

Original Article

AN INNOVATIVE STRATEGY BASED ON UNCERTAINTY PROFILE FOR THE VALIDATION OF MICROBIOLOGICAL METHODS FOR COUNTING ENTEROBACTERIACEAE IN FOODS

FAYSSAL JHILAL^{a*}, BOUCHAIB IHSSANE^b, MOUHCINE SFAIRA^a, EL-MESTafa EL-HADRAMI^b, HOUDA BOUCHAFRA^b, SAAD S. ALAOU^b, YASSINE H. BENCHAKROUN^b, HASNAA HAIDARA^b, OMAR A. HAMEDANE^b, TAOUFIQ SAFFAJ^b

^aEngineering Materials, Environment and Modeling Laboratory (LIMME). Faculty of Sciences Dhar El Mahraz BP 1796 Atlas Fez, Morocco,
^bApplication Organic Chemistry Laboratory, Faculty of Sciences and Techniques, BP 2202 route d'immouzer-Fès-University Sidi Mohamed Ben Abedallah-Morocco
Email: fayssaljhilal@gmail.com

Received: 20 Dec 2015 Revised and Accepted: 27 Feb 2016

ABSTRACT

Objective: A new and powerful statistical approach known as the uncertainty profile concept has been suggested for both testing the validity and making easy and straightforward interpretation of results obtained during the validation of an analytical method. The main goal of this paper is to confirm the applicability of this new strategy for the validation of a commercial kit, microbiological method, for the enumeration of the *Enterobacteriaceae* in foods and the estimate of the measurement uncertainty by using the newly provided formula and without referring to any additional experiments.

Methods: An innovative formula to assess the uncertainty by using validation data and without recourse to other additional experiments was proposed. The uncertainty was evaluated through the two-sided β -content, γ -confidence tolerance interval, which is computed with three manners: the Mee's approach, the Generalized Pivotal Confidence, and the Modified Large Simple procedure

Results: After the use of the three chemometric method of calculation of tolerance intervals, the obtained results with uncertainty profile show without doubt that the enumeration method is valid over the range of target values given that the upper and the lower 66.7 %-content, 90 %-confidence tolerance limits have fallen within the two acceptance limits of ± 0.25 Log unit. If the β is stretched to 80 %-content, 90 %-confidence, the three computed tolerance intervals lead to different decisions.

Conclusion: we have demonstrated the ability of the uncertainty profile to be used for testing the validity of enumeration method which represents the first application of an uncertainty profile to food microbiological methods, and provides good estimations of the uncertainty measurements for each concentration level.

Keywords: Validation, Uncertainty profile, β -content- γ -confidence tolerance interval, Uncertainty measurement, Microbiological method

© 2016 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)

INTRODUCTION

Actually, it is well known that the analytical, microbiological methods are very crucial on account of their role and efficacy to provide accurate quantification of the analyte in pharmaceutical and medical field. Concerning tests implemented, manufacturers must verify that their microbiological methods are applicable to analyze the pharmaceuticals or medical devices and then yields a confidence in the validity of analytical results that are provided to customers.

An analytical measurement is a tool to make a suitable decision. Therefore, the effort must be heading essentially to the metrological quality of the measurements, which requires introducing two important concepts: analytical validation and estimation of measurement uncertainty. Then, to ensure an analytical measurement, the analyst is called to master both these concepts which are passages mandatory for all laboratories that are trying to be accredited according to ISO 17025 [2].

Thus, the validation of a bioanalytical method is crucial since it is mandatory to prove that an alternative method is able actually to deliver results that can be interpreted in the same way as those obtained with the reference method.

Otherwise, several papers like standards and guidelines have been published in order to propose a procedure for validating analytical and bioanalytical methods [3-9]. In general, they recommend checking a validation set of criteria that must be compared to threshold values. Although these documents have largely contributed to the advancement of the implementation of analytical validation, however, they have weaknesses on the conclusions of tests performed and as to

aid in decision making related to acceptance limits defined for the use of an analytical procedure. In addition, the guide ISO 16140 [10], defined the final decision rules which are principally based on a set of null hypothesis tests, aiming to demonstrate that the estimated parameters for the alternative method do not differ significantly from those obtained with the reference method. This strategy presents many drawbacks that were extensively described in recent papers [11, 12]. Consequently, the SFSTP published a guide entitled 'Harmonization of strategies for the validation of quantitative analytical procedures [3, 4] where a new interpretation strategy has been developed, called accuracy profile, based on global acceptance criteria so that such confusing conclusions can be avoided. This strategy of accuracy profile as a decision tool based on the concept of total error and uses the β -expectation tolerance interval. It has been proved its efficacy to select the most appropriate response function, to determine the limits of quantification and to assess the concentration range. In spite of its applicability and flexibility, this approach has shown, however, gaps in the estimation of measurement uncertainty [13-21]. Undoubtedly, measurement uncertainty is a crucial parameter which the estimation is considered as a major problem for every laboratory. So as to assist analysts, standards and guides have been published to assess the measurement uncertainty [22-26] such as the International Organization for Standardization (ISO) approach [24] (commonly known as "bottom-up" approach) and the Analytical Methods Committee [27] (commonly known as "top-down" approach). From ISO Guide, EURACHEM was conceived and adapted specifically for chemical measurements [28].

