

**Short Communication**

**SCREENING FOR PHYTOCHEMICALS AND FTIR ANALYSIS OF *MYRISTICA DACTYLOIDES* FRUIT EXTRACTS**

**P. RAJIV\*, A. DEEPA, P. VANATHI, D. VIDHYA**

Department of Biotechnology, School of Life Sciences, Karpagam Academy of Higher Education, Eachanari post, Coimbatore 641021, Tamil Nadu, India

Email: rajivsmart15@gmail.com

Received: 02 Feb 2016 Revised and Accepted: 12 Nov 2016

**ABSTRACT**

**Objective:** The present investigation focus on screening of phytochemicals and FT-IR analysis of *Myristica dactyloides* fruit extracts. The fruit extracts were prepared using five different solvents.

**Methods:** The phytochemical analysis and FT-IR (Fourier transform infrared spectroscopy) analysis were performed using standard methods.

**Results:** The results reveals that the alkaloids, steroids, flavonoids, phenolic compounds, proteins, carbohydrates, cardio glycosides and saponins were present in methanolic extract when compared to other solvent extracts. FT-IR analysis shows the presence of different functional groups such as carboxylic acids, aromatics, alkanes, alcohols, phenols, aliphatic amines, alkenes and amine groups in the fruit extracts.

**Conclusion:** The study concluded that the methanolic extract (*M. dactyloides* fruit) has potential bioactive compounds.

**Keywords:** *Myristica dactyloides*, Methanol extract, Phytochemicals, Fourier transform infrared spectroscopy (FT-IR)

© 2017 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>) DOI: <http://dx.doi.org/10.22159/ijpps.2017v9i1.11053>

Medicinal plants are a significant part of natural wealth. They serve as vital therapeutic agents as well as valuable raw materials for manufacturing numerous traditional and modern medicines [1]. Today a number of chemicals obtained from plants are used as vital drugs in more countries in the world [2]. Secondary metabolites from plants are referred to as phytochemicals which are naturally occurring and biologically active compounds that have the potential to prevent diseases. Evaluation of the phytochemical constituents of a medicinal plant is considered to be the main step in medicinal plant research [3]. Phytochemicals with adequate antibacterial activity are reported for the treatment of bacterial infections [4]. In current years, Indian medicinal plants have been investigated by researchers for pharmacological activity. *Myristica dactyloides* belongs to *Myristicaceae* family. The fruit contains oil-cells frequently with phenolic and myristicin. *Myristica dactyloides* fruits of the plant contains many volatile oil compounds (a-pinene, camphene, b-pinene, sabinene, myrcene, a-phellandrene, a-terpinene, limonene, 1, 8-cineole, y-terpinene, linalool, terpinen-4-ol, safrole, methyl eugenol and myristicin). These oil are used to treat diseases like flatulency, diarrhea, nausea, vomiting, chronic bowel complaints, spermatorrhoea, impotency, amenorrhoea, menorrhagia, dysmenorrhoea, ulcers, splenic disorders, rheumatism, asthma, colic, flatulence and dyspepsia [5]. It has pharmacological measures such as aromatic, stimulant, sedative, antiemetic and spasmolytic. Traditionally *M. dactyloides* seed paste was used for treating dysentery [6].

As traditional medicine plays an important role in developing new plant based drugs and *M. dactyloides* is an important medicinal plant,

the present study was aimed to analyse the phytochemicals and functional groups of *M. dactyloides* fruit extracts, which will be useful for characterization purpose of various phytoconstituents.

All the chemicals and solvents were purchased from Sigma-Aldrich, India. *M. dactyloides* fruits were collected from local markets of Tirupur district, Tamil Nadu, India (11.1800 ° N, 77.2500 ° E).

10 gram of fruits was taken in clean sterile soxhlet apparatus and extracted with gradient solvent system by aqueous (100 °C), petroleum ether (40 °C), ethyl acetate (77 °C), methanol (65 °C) and ethanol (78 °C) using hot soxhlet extraction method [7]. After extraction, the extracts were dried. From that extracts were made with suitable concentrations solvents for further analysis.

The screening was performed with some modifications from the method of Harborne [8].

A small quantity fruit extracts were mixed with KBr. The functional groups were analysed using Fourier-transform infrared (FT-IR) (Shimadzu) in the region 4000–400 cm<sup>-1</sup>.

