

Print ISSN: 2656-0097 | Online ISSN: 0975-1491

Vol 15, Issue 1 2023

Original Article

LEFLUNOMIDE TABLET FORMULATION: DEVELOPMENT AND VALIDATION OF AN RP-HPLC TECHNIQUE

POTHURAJU NARESH*, K. VINOD KUMAR

Department of Pharmaceutical Analysis, Raghavendra Institute of Pharmaceutical Education and Research (RIPER)-Autonomous, Anantapur, Andhra Pradesh, India 515721 Email: pothurajunareshdopa18@gmail.com

Received: 03 Oct 2022, Revised and Accepted: 11 Nov 2022

ABSTRACT

Objective: To develop a Reverse Phase-High Performance Liquid Chromatography (RP-HPLC) method for Leflunomide using rapid, cheap, economical, and less composition of the mobile phase. To validate the method's specificity, linearity, precision, accuracy, robustness, and ruggedness were all validated as per regulatory requirements ICH Q_2 [R1] guidelines.

Methods: The method employed solving of Development and validation based on the measurement of absorbance at one wavelength, 251 nm, λ max, Inertsil-ODS C18 analytical column (250 x 4.6 mm, 5 μ). 1.0 ml/min of a mobile phase consisting of water and methanol (40:60v/v) for Leflunomide tablet formulation.

Results: The method showed excellent linear response with correlation coefficient (R^2) values of 0.999 for a Leflunomide. The percent recoveries for a drug were found within the acceptance limit of (99.93%–100.34%). Intra-and inter-day precision studies of the new method were less than the maximum allowable limit percentage of relative standard deviation (%RSD) 2.0. It can be concluded from the results that the present method for validation determination of Leflunomide in tablets is specific, rapid, and simple with good sensitivity.

Conclusion: This analytical method is also applicable in ordinary laboratories and also technique may be used to measure the drug and assess the uniformity and purity of the dosage formulation as well as for quality control of commercial Leflunomide tablets.

Keywords: HPLC, Leflunomide, Validation, Tablet

© 2023 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (https://creativecommons.org/licenses/by/4.0/) DOI: https://dx.doi.org/10.22159/ijpps.2023v15i1.46505. Journal homepage: https://innovareacademics.in/journals/index.php/ijpps.

INTRODUCTION

[N-[4',5'-trifluoromethylphenyl]-5-methylisoxazole-4-Leflunomide carboxamide] is an isoxazole derivative used as an anti-rheumatic drug with a molecular weight of 270.2 (fig. 1) [1]. The mechanism of action is selective inhibition of dihydro-orotate dehydrogenase [2], a crucial enzyme in the denovo synthesis of pyrimidine, and the subsequent suppression of Ribosenucleic acid and Deoxyribonucleic acid synthesis [3]. Leflunomide may be particularly toxic to activated T cells, which mainly generate pyrimidines through the de novo route [4]. Blockade of tumor necrosis factor is one of the leflunomide's immunomodulatory and anti-inflammatory actions, which have recently been reviewed [5]. Reactive oxygen radicals [6] are inhibited by mediated activation of the transcription factor NFêB. Increases in tissue inhibitor of metalloproteinase (matrix metalloproteinases) ratios as a result of polymorphonuclear leucocyte movement into the rheumatoid synovial cavity, suppression of matrix metalloproteinases, and patients with its metabolism [7]. Leflunomide in plasma was determined using LC-MS, HPLC, etc. It has been reported in recent research describes the pharmaceutical determination of leflunomide by FIA-UV. Leflunomide in pharmaceutical formulations may be regularly checked for quality using the approach described in this study, which is quick and sensitive and uses UV detection [8] Linearity, accuracy, precision, and robustness were the criteria used to validate the approach. The robustness and intermediate accuracy of the experimental design were validated [9].

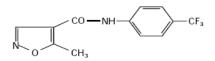


Fig. 1: Structure of leflunomide

Experimental design

Apparatus

HPLC system

Chromatographic separation was achieved using the RP-HPLC Waters system, which includes the Waters Model No. 2690/5 UV-Visible detector, Waters Pump Control Module-II, Waters 515 Solvent Delivery System [10] (pump), Rheodyne-injector (20 μ loop), and Waters Empower-2 software from the Waters Corporation as the data processor. Column for analysis Inertsil-ODS C18 (250 x 4.6 mm, 5 μ). Spincotech Pvt Ltd's Sonicaultra-sonic cleaner was used to degas the mobile phase after it had been passed through a 0.45 μ m membrane filter.