The implementation of the bottom-up approach to measurement uncertainty proposed by the GUM is hardly applicable for complex

analytical or bioanalytical methods because of the difficulty of constructing an uncertainty budget model [29-31]. The propagation of the uncertainties in this model following the law of error propagation is another difficulty. According to the EURACHEM guide [28], uncertainty can be evaluated from the validation data. Thus, research studies have been published and dedicated to the one and only main purpose of simplifying and evaluating the uncertainty of measurement from validation data [32-40]. Consequently, all researchers' concentration was mainly to adopt a global, harmonic and easy to practice strategy to test the validity of an analytical method and to estimate the measurement uncertainty.

In this paper, we try to show the applicability of a new analytical validation strategy and estimation of measurement uncertainty and expand its field of application by applying it to a biological matrix. This approach called the uncertainty profile was developed in our laboratory not simply to validate an analytical method but also to estimate the measurement uncertainty. This means that thanks to this approach, we can perform a full validation.

It is based on the use the two sided β -content, γ -confidence tolerance interval in order to build a powerful graphical tool that allows simply making a decision about the validity of a bioanalytical method [14-21].

The present study aims to illustrate the validity of an alternative method for the quantitative enumeration of *Enterobacteriaceae*.

Three Chemometrics methodologies are uncovered to build the β -content, γ -confidence tolerance interval, namely: the Satterthwaite's approximation, the GPQ method (generalized pivotal confidence) and the MLS procedure (modified large simple). Furthermore, we illustrate the applicability and flexibility of the uncertainty profile to assess the fitness of the purpose of analytical, microbiological methods [20].

METHODS AND EXPERIENCES

Case study

The 3M Petri film *Enterobacteriaceae* Count Plate is a sample-ready culture medium system applicable to the enumeration of *Enterobacteriaceae* in food products and the industrial environment. The measure and is colony-forming unit per gram (CFU/g). Indeed, two major features are met in the original application: the difficulty encountered in defining the measure and for microbiological methods; and the use of experimental data collected in the framework of a collaborative study instead of by a single laboratory. Actually, tests for *Enterobacteriaceae* are mandatory steps in the process hygiene for many foodstuffs [1]. Therefore, enumeration of *Enterobacteriaceae* in foods now shows evidence of increasing interest.

Uncertainty assessment: new approach

The uncertainty is a parameter associated with the result of a measurement that characterizes the dispersion of the values that could reasonably be attributed to the measure and.

A simple and efficient procedure has recently made available to researchers to estimate the uncertainty of their analytical methods without restoring to other studies or experiments. Indeed, this approach is based only on the validation data and brings together two major objectives, the validation of analytical methods and estimation of measurement uncertainty. The estimation of the uncertainty is crucial for the interpretation of the results obtained and especially their comparisons with the statutorily above compliance limits. The declaration of an analytical result is incomplete without knowing the measurement uncertainty. Hence, ISO 17025 requires since 2001 for testing laboratories, to apply procedures for estimating uncertainty and to be able, if necessary, to be associated with the results returned [2]. Generally, the uncertainty is, as it is indicated in references [16, 17, 20], expressed by the following equations:

$$U = \bar{Y} + t(v)u(Z) \text{ EQ. 1}$$

$$L = \bar{Y} - t(v)u(Z) \text{ EQ. 2}$$

We conclude the expression of the uncertainty:

$$u(Z) = \frac{U - L}{2t(v)} \text{ EQ. 3}$$

With:

- U: is the upper β -content tolerance interval;
- L: is the lower β -content tolerance interval;
- $t(v)$ is the $\frac{1+\gamma}{2}$ quantile of Student t distribution with v degrees of freedom. For balanced data, v can be estimated by the Satterthwaite's formula [41].

Construction of uncertainty profile

The building strategy of uncertainty profile, which comes after computing the uncertainty through the equation (3), is based on the following expression:

$$|\bar{Y} \pm ku(Z)| \leq \lambda \text{ EQ. 4}$$

Where

- k is a coverage factor which depends to the level of confidence desired. For an approximate level of confidence of 95%, $k=2$.
- \bar{Y} is the mean of results and λ is the acceptance limits.

We can also build the uncertainty profile by the following expression:

$$|\text{bias} \pm ku(Z)| \leq \lambda \text{ EQ. 5}$$

Where:

$$\text{bias} = \bar{Y} - \mu_T \text{ EQ. 6}$$

And μ_T is the true value

To achieve the uncertainty profile, we have to respect the following steps [14-21]

- i. Choice of the appropriate acceptance limits taking into account the intended use of the method.
- ii. Generate all possible calibration models using the calibration data.
- iii. Calculation of the inverse predicted concentrations of all validation standards according to the selected calibration model.
- iv. Compute the two-sided β -content γ -confidence tolerance intervals for each level, according to one of three approaches proposed below.
- v. Determination of the uncertainty for each level using equation (3).
- vi. Construct the uncertainty profile according to equation (5) and make 2D-graphical representation results for the acceptability and uncertainty limits.
- vii. Compare the interval of uncertainty (L, U) to the acceptance limits $(-\lambda, \lambda)$.
- viii. If (L, U) falls totally within $(-\lambda, \lambda)$, the method is accepted; otherwise, the method is not valid.