Table 1 shows the phytochemical screening of various fruit extracts. Steroids were present in all the extracts. Phenols and Flavonoids were present in ethanol and methanol extracts. Proteins were present in ethanol, methanol, ethyl acetate and aqueous extracts. Cardio glycosides were observed in all the extracts except aqueous extract. Alkaloids were present only in methanolic extracts. The similar results were obtained from *F. religiosa* and *F. bengalensis* [9].

**Table 1: Phytochemical screening of *M. dactyloides* fruit (+: present; -: absent)**

S. No.	Phytochemicals	Fruit extracts				
		Ethanol	Methanol	Petroleum Ether	Ethyl Acetate	Aqueous
1.	Alkaloids	-	+	-	-	-
2.	Flavonoids	+	+	-	-	-
3.	Steroids	+	+	+	+	+
4.	Phenol	+	+	-	-	-
5.	Proteins	+	+	-	+	+
6.	Carbohydrates	+	+	-	+	-
7.	Cardio glycosides	+	+	+	+	-
8.	Saponins	+	+	+	+	-

Fig. 1-5 shows the FTIR spectra of *M. dactyloides* fruit extracts. The peak at  $3332.99\text{ cm}^{-1}$  revealed the presence of alcohols, phenols (O-H stretch, H-bonded). The peak at  $2970.38$  and  $2885.51\text{ cm}^{-1}$  refers to the presence of alkanes (C-H stretch). The peak at  $1759.08$  and  $1666.50\text{ cm}^{-1}$  corresponds to the carboxylic acid group (C=O stretch). A peak at  $1597.06\text{ cm}^{-1}$  denotes the  $1^\circ$  amines (N-H bend). A peak of

$1327.03\text{ cm}^{-1}$  showed the presence of aromatic amines (C-N stretch). The peaks of  $1273.02$ ,  $1087.85$  and  $1049.28\text{ cm}^{-1}$  indicate the alcohols, carboxylic acids, esters, ethers (C-O stretch). A peak of  $879.54\text{ cm}^{-1}$  revealed the alkenes (=C-H bend). FTIR spectrum analysis by *Caralluma fimbriata* was reported in earlier research and found the presence of phenols, alkanes, aromatic amines [10].

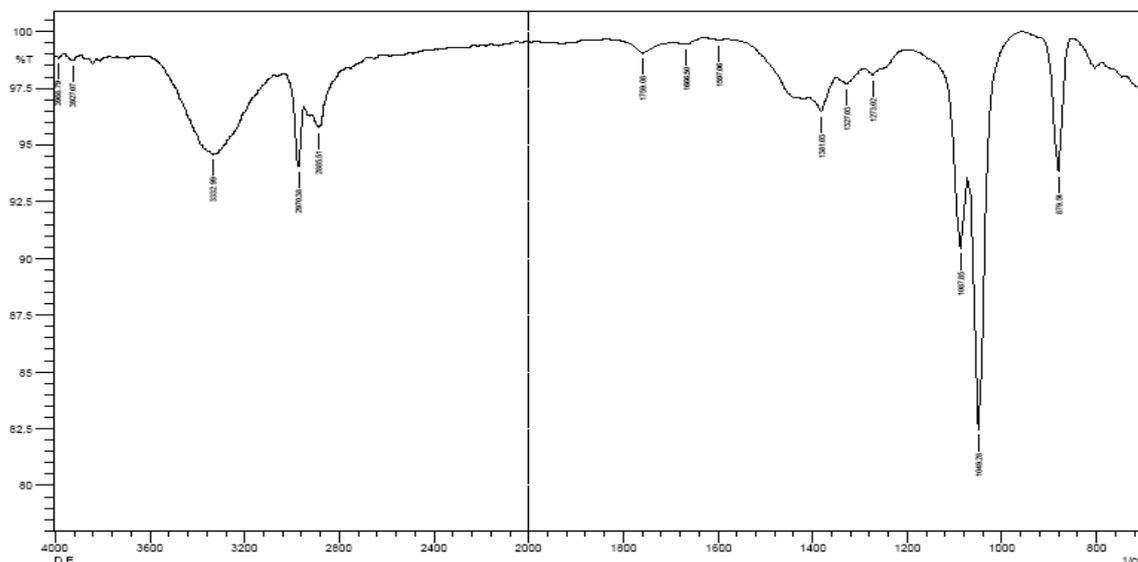


Fig. 1: FT-IR Spectra of *M. dactyloides* ethanolic fruit extract

The broad peaks at  $2831.50$ ,  $1442.75$ ,  $1111.00$  and  $1026.13\text{ cm}^{-1}$  represents the presence of functional groups such as alcohols, phenols (O-H stretch, H-bonded), carboxylic acids (O-H stretch), aromatics (C-C stretch (in-ring), aliphatic amines (C-N stretch) and alcohols, carboxylic acids, esters, ethers (C-O stretch) (fig. 2). Similar results were obtained from an ethanolic extract of