Reagents

Methanol and Acetonitrile of HPLC grade (A. R. grade) were provided from (MerckIndia). Sun Pharmaceuticals Ltd. Baroda, India provided a pure sample of the medication and an internal standard [11]. All solutions for the procedure were made with ultra-pure water made with a Milli-Q® UF-Plus device (Millipore) [12]. Leflunomide in conventional formulations was determined using [lefra®] 20 mg tablets.

Chromatographic condition

By completing several trials using various mobile phases and altering their compositions and flow rates, the chromatographic conditions were eventually optimized, resulting in the development of an optimized chromatogram [13] [table 6].

Preparation of mobile phase

Using a vacuum filtration method, the mobile phases, water and methanol were taken in a 40:60 ratio v/v. Furthermore, the mobile phases were filtered through a membrane filter and sonicated for 15

min in an ultrasonic water bath. (Millipore nylon disc filter 0.45 μ m) [14] Before usage [15].

Standard stock and standards solution preparation

10 mg of Leflunomide was accurately weighed and added into a 10 ml volumetric flask, first dissolving it in a required amount of methanol, followed by sonication for 10 min [16]. Methanol should be added after the solution has been made up, obtaining a concentration of 1000, 100, 10 mcg/ml [17].

Extraction of leflunomide from tablets

20 Leflunomide containing lefra® tablets, equivalent to 20 mg, were weighed and added to a 100 ml volumetric flask [1]. 90 ml of methanol was added, and the mixture was centrifuged at 1000 rpm for 30 min after being steeped in an ultrasonication bath for 10 min. To volume, the supernatant was diluted with the same solvent. Furthermore, a 0.45 μ m filter was used to filter the solution, and the filtrate was used to make sample solutions in various concentrations [18].

Preparation of calibration curve standards

The concentration range for the calibration curve was $20-70 \mu cg/ml$, and the required amount of mobile phase was added. Through the use of 0.45 μ m membrane filter paper, the formed solutions were filtered, and the filtrate was used for analysis [19].

Optimized method development and validation

We have created a quick and accurate RP-HPLC technique for extract sample quantification for this investigation. ODS C18 Column Inertsil [250 x 4.6 mm, 5 μ] [20]. Water and methanol were utilized as the mobile phase and column in a ratio of 40:60 v/v. 1.0 ml/min flow rate. The 256 nm wavelength was used for the detecting process.

The developed technique the created procedure was precise and accurate [21] [fig. 3].

RESULTS AND DISCUSSION

Leflunomide 10μ cg/ml and the internal standard Reslizumab 10μ cg/ml could be separated well using the chromatographic conditions that were used [fig. 2]. No drug deterioration was detected during the analysis. The following measures were used to validate the Liquid chromatography technique [9].

Validation

The optimized chromatographic method was completely validated to the procedures in ICH guidelines validation of analytical methods ICH $Q_{2 [R1]}$ [22].

Linearity: [n=6]

By using the mobile phase, chromatography was performed on leflunomide and an internal standard [23]. Leflunomide was used to examine the linearity of peak area responses to concentrations from 20 to 70μ cg/ml. Over the studied concentration range, a linear response was seen. The results are tabulated in [table 3].

Accuracy and precision

Leflunomide concentrations of 20, 40, and 60 μ cg were included in three separate solutions used to determine accuracy [24]. The obtained values were within the range of 99.93%100.24%, 100.34% mean (Relative Standard Deviation) RSD% was 0.00431, satisfying the conditions for the study's acceptance. The reproducibility was determined with five injections of Leflunomide with an analytical concentration of approximately 40 μ cg [25]. The RSD% was 0.00352 and 0.00373, respectively [table 2].