Tolerance intervals

Before computing the uncertainty, we have to estimate the tolerance interval. Generally, two kinds of tolerance intervals are largely used, which are the β -expectation tolerance interval and β -content, γ -confidence tolerance interval. In this section, our main interest lies in β -content, γ -confidence tolerance intervals which are also called guaranteed coverage tolerance intervals [20].

In the following, we will detail three approaches to the construction of a two-sided β -content, γ -confidence tolerance interval using Mee's approach [41], Hoffman-Kringle approach [42] and Liao-Lin-Iyer approach [43, 44].

Mee's approach

In this approach, the estimation of Chi-square is based on the Satterthwaite's approximation.

The computation of uncertainty profile is based on the β - γ -content tolerance interval [41]. This later is calculated according to Mee's approach and can be expressed as [16, 17, 20]:

$$\bar{Y} \pm K_s \hat{\sigma}_{IP} \text{ EQ. 7}$$

Where:

$$K_s = \sqrt{\frac{v^* \chi_{1;\beta}^2(h)}{\chi_{v^*;1-\gamma}^2}} \text{ EQ. 8}$$

K_s is the estimation of tolerance factor which should for a bilateral β - γ -content tolerance interval in a balanced one-way ANOVA.

$$\hat{\sigma}_{IP}^2 = \hat{\sigma}_e^2 + \hat{\sigma}_b^2 \text{ EQ. 9}$$

And:

$\hat{\sigma}_b^2$: is the variance between laboratories.

$\hat{\sigma}_e^2$: is the variance within laboratories.

Liao-Lin-Iyer approach

Liao and al proposed an approach that focuses on the β -content tolerance interval. Indeed, have used the generalized pivotal confidence interval and Monte Carlo simulation to estimate the β -content tolerance intervals [43, 44].

We consider the case of one-way random model with balanced data; Note that, in order to derive a two-sided tolerance interval, we have to obtain a margin of statistic error D, a function of σ_b^2 and σ_e^2 , so that:

$$P_{\bar{Y}, \hat{\sigma}_e^2, \hat{\sigma}_b^2} \{P_Y[\bar{Y} - D < Y < \bar{Y} + D | \bar{Y}, \hat{\sigma}_{IP}] \geq \beta\} = \gamma \text{ EQ. 10}$$

Where $Y_{ij} \sim N(\mu, \sigma_b^2 + \sigma_e^2)$. Once D is obtained, the tolerance interval is given by:

$$\bar{Y} \pm D \text{ EQ. 11}$$

We define

$$\sigma_1^2 = \sigma_b^2 + \sigma_e^2 \text{ And } \sigma_2^2 = \frac{n\sigma_b^2 + \sigma_e^2}{an}$$

And

$$\sigma_1^2 + \sigma_2^2 = \left(1 + \frac{1}{a}\right) \frac{n\sigma_b^2 + \sigma_e^2}{n} \left(1 - \frac{1}{n}\right) \sigma_e^2 \text{ EQ. 12}$$

To calculate D, we must determine γ upper confidence limit for $\sigma_1^2 + \sigma_2^2$ using the generalized confidence interval idea.

Once the data are obtained, compute the observed values \bar{Y} , SS_b and SS_e . Let M denote the number of simulation runs. For $i=1, 2, 3, \dots, M$. Then, Generate independent random variables $A_{b,i} \sim \chi_{a-1}^2$ and $A_{e,i} \sim \chi_{a(n-1)}^2$.

Finally compute:

$$D_{1,i} = \frac{\left(\frac{1}{n}\right) \left(1 - \frac{1}{a}\right) SS_b}{A_{b,i}^2} + \frac{\left(1 - \frac{1}{n}\right) SS_e}{A_{e,i}^2} \text{ EQ. 13}$$

The γ quantile of the $D_{1,i}$ values gives an estimate of the γ upper confidence limit.

The square root of this upper confidence limit, multiplied by $Z_{(1+\beta)/2}$ provides a margin of error statistic D needed for computing the two-sided β -content tolerance interval. In the above, $Z_{(1+\beta)/2}$ denotes the $(1+\beta)/2$ quantile of a standard normal distribution.

