in conventional ointments. For both procedures, framycetin sulfate was used as a standard. In the excision approach, ethanolic and acetone extracts showed 97.11% and 95.95% activity, respectively, and showed long-term effects in the incision method [82].

CONCLUSION

The main objective of this review was to resolve the pharmacological and medicinal activities of *L. coromandelica* that exposed plant possesses analgesic medication, anti-inflammatory, antipyretic, and antioxidant. These activities are commenced in its root, stem, leaf, and seeds. It states that any of the solvent either ethanolic or methanolic extracts is widely used for examination and identification purposes and in the future, principally *in vivo* studies based on animal models may be considered for higher effective results.

CONFLICT OF INTEREST

Nil.

REFERENCES

- Achika J, Ayo R, Oyewale A, James D. Chemical investigation and antioxidant activity of fractions of *Lannea humilis* (Oliv.) Engl. J Turk Chem Soc Sec A Chem 2017;4:563-72.
- Alam B, Hossain S, Habib R, Rea J, Islam A. Antioxidant and analgesic activities of *Lannea coromandelica* Linn. Bark extract. Int J Pharmacol 2012;8:224-33.
- Anderson DM, Hendrie A. The structure of *Lannea coromandelica* gum. Carbohydr Res 1973;26:105-15.
- Antimicrobial Activity of *Anogeissus leiocarpus* and *Lannea microcarpa* on Some Microbes Isolated from Vegetables in Sokoto. Dubai, UAE: International Conference on Chemical, Environmental and Biological Sciences; 2015.
- Kumar RB. Flora of sacred groves at Sriharikota Island, Andhra Pradesh, India. Ethnobot Leaflet 2010;14:420-6.
- Bingchun G, Rongtao L, Xinquan Y, Daolin D. Ethnobotanical study on medicinal plants used by Li nationality in Wuzhishan area of Hainan province. Afr J Pharm Pharmacol 2007;7:???
- Chandrajith VG, Marapana R. Physicochemical characteristics of bark exudates of *Lannea coromandelica* and its application as a natural fruit coating. J Pharmacogn Phytochem 2018;7:1798-1802.
- Clarke AE, Anderson RL, Stone BA. Form and function of arabinogalactans and arabinogalactan-proteins. Phytochemistry 1979;18:521-40.
- Das G, Ojha D, Bhattacharya B, Samanta M, Ghosh S, Datta S, et al. Evaluation of antimicrobial potentialities of leaves extracts of the plant *Cassia tora* Linn. (Leguminosae/Caesalpinioideae). J Phytol 2010;2:64-72.
- Dinesh V, Sharma P. Traditional uses of plants in indigenous folklore of Nizamabad district, Andhra Pradesh, India. Ethnobot Leaflet 2010;14:29-45.
- Franco FM, Narasimhan D. Plant names and uses as indicators of knowledge patterns. Indian J Tradit Knowl 2009;8:645-8.
- Galanki V, Venkatesham A, Chitturi D. Antidiabetic activity of *Lannea coromandelica* Houtt leaves in alloxan-induced diabetic rats. Int J Pharm Biol Sci 2014;4:108-14.
- Muhaisen HM. Chemical constituents from the bark of *Lannea acida* rich (Anacardiaceae). Pharm Chem 2013;5:88-96.
- Hossain J, Biswas S, Shahriar M, Chowdhury MM, Islam S, Chowdhury RA. Phytochemical screening, antimicrobial activity, antioxidant capacity, and *in vivo* anticancer activity of *Lannea coromandelica* bark extracts. IOSR J Pharm Biol Sci 2018;13:19-25.
- Imam MZ, Moniruzzaman M. Antinociceptive effect of ethanol extract of leaves of *Lannea coromandelica*. J Ethnopharmacol 2014;154:109-15.
- Islam MT, Tahara S. Dihydroflavonols from *Lannea coromandelica*. Phytochemistry 2000;54:901-7.