Table 1: Data on the accurac	v of leflunomide [n=	31 where "n"=three	different concentration
Table 1. Data on the accurac	y of ichunonnuc m-	J where n -unce	unici chi concenti ation

S. No.	Amount added mcg/ml	Recovery level	Amount recovered mcg/ml	% Recovery [n=3]	%RSD
1	20	50%	19.98	99.93%	0.00342
2	40	100%	40.09	100.24%	0.00431
3	60	150%	60.20	100.34%	0.00145

Values are presented in the form of mean±RSD

Table 2: Leflunomide repeatability data

S. No.	Concentration µg/ml	Intraday [n=3]			Interday [n=3		
1	40 μcg/ml	Ι	II	III	Ι	II	III
		3058687.12	3058588.92	3058782.28	3058349.65	3058594.07	3058580.67
Mean		3058632.20			3058537.12		
SD		107.9303			114.2974		
%RSD		0.00352			0.00373		

n=peak area of three determination, Values are presented in the form of mean±SD

Specificity

In the drug's High-performance liquid chromatography, the chromatograms showed relatively no peaks within a 6 min retention time range. No interference was observed from the

additives and by-products. This method was found to be specific [26]. The stability of the stock solution was evaluated under two conditions, at room temperature, stored in the refrigerator (2-8 °C). Consequently, it was determined that the peak is peculiar for this particular Leflunomide [27].

Table 3: Data on the linearity of Leflunomide

A statistical attribute	HPLC
Concentration range (mcg/ml)	20-70
Regression equation	y = 76034x+9579.
Correlation coefficient (r)	$R^2 = 0.9999$
Slope	76034
y-Intercept	9579
Limit of detection [LOD] (µg/ml)	0.0041
Limit of quantification [LOQ] (µg/ml)	0.0126

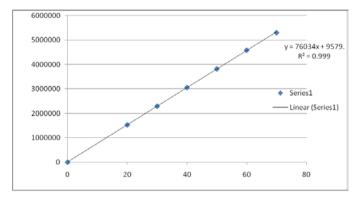


Fig. 2: Plot of linearity (Concentration Vs Peak area) [n=6], n=peak area of six determination, % RSD; Percentage relative standard deviation

Robustness

According to the International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use(ICH), an analytical procedure's robustness is its capacity to be unaffected by minor and intentional changes to the method's parameters [ICH, 1997] [28]. In robustness testing, a multivariate strategy incorporating the design of experiments is advised to explore the simultaneous change of the variables on the taken-in responses. According to the system appropriateness criteria, theoretical plates and asymmetry were determined to be in excellent condition [29]. Thus, the investigation supports the validity of the test technique for detecting even little chromatographic condition changes. Thus, the approach may be described as robust [30].

	Table 4	4: Leflunomid	e robustness stud	dy, SD; Standard deviation
--	---------	---------------	-------------------	----------------------------

Flow rate	STD. peak mean	Tailing factor mean	SD	%RSD	
0.8 ml	1924774.87	1.112	87.2224	0.00453	
1.0 ml	3058552.29	1.113	127.5140	0.00416	
1.2 ml	4132485.35	1.115	134.8511	0.00326	

Values are presented in the form of mean±SD

Parameter	Criteria	Formula	Results	
LOD	S/N = 3	3.3 x S. D/Slope	0.0041µg/ml	
LOQ	S/N =10	10 x S. D/Slope	0.0126µg/ml	

Table 5: Limit of quantification and LIMIT of detection study of leflunomide

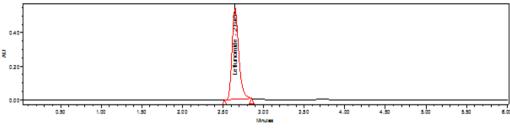


Fig. 3: The leflunomide chromatogram [25 mg]

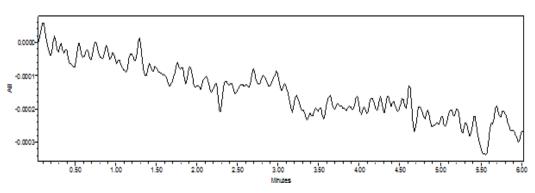
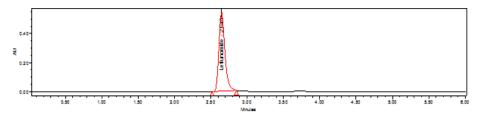
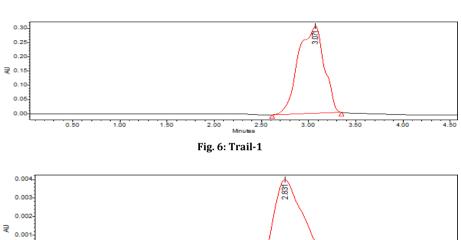