Hoffman-Kringle approach

It's a strategy based on the Modified Large Simple (MLS) procedure to compute the two-sided b-content tolerance interval in balanced and unbalanced one-way random models [42]. For a balanced one-way random model, we define:

$$\sigma_1^2 = \sigma_b^2 + \sigma_e^2 \text{ And } \sigma_2^2 = \frac{n\sigma_b^2 + \sigma_e^2}{an}$$

The MLS upper confidence limit for $\sigma_b^2 + \sigma_e^2$ is given by:

$$S = \left[\left(1 + \frac{1}{a}\right) \frac{n\sigma_b^2 + \sigma_e^2}{n} + \left(1 - \frac{1}{n}\right) \sigma_e^2 \right] + \left[\left(1 + \frac{1}{a}\right)^2 \frac{(n\sigma_b^2 + \sigma_e^2)^2}{n^2} \left(\frac{a-1}{\chi_{a-1;1-\gamma^2}} - 1\right)^2 + \left(1 - \frac{1}{n}\right)^2 \sigma_e^4 \left(\frac{a(n-1)}{\chi_{a(n-1);1-\gamma^2}} - 1\right)^2 \right]^{0.5} \text{ EQ. 14}$$

Once the S is obtained, the β -content, γ -confidence tolerance interval is given by:

$$\bar{Y} \pm Z_{\frac{(1+\beta)}{2}} \sqrt{S} \text{ EQ. 15}$$

Evaluation of the enumeration method

In many classical guides, validation is habitually limited to calculate a set of performance criteria. But according to the definition of validation proposed by ISO, it must also contain a demonstration of the fitness of the method to a defined goal based on these performance criteria; therefore, uncertainty profile is a possible solution. It consists of a graphical decision-making tool to help the analyst decide whether an analytical procedure is valid. It is obtained by simultaneously combining β -content, γ -confidence tolerance interval and acceptability limits in graphical format.

In order To demonstrate the applicability of the uncertainty profile strategy to the microbiological field, we evaluate the performance of the quantitative enumeration procedure which is selected from the literature [1]. In this procedure, the dosage is direct and we haven't to calibration data, and so we aren't in need of response function or linearity study.

For validation standards, three concentration levels have involved in experiment design. Each validation sample was analyzed two times ($n= 2$) for fourteen different days ($a= 14$).

RESULTS AND DISCUSSION

At each concentration level, recovery, Bias, Standard deviation (SD) between and within laboratories and freedom degrees are calculated. All obtained results are summarized in table 1.

Table 1: Statistical results in Log₁₀ (CFU/g)

Statistic	Concentration levels		
	First	Second	Third
Concentrations	2.267	3.23	4.176
Number of days (a)	14	14	14
Number of replicates (n)	2	2	2
Bias	-0.043	0.0135	0.047
Recovery (%)	98.11	100.42	101.12
SD between laboratories	0.0308	0.0524	0.0711
SD within laboratories	0.1221	0.1100	0.1044
Freedom degree	26.74	25.75	23.98
Lower acceptability (-λ)	-0.25 Log unit	-0.25 Log unit	-0.25 Log unit
Upper acceptability (+λ)	0.25 Log unit	0.25 Log unit	0.25 Log unit

In order to calculate the tolerance intervals, we set a value of $\beta = 66.7\%$ of the content preferred to be compliant, the population of future measures included within the acceptance limits with confidence levels set at 90 % and 95 %. We expanded the value of β to 80 % and verified the validity of the method for the two risk values.

As known, the uncertainty is a key parameter in the strategy of the uncertainty profile. Its estimation is, therefore, fundamental to the

interpretation of the results and is quite logically becomes an unavoidable obligation for laboratories in the various sectors where quantitative analytical procedures are applied to provide analytical results. Indeed, since 2001, the standard ISO 17025 requires testing laboratories to apply procedures for the estimation of the uncertainty of obtained results [45, 46]. In table 2 all results about uncertainty, expanded uncertainty and uncertainty limits are presented.

Table 2: Point estimates of the uncertainty measurement and uncertainty limits obtained for each concentration level using the 66.7 and 80 %-content, 90 and 95 %-confidence tolerance interval computed by Mee, Liao-Lin-Iyer (GPQ with N=100000) and Hoffman-Kringle (MLS) methods

β -content	γ -confidence	Approach	Target values	Uncertainty (Log ₁₀ (CFU/g))	Expanded (%) Uncertainty	Lower uncertainty limit	Upper uncertainty limit
0.667	0.95	Mee	2.267	0.0810	7.1459	2.0580	2.3905
			3.23	0.0794	4.7853	3.0802	3.4069
			4.176	0.0833	3.9899	4.0510	4.3949
		GPG	2.267	0.0832	7.3416	2.0534	2.3951
			3.23	0.0809	4.8735	3.0772	3.4099
			4.176	0.0842	4.0324	4.0492	4.3967
		MLS	2.267	0.0820	7.2339	2.0559	2.3926
			3.23	0.0799	4.8103	3.0793	3.4078
			4.176	0.0833	3.9887	4.0511	4.3949
0.667	0.90	Mee	2.267	0.0909	8.0198	2.0694	2.3791
			3.23	0.0889	5.5046	3.0919	3.3952
			4.176	0.093	4.4529	4.0639	4.3820
		GPG	2.267	0.0940	8.2929	2.0641	2.3844
			3.23	0.0912	5.4914	3.0880	3.3991
			4.176	0.0947	4.5339	4.0610	4.3849
		MLS	2.267	0.0919	8.1042	2.0677	2.3808
			3.23	0.0894	5.3850	3.0910	3.3961
			4.176	0.0931	4.4608	4.0636	4.3823
0.8	0.95	Mee	2.267	0.1072	9.4583	2.0042	2.4443
			3.23	0.1051	6.3337	3.0273	3.4598
			4.176	0.1103	5.2808	3.9954	4.4505
		GPG	2.267	0.1099	9.6937	1.9987	2.4498
			3.23	0.1071	6.4500	3.0233	3.4637
			4.176	0.1112	5.3240	3.9935	4.4524
		MLS	2.267	0.1085	9.5763	2.0014	2.4471
			3.23	0.1057	6.3679	3.0261	3.4609
			4.176	0.1103	5.2802	3.9954	4.4505
0.8	0.90	Mee	2.267	0.1203	10.6150	2.0192	2.4293
			3.23	0.1176	7.0869	3.0428	3.4443
			4.176	0.1231	5.8937	4.0124	4.4335
		GPG	2.267	0.1242	10.9606	2.0126	2.4359
			3.23	0.1205	7.2616	3.0379	3.4492
			4.176	0.1255	6.0082	4.0083	4.4376
		MLS	2.267	0.1216	10.7284	2.0170	2.4315
			3.23	0.1183	7.1286	3.0416	3.4454
			4.176	0.1233	5.9052	4.0120	4.4339