- Islam MT, Ito T, Sakasai M, Tahara S. Zoosporicidal activity of polyflavonoid Tannin identified in *Lannea coromandelica* stem bark against phytopathogenic oomycete *Aphanomyces cochlioides*. J Agric Food Chem 2002;50:6697-703.
- Islam R, Khan AS, Benozir S, Islam S, Alam J. Anti-diabetic properties of *Lannea coromandelica* L. bark extract on alloxan induced Type-2 diabetic rats. Eur J Pharm Med Res 2018;5:31-8.
- John HB. Allergic Asian *Anacardiaceae*. Clin Dermatol 1986;4:191-203.
- Manzur-Ul-Kadir Mia M, Kadir MF, Hossain MS, Rahmatullah M. Medicinal plants of the Garo tribe inhabiting the Madhupur forest region of Bangladesh. Am Eur J Sustain Agric 2009;3:165-71.
- Kamalkishor HN, Kulkarni KM. Fish stupefying plants used by the Gond tribal of Mendha village of Central India. Indian J Tradit Knowl 2009;8:531-4.
- Kantamreddi V, Lakshmi YN, Kasapu VV. Preliminary phytochemical analysis of some important Indian plant species. Int J Pharm Biol Sci 2010;1:358.
- Kaur R, Jaiswal ML, Jain V. Protective effect of *Lannea coromandelica* Houtt. Merrill. Against three common pathogens. J Ayurveda Integr Med 2013;4:224-8.
- Kumar T, Jain V. Appraisal of total phenol, flavonoid contents, and antioxidant potential of folkloric *Lannea coromandelica* using *in vitro* and *in vivo* assays. Scientifica 2015;2015:203679.
- Lestari DW, Satria Y. The Utilization of Kudo Bark (*Lannea coromandelica*) as the Source of Natural Dye in Dyeing of Silk Batik. Vol. 1. Indonesia: Proceeding Indonesian Textile Conference; 2019. p. 1-6.
- Manik MK, Wahid MA, Islam SM, Pal A, Ahmed KT. A comparative study of the antioxidant, antimicrobial and thrombolytic activity of the bark and leaves of *Lannea coromandelica* (anacardiaceae). Int J Pharma Sci Res 2013;19:2609-14.
- Mannan A, Das H, Rahman M, Jesmin J, Siddika A, Rahman M, et al. Antihyperglycemic activity evaluation of *Leucas aspera* (Willd.) link leaf and stem and *Lannea coromandelica* (Houtt.) Merr. bark extract in mice. Adv Nat Appl Sci 2010;4:385-8.
- Stanislaus N, Sule MI, Pateh UU, Hassan HS, Ahmed MM, Abdullahi ST, et al. Phytochemical and antimicrobial activity of the leaves of *Lannea kerstingii* Engl and k. Krause (Anacardiaceae). J Health Allied Sci NU 2014;4:4-9.
- Olatokunboh AO, Mofomosara SH, Ekene OA. Evaluation of the anti-diarrhoeal effect of *Lannea welwitschii* Hiern (Anacardiaceae) bark extract. Afr J Pharm Pharmacol 2010;4:165-9.
- Picerno P, Mencherini T, Loggia RD, Meloni M, Sanogo R, Aquino RP. An extract of *Lannea microcarpa*: Composition, activity, and evaluation of cutaneous irritation in cell cultures and reconstituted human epidermis. J Pharm Pharmacol 2006;58:981-8.
- Prabhakaran M, Sundarapandian SM, Elumalai S. Phytochemical studies and antibacterial activity of the leaf, bark, and root powders of *Odina wodier* Roxb. Biosci Biotech Res Asia 2010;7:857-77.
- Premjanu N, Jaynthy C, Diviya S. Antifungal activity of endophytic fungi isolated from *Lannea coromandelica*-an *in silico* approach. Int J Pharm Pharm Sci 2016;8:207-10.
- Rahman MS, Begum B, Chowdhury R, Rahman KM, Rashid MA. Preliminary cytotoxicity screening of some medicinal plants of Bangladesh. Dhaka Univ J Pharm Sci 2008;7:47-52.
- Rahman M, Khatun A, Uddin SJ, Shilpi JA. Comparative effect of *Lannea coromandelica* (Houtt.) Merr. leaves and stem barks on acetic acid-induced pain model in mice and chromogenic reagents: Exploring the analgesic potential and phytochemical groups. Pharmacol Online 2016;1:146-52.
- Ramachandran R, Joshi BC. Examination of the gum from *Lannea coromandelica*. Phytochemistry 1968;7:2057-9.