*Tylophora pauciflora* and observed the peak at  $2800$ ,  $2862.36$  and  $2926.01\text{ cm}^{-1}$  which correspond to lipids, alkanes, and hydroxyl compounds and the peak at  $1730$ , and  $1708\text{ cm}^{-1}$  shows the presence of ester carbonyl and unsaturated carbonyl groups. The strong absorption bands at  $2931$  and  $1458\text{ cm}^{-1}$  are due to CH and  $\text{CH}_2$  groups respectively [11].

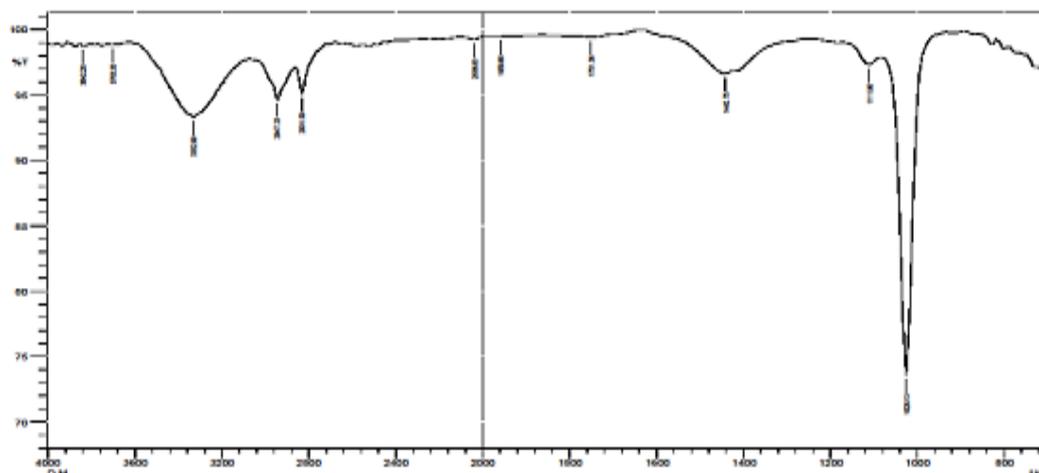


Fig. 2: FT-IR Spectra of *M. dactyloides* methanolic fruit extract

The peaks at  $3170.97$ ,  $2954.95$ ,  $2924.09$ ,  $2870.08$ ,  $1759.08$ ,  $1658.78$ ,  $1604.77$ ,  $1458.18$ ,  $1242.16$ ,  $1056.99$ ,  $910.40$ ,  $864.11$  and  $732.95\text{ cm}^{-1}$  indicates the presence of functional groups such as carboxylic acids (O-H stretch), alkanes (C-H stretch), carboxylic acids (C=O stretch), alkenes (C=C-stretch),  $1^\circ$  amines (N-H bend), aromatics (C-C stretch (in-ring), alcohols, carboxylic acids, esters, ethers (C-O stretch), carboxylic acids (O-H bend) and aromatics (C-H) (fig. 3).

Ragavendran *et al.* analysed leaf extract of *Aerva lanata* by FTIR and reported that the functional groups of halogens, amines, sulphur derivatives, polysaccharides, organic hydrocarbons and carboxylic acids are present in the extract and also reported that the strong absorption band were observed around  $3373\text{--}3422\text{ cm}^{-1}$  may be due to the presence of bonded N-H/C-H/O-H stretching of amines and amides [12].

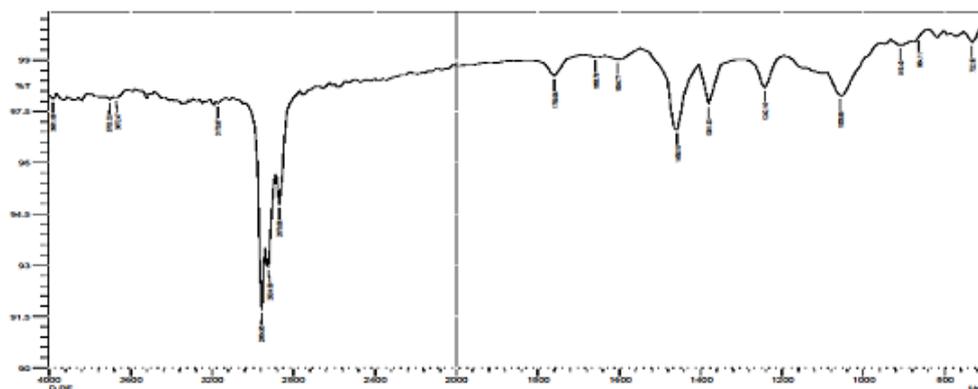


Fig. 3: FT-IR Spectra of *M. dactyloides* fruit (Petroleum ether extract)