Table 3: Uncertainty limits with difference to target values for two different values of β -content and γ -confidence

β -content	γ -confidence	Target values	Tolerance interval with Difference to target values		
			Mee's approach	GPQ approach	MLS approach
0.667	0.95	2.267	[-0.2090;0.1235]	[-0.2136;0.1281]	[-0.2111;0.1256]
		3.230	[-0.1498;0.1769]	[-0.1528;0.1799]	[-0.1507;0.1778]
		4.176	[-0.1250;0.2189]	[-0.1268;0.2207]	[-0.1249;0.2189]
	0.90	2.267	[-0.1976;0.1121]	[-0.2029;0.1174]	[-0.1993;0.1138]
		3.230	[-0.1381;0.1652]	[-0.1420;0.1691]	[-0.1390;0.1661]
		4.176	[-0.1121;0.2060]	[-0.1150;0.2039]	[-0.1124;0.2063]
0.8	0.95	2.267	[-0.2628;0.1773]	[-0.2683;0.1828]	[-0.2656;0.1801]
		3.230	[-0.2027;0.2298]	[-0.2067;0.2337]	[-0.2309;0.2309]
		4.176	[-0.1806;0.2745]	[-0.1825;0.2764]	[-0.1806;0.2745]
	0.90	2.267	[-0.2478;0.1623]	[-0.2544;0.1689]	[-0.2500;0.1645]
		3.230	[-0.1872;0.2143]	[-0.1921;0.2192]	[-0.1884;0.2154]
		4.176	[-0.1636;0.2575]	[-0.1677;0.2616]	[-0.1640;0.2579]

The uncertainty profiles are constructed from the results in table 3 which contain the tolerance intervals with the difference to the target levels.

The upper and lower uncertainty limits expressed in absolute value are presented in fig. below as a function of the target values generated by the reference method. As can be seen from the results,

the method was considered valid since the uncertainty intervals with the difference to target value are included in the ± 0.25 Log unit acceptance limits for all levels tested.

Legend

The continuous gray lines are the tolerance limits computed by MLS method, the dashed green lines represent the uncertainty limits computed by GPQ method, the blue dotted lines are the upper and

lower β -content, γ -confidence tolerance limits calculated by Mee's method and the red dashed lines are set at ± 0.25 Log unit.

In order to illustrate this methodology, it was applied to the validation of enumeration method of *Enterobacteriaceae* in foods. As indicated several times above, the uncertainty profile, directly reflecting the analytical procedure potential, makes possible to appreciate the adequacy of different practices and to make decisions.

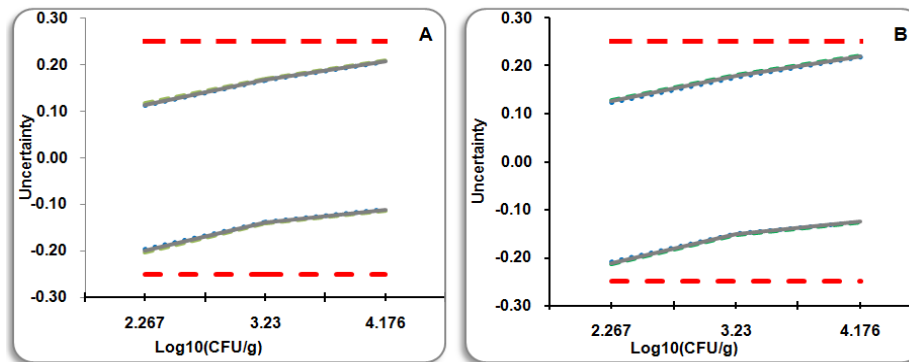


Fig. 1: Uncertainty profile of the alternative method of enumeration of *Enterobacteriaceae* using Mee, GPQ and MLS methods (A): profile carried out with $\beta=66.7\%$, $\gamma=90\%$, (B): profile carried out with $\beta=66.7\%$, $\gamma=95\%$