Fig. 4: The leflunomide blank chromatogram

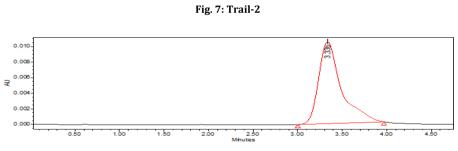
Table 6: Separations trails with different mobile phase compositions on C18 column (ODS)

Trial No	Mobile phase composition	Flow rate	t _R	Remarks
1	Acetonitrile: Water [90:10 v/v]	1.0 ml/min	3.071	A bold Peak was observed
2	Acetonitrile: methanol [45:55 V/V.]	1.0 ml/min	2.831	Got noise baseline, peak shape was not good
3	Acetonitrile: Methanol [50:50 V/V.]	1.0 ml/min	3.336	Peak Tailing was observed.
4	Methanol: Water [60:40 V/V]	1.0 ml/min	2.650	The sharp chromatogram and peak shape were good.









2.50 Minutes

2.00

1.50

3.00

3.50

4.00

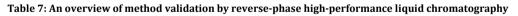
4.50

0.000

0.50

1.00

Fig. 8: Trail-3



Parameters	Leflunomide
Specificity	Peak Purity-No interference (by Uv-PDA detector)
Linearity and Range	20-70 mcg/ml
Regression equation	y = 76034x + 9579
Correlation coefficient	0.999
Accuracy-50%	99.93%
100%	100.24%
150%	100.34%
Precision-Intraday	0.00352
Inter day	0.00373
Repeatability	0.00362
LOD	0.0041µg/ml
LOQ	0.0126µg/ml
Robustness	The system suitability parameters were determined to be well within the accepted standards, so that method
	should be robust

An overview of the conditions for method validation

The whole list of unique validation parameters produced by the Reverse phase-High-performance liquid chromatography technique for the Leflunomide tablet.

CONCLUSION

The suggested high-performance liquid chromatographic technique was assessed for linearity, precision, accuracy, and suitability; it was found to be practical and successful for the quality control of Leflunomide in dosing types for pharmaceuticals. With a correlation value of 0.999, it was demonstrated that the measured signal was exact, accurate, and linear across the concentration range examined (20-70 mcg). Additionally, the chromatographic process is economical and ecologically beneficial due to the minimum solvent consumption and the brief analytical run time of 6.0 min. It is clear from the findings that the suggested approach may be used to determine Leflunomide with reliable sensitivity and without causing any interference. As a result, the proposed methodology is quick, selective, and only needs a quick sample preparation step, and provides a good method for making tablets with Leflunomide.

ACKNOWLEDGMENT

I express my special thanks to S. MahaboobBasha and B. Ravikumar PG lab assistant, Instrumentation lab for their continuous assistance throughout my work. It is my privilege to utilize the moment and bow myself to acknowledge my family and my friends.

FUNDING

Nil

AUTHORS CONTRIBUTIONS

The investigation, original draft, writing, formal analysis, data curation, validation, editing: Pothuraju Naresh, Conceptualization, resources, supervision: Dr. K. Vinod Kumar, All data were generated in-house, and no paper mill was used. All authors agree to be accountable for all aspects of this work, ensuring integrity and accuracy. All authors have read and agreed to the published version of the manuscript.

CONFLICT OF INTERESTS

No conflicts of interest are reported by the authors.

REFERENCES

- Srinivas RV, Narasimha RM, Allam AR, Maheswari I, Srinubabu G. Development and validation of LC method for the determination of leflunomide in pharmaceutical formulations using an experimental design. Afr J Pure Appl Chem 2008;2(2):10-7.
- Rastogi RP, Mehrotra BN. Compendium of Indian medicinal plants. Vol. 2. Lucknow and New Delhi: Central Drug Research Institute and National Institute of Science Communication; 1980. p. 473-5.
- Schiff MH, Whelton A. editors. Renal toxicity associated with disease-modifying antirheumatic drugs used for the treatment of rheumatoid arthritis. Semin Arthritis Rheum. 2000;30(3):196-208. doi: 10.1053/sarh.2000.16641, PMID 11124283.
- Herrmann ML, Schleyerbach R, Kirschbaum BJ. Leflunomide: an immunomodulatory drug for the treatment of rheumatoid arthritis and other autoimmune diseases. Immunopharmacology. 2000;47(2-3):273-89. doi: 10.1016/s0162-3109(00)00191-0, PMID 10878294.
- 5. Viviano KR. Pharmacotherapeutics of immune-mediated disease. Pharmacother Vet Dispensing. 2019:339-60.
- Gupta S, George M, Singhal M, Sharma GN, Garg V. Leaves extract of Murraya koenigii Linn for anti-inflammatory and analgesic activity in animal models. J Adv Pharm Technol Res. 2010;1(1):68-77. PMID 22247833.
- Stomrud E, Bjorkqvist M, Janciauskiene S, Minthon L, Hansson O. Alterations of matrix metalloproteinases in the healthy elderly with increased risk of prodromal Alzheimer's disease. Alzheimers Res Ther. 2010;2(3):20. doi: 10.1186/alzrt44, PMID 20576109.