36. Rao VS, Einstein JW, Das K. Hepatoprotective and antioxidant activity of *Lannea coromandelica* Linn on thioacetamide induced hepatotoxicity in rats. *Int Lett Nat Sci* 2014;8:30-43.
37. Reddy AK, Joy JM, Kumar CK. *Lannea coromandelica*: The researcher's tree. *J Pharm Res* 2011;4:577-9.
38. Sathish R, Ahmed MH, Natarajan K, Lalitha KG. Evaluation of wound healing and antimicrobial activity of *Lannea coromandelica* (Houtt) Merr. *J Pharm Res* 2010;3:1225-8.
39. Selvaraj D, Kotapadu A, Sampurna B, Balaji P, Abraham AA, Kesavan SK, et al. Detection of active constituents from the leaf extract of *Lannea coromandelica* by GC-MS testing and assessment of its pharmacological activity. *Int J Pharm Sci Res* 2015;1:1217-21.
40. Seth MK. Trees and their economic importance. *Bot Rev* 2003;69:321-76.
41. Shahriyar SA, Sultana N, Haque M, Islam SM. Antihyperglycemic and antinociceptive activity of *Lannea coromandelica* (houtt.) Merr. *Bark in vivo*. *World J Pharm Pharm Sci* 2016;5:171-84.
42. Shrimal RL, Vyas LN. Community structure of the Mota Magra forest block (Udaipur). *Vegetatio* 1977;34:127-32.
43. Singh S, Singh GB. Hypotensive activity of *Lannea coromandelica* bark extract. *Phytother Res* 1996;10:429-30.
44. Subramanian SS, Nair AG. Polyphenols of *Lannea coromandelica*. *Phytochemistry* 1971;10:1939-40.
45. Upadhyay B, Parveen, Dhaker AK, Kumar A. Ethnomedicinal and ethnopharmaco-statistical studies of Eastern Rajasthan, India. *J Ethnopharmacol* 2010;129:64-86.
46. Vadivel K, Thangabalan B, Chetanajessygrace B, Kumar DP, Manoharbabu S, Narayana KV. Preliminary phytochemical evaluation of leaf extracts of *Lannea coromandelica* L. *Int J Pharmacol Res* 2012;2:64-68.
47. Valli G, Jeyalakshmi M. Preliminary phytochemical and antioxidant study of *Odina woodier* leaf extract. *Asian J Pharm Res* 2012;2:153-5.
48. Venkatesham A, Vasantha G, Dayakar CH. Hypolipidemic activity of *Lannea coromandelica* Houtt. leaves in triton X-100 induced hyperlipidemic rats. *Indian J Pharm Pharmacol* 2019;5:207-10.
49. Walunj S, Gupta R, Joshi S, Sabharwal S, Joshi K. *Lannea coromandelica* attenuates glucagon and oxyntomodulin mediated cAMP formation in HEK cells stably-expressing human glucagon receptors. *J Herb Med* 2015;5:153-7.
50. Wollenweber E, Dietz VH. Occurrence and distribution of free flavonoid aglycones in plants. *Phytochemistry* 1981;20:869-932.
51. Yun XJ, Shu HM, Chen GY, Ji MH, Ding JY. Chemical constituents from barks of *Lannea coromandelica*. *Chin Herb Med* 2014;6:65-9.
52. Wachtel-Galor S, Benzie IF. Herbal medicine An Introduction to Its History, Usage, Regulation, Current Trends, and Research Needs. Florida, United States: CRC Press/Taylor and Francis; 2011.
53. Ghasemian M, Owlia MB. A different look at pulsed glucocorticoid protocols; is high dose oral prednisolone necessary just after initiation of pulse therapy? *J Case Rep Pract* 2015;13:1-3.
54. Rai PK, Lalramnghinglova H. Ethnomedicinal plant resources of Mizoram, India: Implication of traditional knowledge in health care system. *Ethnobot Leaflet* 2010;14:274-305.
55. Beaman JH. Allergenic Asian *Anacardiaceae*. *Clin Dermatol* 1986;4:191-203.
56. Garcia-Ruiz JC, Amutio E, Ponta J. Invasive fungal infection in immunocompromised patients. *Rev Iberoam Micol* 2004;21:55-62.
57. Dismukes WE. Introduction to antifungal drugs. *Clin Infect Dis* 2000;30:653-7.
58. Tan RX, Zou WX. Endophytes: A rich source of functional metabolites. *Nat Prod Rep* 2001;18:448-59.
59. Strobel GA. Rainforest endophytes and bioactive products. *Crit Rev Biotechnol* 2002;22:315-8.
60. Ding G, Liu S, Guo L, Zhou Y, Che Y. Antifungal metabolites from the plant endophytic fungus *Pestalotiopsis foedan*. *J Nat Prod* 2008;71:615-8.