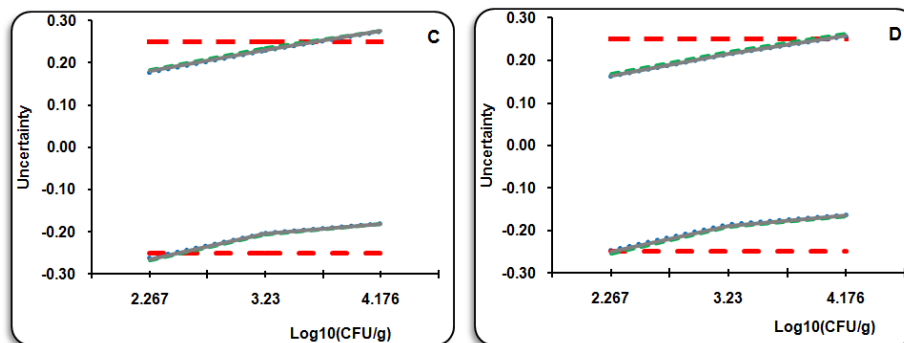


Fig. 2: Uncertainty profile of the alternative method of enumeration of *Enterobacteriaceae* using Mee, GPQ and MLS methods (C): profile carried out with $\beta=80\%$, $\gamma=90\%$, (D): profile carried out with $\beta=80\%$, $\gamma=95\%$

All the steps listed below have been applied successfully to evaluate the performance of the alternative method of enumeration. On the other hand, the proposed strategy allows a good estimate of measurement uncertainty by using validation data [1] and without having recourse to other experiments by using the formula equation (3) for estimating uncertainty based on the β -content, γ -confidence tolerance interval and computed by three different methods: the Satterthwaite's approximation (Mee's method) [41], the GPQ method (generalized pivotal confidence) [43, 44] and the MLS procedure (modified large simple).

All for $\beta=66.7\%$, $\gamma=90\%$ tolerance limit values are included within the acceptability limits, and it can be concluded that the alternative method is fully validated for all the three chemometric methods. Indeed, the tolerance intervals computed by Mee, GPQ and MLS methods are, as can be shown in fig.1. A, within the acceptance limits for all target levels. Even if we take a risk of 5%, all tolerance limits, presented in fig.1. B, are always inside the acceptance limits.

Consider that over the validation domain; the tolerance interval is included in the acceptability limits; the method can be declared as valid over this range. This can be interpreted as follows: by definition, the $\beta=66.7\%$, $\gamma=90\%$ & 95% tolerance limits is supposed to contain, on average, 66.7% of the predicted future

measurements. Therefore, the analyst can guarantee that, on average, 66.7% of the future results will fall within the limits of this prediction interval. As far as this percentage is included in the acceptability limits, the analyst can guarantee that future measurements will be equivalent to those obtained by the reference method, with an acceptance of ± 0.25 Log unit.

The other hand, we compared the results obtained from the uncertainty profile, with that calculated by the accuracy profile, each time led to the same decision even if we chose a value of ± 0.25 Log unit as acceptability limits. But, our preference is for the uncertainty interval, because this type of strategy allows a better estimate of the uncertainty measurement compared to the total error interval.

If the analyst wants more future measurements included in the limits of acceptability, there is the possibility to extend the β value to 80%. As a result, for an acceptance of ± 0.25 Log unit, the method is not valid on all range of target levels, fig. 2, for the tolerance intervals Mee, MLS, and GPG for both risks 5% and 10%.

Otherwise, if we choose an acceptance of ± 0.3 Log unit, the uncertainty limits are included in the acceptability limits; the method, as shown in fig. 3, can be declared as valid over this range.

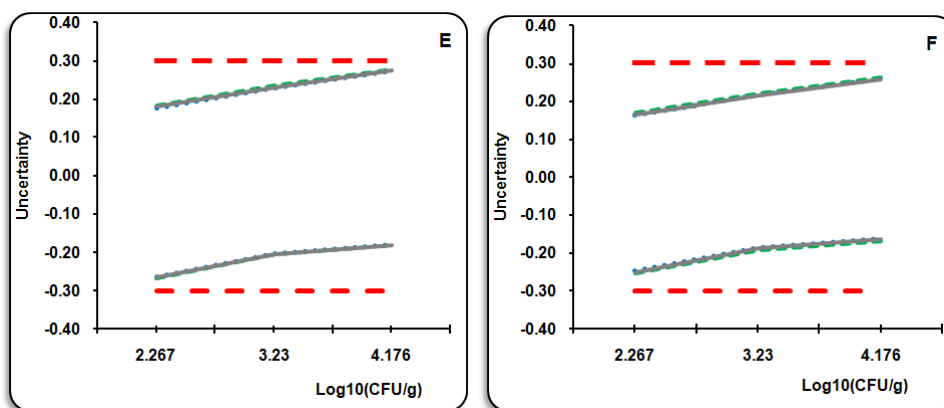


Fig. 3: Uncertainty profile of the alternative method of enumeration of *Enterobacteriaceae* using Mee, GPQ and MLS methods (C): profile carried out with $\beta=80\%$, $\gamma=90\%$, (D): profile carried out with $\beta=80\%$, $\gamma=95\%$. Acceptance limits are set at ± 0.3 Log unit

CONCLUSION

The validation of any analytical method is the ultimate step before the routine use. In this work, we have demonstrated the ability of the uncertainty profile to be used for testing the validity of enumeration method which represents the first application of an uncertainty profile to food microbiological methods, and provides good estimations of the uncertainty measurements for each concentration level.