- Nussbaumer S, Bonnabry P, Veuthey JL, Fleury Souverain S. Analysis of anticancer drugs: a review. Talanta. 2011;85(5):2265-89. doi: 10.1016/j.talanta.2011.08.034, PMID 21962644.
- Srinubabu G, Raju ChA, Sarath N, Kumar PK, Rao JV. Development and validation of a HPLC method for the determination of voriconazole in pharmaceutical formulation using an experimental design. Talanta. 2007;71(3):1424-9. doi: 10.1016/j.talanta.2006.04.042, PMID 19071468.
- 10. Gokul P, Ravichandran S. Bioanalytical method development and validation for simultaneous estimation of lercanidipine and atenolol in human plasma by using RP-HPLC; 2017.
- Gannu R, Yamsani VV, Rao YM. New RP-HPLC method with UV-detection for the determination of carvedilol in human serum. J Liq Chromatogr Relat Technol. 2007;30(11):1677-85. doi: 10.1080/10826070701224937.
- Mendez AS, Steppe M, Schapoval EE. Validation of HPLC and UV spectrophotometric methods for the determination of meropenem in the pharmaceutical dosage form. J Pharm Biomed Anal. 2003;33(5):947-54. doi: 10.1016/s0731-7085(03)00366-2, PMID 14656585.
- Guichard N, Fekete S, Guillarme D, Bonnabry P, Fleury Souverain S. Computer-assisted UHPLC-MS method development and optimization for the determination of 24 antineoplastic drugs used in hospital pharmacy. J Pharm Biomed Anal. 2019;164:395-401. doi: 10.1016/j.jpba.2018.11.014, PMID 30439666.
- 14. Sebesta DW. Glycosidase induction in Pseudomonas stutzeri and properties of one of its amylases maltotetraohydrolase: University of London. Royal Holloway and United Kingdom: Bedford New College; 1987.
- Sánchez Brunete C, Rodriguez A, Tadeo JL. Multiresidue analysis of carbamate pesticides in soil by sonication-assisted extraction in small columns and liquid chromatography. J Chromatogr A. 2003;1007(1-2):85-91. doi: 10.1016/s0021-9673(03)00953-1, PMID 12924554.
- Ahmad S, Ahire S, Tamiya A. QbD approach method development and validation for the simultaneous estimation of methotrexate and folic acid by UV spectrophotometric and RP-HPLC in bulk and tablet dosage forms. Journal of Pharmaceutical Sciences and Research. 2021;13(9):553-8.
- 17. Sakhare R. Development and validation of a stability indicating RP-HPLC method for the determination of adefovir dipivoxil in bulk and tablet dosage form. Int J Pharm Res. 2015;7(4):25.
- Horowitz AJ, Elrick KA, Colberg MR. The effect of membrane filtration artifacts on dissolved trace element concentrations. Water Res. 1992;26(6):753-63. doi: 10.1016/0043-1354(92)90006-P.
- Kennedy VC, Zellweger GW, Jones BF. Filter pore-size effects on the analysis of Al, Fe, Mn, and Ti in water. Water Resour Res. 1974;10(4):785-90. doi: 10.1029/WR010i004p00785.
- Zhou G, Chen Y, Tang Y. Total content of piperidine analysis in artane by RP-HPLC using pre-column derivatization with 4toluene sulfonyl chloride. J Chromatogr Sci. 2022;60(7):613-9. doi: 10.1093/chromsci/bmab099, PMID 34343261.
- Zhang A, Wan L, Wu C, Fang Y, Han G, Li H. Simultaneous determination of 14 phenolic compounds in grape canes by HPLC-DAD-UV using wavelength switching detection. Molecules. 2013;18(11):14241-57. doi: 10.3390/molecules181114241, PMID 24252994.
- 22. Borman P, Elder D. Q2 [R1] validation of analytical procedures. ICH Quality Guidelines. 2017;5:127-66.
- Schmidt A, Schwind B, Gillich M, Brune K, Hinz B. Simultaneous determination of leflunomide and its active metabolite, A77 1726, in human plasma by high-performance liquid chromatography. Biomed Chromatogr. 2003;17(4):276-81. doi: 10.1002/bmc.244, PMID 12833393.
- Shamonki MI, Spandorfer SD, Rosenwaks Z. Ultrasound-guided embryo transfer and the accuracy of trial embryo transfer. Hum Reprod. 2005;20(3):709-16. doi: 10.1093/humrep/deh546, PMID 15689350.
- 25. Chan V, Charles BG, Tett SE. Rapid determination of the active leflunomide metabolite A77 1726 in human plasma by high-performance liquid chromatography. J Chromatogr B Analyt