61. Strobel G, Daisy B. Bioprocessing for microbial endophytes and their natural products. *Microbiol Mol Biol Rev* 2003;67:491-502.
62. Mousa WK, Raizada MN. The diversity of anti-microbial secondary metabolites produced by fungal endophytes: An interdisciplinary perspective. *Front Microbiol* 2013;4:65.
63. Donadio S, Monciardini P, Alduina R, Mazza P, Chiocchini C, Cavaletti L, et al. Microbial technologies for the discovery of novel bioactive metabolites. *J Biotechnol* 2002;99:187-98.
64. Queiroz EF, Kuhl C, Terreaux C, Mavi S, Hostettmann K. New dihydroalkylhexenones from *Lannea edulis*. *J Nat Prod* 2003;66:578-80.
65. Merlin FF, Narasimhan D. Plant names and uses as indicators of knowledge patterns. *Indian J Tradit Knowl* 2009;8:645-8.
66. Rajagopal K. Endophytic fungi of a palm, *Borassus flabellifer* L. A first report from India. *Asian J Microbiol Biotechnol Environ Sci* 2004;6:541-4.
67. Venkatachalam CM, Jiang X, Oldfield T, Waldman M. Ligandfit: A novel method for the shape-directed rapid docking of ligands to protein active sites. *J Mol Graph Model* 2003;21:289-91.
68. Wicklow DT, Roth S, Deyrup ST, Gloer JB. A protective endophyte of maize: *Acremonium zeae* antibiotics inhibitory to *Aspergillus flavus* and *Fusarium verticillioides*. *Mycol Res* 2005;109:610-8.
69. Park JH, Choi GJ, Lee SW, Jang KS, Choi YH, Cho KY, et al. Screening for antifungal endophytic fungi against six plant pathogenic fungi. *Mycobiology* 2003;31:179-82.
70. Liu L, Liu S, Niu S, Guo L, Chen X, Che Y. Isoprenylated chromone derivatives from the plant endophytic fungus *Pestalotiopsis fici*. *J Nat Prod* 2009;72:1482-6.
71. Shekhawat KK, Rao DV, Batra A. *In vitro* antimicrobial activities of endophytic fungi isolates from medicinal tree-*Melia azedarach* L. *J Microbiol Res* 2013;3:19-24.
72. Liu JY, Song YC, Zhang Z, Wang L, Guo ZJ, Zou WX, et al. *Aspergillus fumigatus* CY018, an endophytic fungus in cynodon dactylon as a versatile producer of new and bioactive metabolites. *J Biotechnol* 2004;114:279-87.
73. Zhang CL, Zheng BQ, Lao JP, Mao LJ, Chen SY, Kubicek CP, et al. Clavatul and patulin formation as the antagonistic principle of *Aspergillus clavatonanicus*, an endophytic fungus of *Taxus mairei*. *Appl Microbiol Biotechnol* 2008;78:833-40.
74. Zhang YX, Li M, Wang CY, Wang BG. A new naphtha quinoneimine derivative from the marine algal-derived endophytic fungus *Aspergillus niger* EN-13. *Chin Chem Lett* 2007;18:951-3.
75. Loesgen S, Magull J, Schulz B, Draeger S, Zeeck A. Isofusidienols: Novel chromone-3-oxepines produced by the endophytic fungus *Chalara* sp. *Eur J Org Chem* 2008;4:698-703.
76. Kim JH, Campbell BC, Chan KL, Mahoney N, Haff RP. Synergism of antifungal activity between mitochondrial respiration inhibitors and kojic acid. *Molecules* 2013;18:1564-81.
77. Hoekstra WJ, Garvey EP, Moore WR, Rafferty SW, Yates CM, Schotzinger RJ. Design and optimization of highly-selective fungal CYP51 inhibitors. *Bioorg Med Chem Lett* 2014;24:3455-8.
78. Strushkevich N, Usanov SA, Park HW. 3 Structural basis of human CYP51 inhibition by antifungal azoles. *J Mol Biol* 2010;397:1067-78.
79. Toda M, Okubo S, Ohnishi R, Shimamura T. Antibacterial and bactericidal activity of Japanese green tea. *Nihon Saikingaku Zasshi* 1989;44:669-72.
80. Fernandez MA, Garcia MD, Saenz MT. Antibacterial activity of the phenolic acids fraction of *Scrophularia frutescens* and *Scrophularia sambusifolia*. *J Ethnopharmacol* 1996;53:11-4.
81. Hoult JR, Paya M. Pharmacological and biochemical actions of simple coumarins: Natural products with therapeutic potential. *Gen Pharmacol* 1996;27:713-22.
82. Krishnaiah PV, Ratnam KV, Raju RR. Preliminary phytochemical evaluation of certain anticancer crude drugs used by adivasis of Rayalaseema Region, Andhra Pradesh, India. *Ethnobot Leaflet* 2008;12:693-7.