The uncertainty profile is built from the β, γ -tolerance interval. Three chemometric methods have been used to assess the tolerance interval: the Satterthwaite's approximation (Mee), the GPQ method (generalized pivotal confidence) and the MLS procedure (modified large simple). In the light of the obtained results, we have shown that these three statistic methods can easily be applied to evaluate the performance of enumeration method as well as to assess the uncertainty of results and led to the same decision.

CONFLICT OF INTERESTS

Declared none

REFERENCES

1. Feinberg M, Sohier D, Jean-Francois D. Validation of an alternative method for counting *Enterobacteriaceae* in foods based on accuracy profile. *J AOAC Int* 2009;92:527-37.
2. ISO/DIS 17025, General requirements for the competence of calibration and testing laboratories, ISO, Geneva, Switzerland; 2000.
3. Hubert Ph, Nguyen-Huu JJ, Boulanger B, Chapuzet E, Cohen N, Compagnon PA, *et al.* Harmonization of strategies for the validation of quantitative analytical procedures-A SFSTP proposal-Part III. *J Pharm Biomed Anal* 2007;45:82-96.
4. Hubert Ph, Nguyen-Huu JJ, Boulanger B, Chapuzet E, Chiap P, Cohen N, *et al.* Harmonization of strategies for the validation of quantitative analytical procedures-A SFSTP proposal-Part II. *J Pharm Biomed Anal* 2007;45:70-81.
5. International Conference on Harmonization (ICH) of Technical Requirements for Registration of Pharmaceuticals for Human Use, Topic Q2 (R1): Validation of Analytical Procedures: Text and Methodology, Geneva; 2005.
6. Food and Drug Administration, Guidance for Industry (draft), Analytical Procedures and Methods Validation; 2000.
7. Food and Drug Administration, Guidance for Industry, Bioanalytical Methods Validation; 2001.
8. International Organization for Standardization (ISO), ISO-5725-2, Accuracy (Trueness and Precision) of Measurement Methods and Results-Part 2: Basic Method for the Determination of Repeatability and Reproducibility of a Standard Measurement Method, ISO, Geneva, Switzerland; 1994.
9. AOAC International, Method Validation Programs (OMA/PVM Department), is including Appendix D: Guidelines for Collaborative Study Procedures to Validate Characteristics of a

Method of Analysis; 2000. Available from: <http://www.aoac.org/vmeth/devmethno.htm>. [Last accessed on 20 Nov 2015].

10. ISO 16140 Microbiology of Food and Animal Feeding Stuffs-Protocol for the Validation of Alternative Methods, International Organization for Standardization, Geneva, Switzerland; 2003.
11. Feinberg M. Validation of analytical methods are based on accuracy profiles. *J Chromatogr A* 2007;1158:174-83.
12. Rozet E, Wascotte V, Lecouturier N, Pr at V, Dew  W, Boulanger B, *et al.* Improvement of the decision efficiency of the accuracy profile by means of a desirability function for analytical methods validation application to a diacetylmonoxime colorimetric assay used for the determination of urea in transdermal iontophoretic extracts. *Anal Chim Acta* 2007;591:239-47.
13. Marini RD, Chiap P, Boulanger B, Rudaz S, Rozet E, Crommen J, Hubert Ph. LC method for the determination of R-timolol in S-timolol maleate: Validation of its ability to quantify and uncertainty assessment. *Talanta* 2006;1120:322-6.
14. Jhilal F, Ihssane B, Bouchafra H, Sfaira M, Hadrami EL M, Saffaj T. Uncertainty profile: a new global strategy for the analytical validation and the estimation of measurement uncertainty. *Les Technologies De Laboratoire* 2013;7:114-20.
15. Bouchafra H, Elkarbane M, Ihssane B, Azougagh M, Jhilal F, Sosse SA, *et al.* *J Chem Pharm Res* 2014;6:2610-23.
16. Ihssane B, Sbai M, El Hadrami EM, Achour S, Saffaj T. A statistical approach based on the total error concept for validation the bioanalytical method: Application to the spectrophotometric determination of traces amount of acetaminophen in human plasma. *Int J Pharm Pharm Sci* 2014;6:413-21.
17. Saffaj T, Ihssane B. Uncertainty profiles for the validation of analytical methods. *Talanta* 2011;85:1535-42.
18. Saffaj T, Ihssane B. Response to comments on "Uncertainty profiles for the validation of analytical methods". *Talanta* 2012;94:361-72.
19. Saffaj T, Ihssane B. Remarks on "Reply to the responses to the comments on "uncertainty profiles for the validation of analytical methods" by Saffaj and Ihssane". *Talanta* 2012;106:155-67.
20. Saffaj T, Ihssane B, Jhilal F, Bouchafra H, Laslami S, Sosse SA. An overall uncertainty approaches for the validation of analytical separation methods. *Analyst* 2013;138:4677-91.
21. Saffaj T, Ihssane B. Comments on "innovative method for carbon dioxide determination in human postmortem cardiac gas samples using headspace-gas chromatography-mass spectrometry and stable labeled isotope as internal standard" by varlet *et al.* *Anal Chim Acta* 2014;810:39-42.
22. ISO/DTS 21748, Guide to the use of repeatability, reproducibility and trueness estimates in measurement uncertainty estimation, ISO, Geneva; 2004.
23. Eurachem/CITAC Guide, quantifying uncertainty in analytical measurement, Eurachem/CITAC, Budapest. 2nd edn; 2000.