FTIR spectroscopic analysis of ethyl acetate extract (*M. dactyloides* fruit) was shown in fig. 4. The peaks at 3155.54, 2985.81, 1743.65, 1442.75, 1242.16, 1049.28, 933.55, 848.68, 786.96 and 702.09  $\text{cm}^{-1}$  corresponds to the functional groups such as carboxylic acids (O-H stretch), alkenes (C-H stretch), carboxylic acids (C=O stretch), aromatics (C-C stretch (in-ring), alcohols, carboxylic acids, esters,

ethers (C-O stretch), aliphatic amines (C-N stretch), carboxylic acids (O-H bend) and 1°, 2° amines (N-H wag). Similar research was carried out in FTIR spectral analysis of *Ampelocissus latifolia* extract and reported that the presence of functional groups such as metal carbonyl compounds, alkanes, amides and aliphatic fluoro compounds were responsible for potential medicinal properties [13].

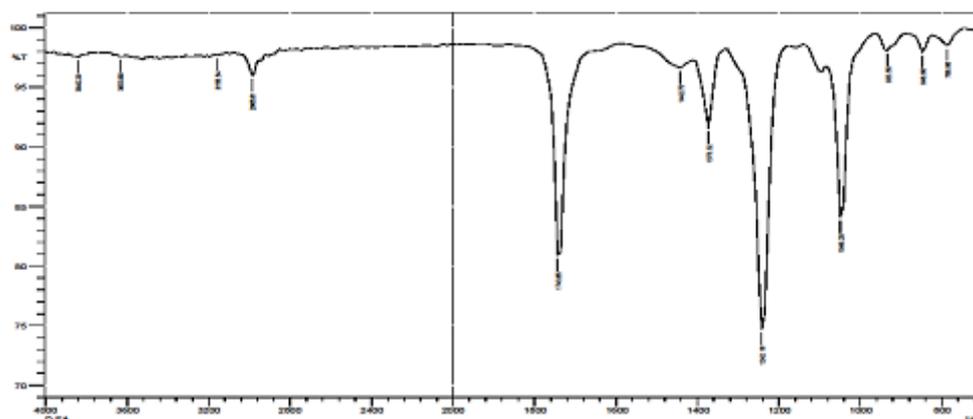


Fig. 4: FT-IR Spectra of *M. dactyloides* fruit (Ethyl acetate extract)

The peaks at 2723.49, 2245.14, 1743.65, 1689.64, 1651.07, 1527.62, 1404.18, 1273.02, 1126.43, 933.55, 833.25 and 740.67  $\text{cm}^{-1}$  refers to the presence of functional group such as aldehydes (H-C=O: C-H stretch), nitriles (C≡N stretch), carboxylic acids (C=O stretch), carbonyls (general) (C=O stretch), alkenes (-C=C-stretch), nitro compounds (N-O asymmetric stretch), aromatics (C-C stretch (in-ring)), alcohols, carboxylic acids, esters, ethers (C-O stretch),

carboxylic acids (O-H bend) and 1°, 2° amines (N-H wag). Muruganantham *et al.* analyzed the FTIR spectral analysis of medicinal plants such as *Eclipta alba* and *Eclipta prostrata* and reported that the very strong absorption band appearing in the region 2933–2922  $\text{cm}^{-1}$  for whole plant parts is due to N-H stretching and also reported the presence of functional groups like carboxylic acids, amines, polysaccharides, nitrates and carbohydrate [14].