Technol Biomed Life Sci. 2004;803(2):331-5. doi: 10.1016/j.jchromb.2004.01.016, PMID 15063344.

- Abdelwahab NS, Abdelrahman MM, Boshra JM, Taha AA. Different stability-indicating chromatographic methods for specific determination of paracetamol, dantrolene sodium, their toxic impurities and degradation products. Biomed Chromatogr. 2019;33(9):e4598. doi: 10.1002/bmc.4598, PMID 31108565.
- Alt FW, Kellems RE, Bertino JR, Schimke RT. Selective multiplication of dihydrofolate reductase genes in methotrexate-resistant variants of cultured murine cells. J Biol Chem. 1978;253(5):1357-70. doi: 10.1016/S0021-9258(17)34875-5, PMID 627542.
- 28. Shabir GA. Validation of high-performance liquid chromatography methods for pharmaceutical analysis. Understanding the differences and similarities between validation requirements of the US Food and Drug Administration, the US Pharmacopeia and the International Conference on Harmonization. J Chromatogr A. 2003;987(1-2):57-66. doi: 10.1016/s0021-9673(02)01536-4, PMID 12613797.
- Sahu PK, Ramisetti NR, Cecchi T, Swain S, Patro CS, Panda J. An overview of experimental designs in HPLC method development and validation. J Pharm Biomed Anal. 2018;147:590-611. doi: 10.1016/j.jpba.2017.05.006, PMID 28579052.
- Bhatt DA, Rane SI. QbD approach to analytical RP-HPLC method development and its validation. Int J Pharm Pharm Sci. 2011;3(1):179-87.

- 31. Rao PP. A new method for the simultaneous estimation of methotrexate and folic acid by using RP-HPLC in the bulk and pharmaceutical dosage form. YMER Journal. 2022 July;21(7):880-90.
- Chen J, Fu B, Liu T, Yan Z, Li K. A graphene oxide-DNA electrochemical sensor based on glassy carbon electrode for sensitive determination of methotrexate. Electroanalysis. 2018;30(2):288-95. doi: 10.1002/elan.201700615.
- Khoury S, Chouchou F, Amzica F, Giguere J, Denis R. The authors declare no conflicts of interest with the work presented in this manuscript. 2013.
- 34. Müller L, Mauthe RJ, Riley CM, Andino MM, Antonis DD, Beels C. A rationale for determining, testing, and controlling specific impurities in pharmaceuticals that possess the potential for genotoxicity. Regul Toxicol Pharmacol. 2006;44(3):198-211. doi: 10.1016/j.yrtph.2005.12.001, PMID 16412543.
- Validation of analytical methods primer. ICH Q3A (R2), Impurities in New Drug Substances. Agilent publication; 2006. p. 5990-5140EN.
- ICH. Validation of analytical procedures: definitions and methodology, Geneva. Vol. Q2 (R1); 2005. p. 2005.
- Srinivasan G, Shetty A. Advancements in dry powder inhaler. Asian J Pharm Clin Res. 2017;10(2):8-12. doi: 10.22159/ajpcr.2017.v10i2.14282.
- Liu DQ, Sun M, Kord AS. Recent advances in trace analysis of pharmaceutical genotoxic impurities. J Pharm Biomed Anal. 2010;51(5):999-1014. doi: 10.1016/j.jpba.2009.11.009, PMID 20022442.