- Available from: <http://www.eurachem.org/>. [Last accessed on 20 Nov 2015].
24. Guide to the Expression of Uncertainty in Measurement, ISO, Geneva, Switzerland; 1995.
 25. EA-4/16, EA Guidelines on the Expression of Uncertainty in Quantitative Testing; 2004. Available from: <http://www.europeanaccreditation.org/>. [Last accessed on 20 Nov 2015].
 26. Barwick VJ, Ellison LR, VAM Project 3.2.1, development and harmonization of measurement uncertainty principles. Part D: protocol uncertainty for evaluation from validation data; 2000.
 27. Analytical Method Committee, Uncertainty of measurement: implications of its use in analytical science. *Analyst* 1995;120:2303-8.
 28. Eurachem/CITAC Guide, quantifying uncertainty in analytical measurement, Eurachem/CITAC, Budapest. 3rd edn; 2000. Available from; <http://www.eurachem.org/>. [Last accessed on 20 Nov 2015].
 29. Horwitz W, Albert R. The concept of uncertainty as applied to chemical measurements. *Analyst* 1997;122:615-27.
 30. Visser R. Measurement uncertainty: opinions of the government, the accreditation council, and the candidate accredited laboratory. *Accredit Qual Assur* 2002;7:124-5.
 31. Visser R. Measurement uncertainty: practical problems encountered by accredited testing laboratories. *Accredit Qual Assur* 2004;9:717.
 32. Feinberg M, Boulanger B, Dewé W, Hubert Ph. New advances in method validation and measurement uncertainty aimed at improving the quality of chemical data. *Anal Bioanal Chem* 2004;380:502-14.
 33. Gonzalez AG, Herrador MA. Accuracy profiles from uncertainty measurements. *Talanta* 2006;70:896-901.
 34. Gonzalez AG, Herrador MA, Asuero AG. Practical digest for evaluating the uncertainty of analytical assays from validation data according to the LGC/VAM protocol. *Talanta* 2005;65:1022-30.
 35. Quintela M, Bagueña J, Gotor G, Blanco MJ, Broto F. Estimation of the uncertainty associated with the results based on the validation of chromatographic analysis procedures: application to the determination of chlorides by high-performance liquid chromatography and of fatty acids by high-resolution gas chromatography. *J Chromatogr A* 2012;1223:107-17.
 36. Galban J, Ubide C. Uncertainty due to the quantification step in analytical methods. *Talanta* 2007;71:1339-44.
 37. Hund E, Massart DL, Smeyers-Verbeke J. Comparison of different approaches to estimate the uncertainty of a liquid chromatographic assay. *Anal Chim Acta* 2003;480:39-52.
 38. De Beer JO, Baten P, Nsengyumva C, Smeyers-Verbeke J. Measurement uncertainty from validation and duplicate analysis results in HPLC analysis of multivitamin preparations and nutrients with different galenic forms. *J Pharm Biomed Anal* 2003;32:767-11.
 39. Dehouck P, Vander Heyden Y, Smeyers-Verbeke J, Massart DL, Marini RD, Chiap P, *et al.* "Interlaboratory study of a liquid chromatography method for erythromycin: determination of uncertainty". *J Chromatography A* 2003;1010:63-74.
 40. Gonzalez AG, Herrador MA, Tr AC, A practical guide to analytical method validation, including measurement uncertainty and accuracy profiles. *Trends Anal Chem* 2007;26:227-38.
 41. Mee R. β -expectation and β -content tolerance limits for balanced one-way ANOVA random model. *Technometrics* 1984;26:251-4.
 42. D Hoffman, R Kringle. Two-sided tolerance intervals for balanced and unbalanced random effects models. *J Biopharm Stat* 2005;15:283-93.
 43. Liao CT, Iyer HK. A tolerance interval for the normal distribution with several variance components. *Statistica Sinica* 2004;14:217-29.
 44. Liao CT, Lin TY, Iyer HK. One and two-sided tolerance intervals for general balanced mixed models and one-way unbalanced random models. *Technometrics* 2005;47:323-35.
 45. Wood R. Food analysis—the past, present, and future, valid analytical measurement bulletin. *National Measurement System* 2005;32:4–7.
 46. Van der Voet H, van Rhijn JA, Van de Wiel HJ. Inter-laboratory, time, and fitness-for-purpose aspects of effective validation, *Anal Chim Acta* 1999;391:159–71.