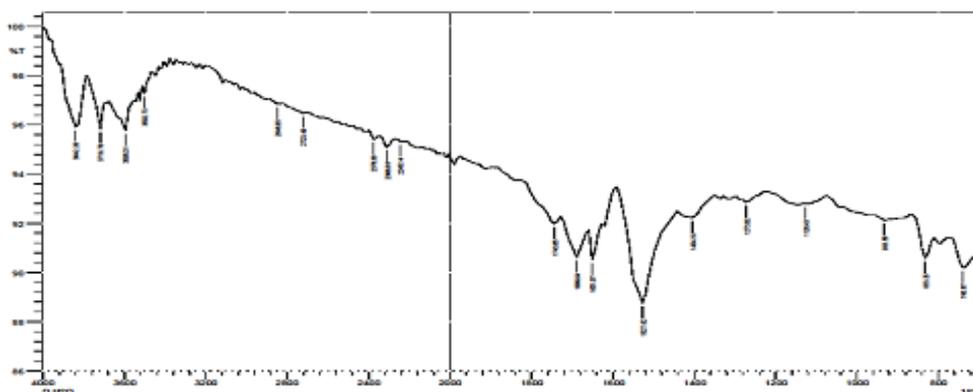


Fig. 5: FTIR analysis of *M. dactyloides* Aqueous fruit extract

The study concluded that the methanolic extract (*M. dactyloides* fruit) has potential bioactive compounds like alkaloids, glycosides, flavonoids, and tannins. FTIR spectra showed the presence of the functional group in all the extracts which have medicinal properties and can be used as antimicrobial and anticancer agents.

#### ACKNOWLEDGEMENT

We thank Management of Karpagam University, Coimbatore, Tamil Nadu, India for providing necessary facilities to carry out this work.

#### CONFLICT OF INTERESTS

Declared none

#### REFERENCES

- Nayak BS, Krishna M. Influence of ethanolic extract of *Jasminum grand forum* Linn flower on wound healing activity in rats. *Indian J Physiol Pharmacol* 2007;51:189-94.
- Debnath M, Malik CP, Bisen PS. Micropropagation: a tool for the production of high-quality plant-based medicines. *Curr Pharm Biotechnol* 2006;7:33-49.
- Banso A, Adeyemo SO. Phytochemical and antimicrobial evaluation of ethanolic extract of *Dra-caena manni*. *Bark Nig J Biotechnol* 2007;18:27-32.
- Parekh J, Chanda VS. *In vitro* antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turk J Biol* 2007;31:53-8.
- Jellin JM, Gregory PJ, Batz F, Hitchens K. Prescriber's letter natural medicines comprehensive database. 7th ed. Stockton CA: Therapeutic Research Faculty. 2005. p. 918-9.
- Vaidyanathan D, Salai Senthilkumar MS, Ghouse Basha M. Studies on ethnomedicinal plants used by Malayali tribals in Kolli hills of Eastern Ghats, Tamil Nadu, India. *Asian J Plant Sci Res* 2013;3:29-45.
- Koperuncholan M, Sathish Kumar P, Sathiyarayanan G, Vivek G. Phytochemical screening and antimicrobial studies of some ethnomedicinal plants in the south-eastern slope of western ghats. *Int J Medicobiol Res* 2010;1:48-59.
- Harborne JB. *Phytochemical methods*. 2nd ed. London (NY): Chapman and Hall publications; 1984. p. 288.
- Uma B, Prabhakar K, Rajendran S. *In vitro* antimicrobial activity and phytochemical analysis of *Ficus religiosa L.* and *Ficus bengalensis L.* against *Diarrhoeal Enterotoxigenic E. coli*. *J Ethnobot Leaflets* 2009;13:472-4.
- Packialakshmi N, Naziya S. Fourier transform infrared spectroscopy analysis of various solvent extracts of *Caralluma fimbriata*. *Asian J Biomed Pharm Sci* 2014;4:20-5.
- Starlin T, Arul Raj C, Ragavendran P, Gopalakrishnan VK. Phytochemical screening, functional groups and element analysis of *Tylophora pauciflora* weight and arn. *Int Res J Pharm* 2012;3:182-3.
- Ragavendran P, Sophia D, Arul Raj C, Gopalakrishnan VK. Functional group analysis of various extracts of *Aerva lanata (L.)* by FTIR spectrum. *Pharmacologyonline* 2011;1:358-64.
- Parag A Pednekar, Bhanu Raman. Antimicrobial and antioxidant potential with FTIR analysis of *Ampelocissus latifolia (roxb.)* Planch. Leaves. *Asian J Pharm Clin Res* 2013;6:67-73.
- Murugantham S, Anbalagan G, Ramamurthy N. FT-IR and SEM-EDS comparative analysis of medicinal plants, *Eclipta alba Hassk* and *Eclipta prostrata* Linn. *Rom J Biophys* 2009;19:285-94.

#### How to cite this article

- P Rajiv, A Deepa, P Vanathi, D Vidhya. Screening for phytochemicals and FTIR analysis of *Myristica dactyloides* fruit extracts. *Int J Pharm Pharm Sci* 2017;9(1):315-318.