

Print ISSN: 2656-0097 | Online ISSN: 0975-1491

Vol 15, Issue 12, 2023

Original Article

ANTICANCER POTENTIAL OF SALACIA CHINENSIS L. AND WOODFORDIA FRUTICOSA (L.) KURZ OF THE WESTERN GHATS FORESTS OF INDIA

NINADA KC¹, BHAGYA N.^{2,3} (D), RODRIGUES WJ^{2,4}, CHANDRASHEKAR R.¹, CHANDRASHEKAR K. R.*^{2,5}

¹Department of Pharmacology, A J Institute of Medical Sciences and Research, Mangalore, DK, India. ²Department of Applied Botany, Mangalore University, Mangalagangotri, Mangalore-574199, India. ³Yenepoya Research Centre, Yenepoya (Deemed to be University) Deralakatte, Mangalore-574018, India. ⁴Department of Biochemistry, Mangalore University, Mangalagangotri, Mangalore-574199, India. ⁵Yenepoya Ayush and Pharmacy Research Centre, Yenepoya (Deemed to be University), Mangalore-574018, India *Corresponding author: Chandrashekar K. R.; *Email: profkrchandrashekar@gmail.com

Received: 22 Aug 2023, Revised and Accepted: 06 Nov 2023

ABSTRACT

Objective: Salacia chinensis and Woodfordia fruticosa are medicinal plants from the Western Ghats of India traditionally used in the treatment of diabetes, diarrhea and worm infections. The current study aims to evaluate the cytotoxic potential of methanolic extract of Salacia chinensis and Woodfordia fruticosa against breast and pancreatic cancers.

Methods: Methanolic extract of dried leaves of *Salacia chinensis* and *Woodfordia fruticosa* were obtained by Soxhlet extraction. The cytotoxic potential of the dried extract was evaluated against human breast (MDA-MB-231) and pancreatic (PANC-1) cancers *in vitro* using MTT-based assay.

Results: The study showed a dose-dependent cytotoxicity of *Salacia chinensis* and *Woodfordia fruticosa* leaf extracts against breast and pancreatic cancers with IC₅₀ values of 124 µg/ml against MDA-MB-231 and 230.5 µg/ml against PANC-1 cells, respectively.

Conclusion: Results indicate the presence of cytotoxic phytochemicals in *Salacia chinensis* and *Woodfordia fruticosa*. Further purification of the extract might be beneficial to isolate the anticancer phytochemical.

Keywords: Salacia chinensis, Woodfordia fruticosa, Breast cancer, Pancreatic cancer, Phytochemical, Cytotoxicity

© 2023 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (https://creativecommons.org/licenses/by/4.0/) DOI: https://dx.doi.org/10.22159/ijpps.2023v15i12.49198. Journal homepage: https://innovareacademics.in/journals/index.php/ijpps

INTRODUCTION

Cancer is the second leading cause of death worldwide after cardiovascular diseases [1]. According to GLOBOCON 2020, breast, lung and colorectal cancers are the most commonly seen cancers and pancreatic cancer is one of the rare cancers worldwide [2]. Currently, surgical resection, chemotherapy and radiation therapy are the commonly available treatment options to cancer patients [3]. However, the side effects associated with chemotherapy drugs demand the discovery of newer anticancer drugs [4]. The development of drug resistance, cytotoxicity to normal cells and metastasis of cancer are the major side effects of chemotherapeutic drugs that lead the patient to death [5-9]. The ideal anticancer agent should be able to selectively target the cancerous cells without harming the normal cells [10-12]. Currently, there are several anticancer drugs isolated from plants such as paclitaxel from Taxus brevifolia L., vincristine, vinblastine, and vinorelbine from Catharanthus roseus G. Don that are used to treat cancer patients [11]. In addition, about 16 plant-derived compounds (eg. flavopiridol from the Indian tree Dysoxylum binectariferum, meisoindigo, from the Chinese plant Indigofera tinctoria) are being tested in clinical trials, 13 are in phase I or II and three are in phase III trials [11]. Besides, several polyphenols from different plants are also reported to exhibit antioxidants, antimicrobial, and anticarcinogenic properties [13-16]. The use of in vitro and in vivo experiments is necessary to screen the bioactive potential of phytochemicals from different sources [12, 15]. However, hardly a limited number of plant resources have been pharmacologically screened for the bioactive potential [17].

In this research, we have used two plants viz, *Salacia chinensis* L. of the family Celastraceae and *Woodfordia fruticosa* (L.) Kurz belongs to the family Lytheraceae, both of which have exhibited several health benefits and curative properties with respect to health conditions like type 2 diabetes, mutagenicity, hepatitis, cardiac disorders, mental disorders and insulin resistance; with documented antimicrobial, antioxidant, immunomodulatory, anti-infertility and

antitumor activities in several in vitro studies [18-20]. The water extract of Salacia chinensis L. (SC) stem or 'Kumpang jed chan' in Thai has been used as a folk remedy to treat patients with cirrhosis in a local hospital with promising results [18]. Different parts of this plant contain many biologically active compounds, such as triterpenes, phenolic compounds, flavonoids. The solvent extracts of S. chinensis root and stem showed potent antioxidant, antiulcer, antidiabetic, hypoglycemic, antiobesity and skin-lightening properties [18]. Further, the cytotoxic effect of S. chinensis was reported against lung (LU), epidermal (KB), liver (Hep-G2) and breast (MCF-7) cells were also reported [21]. A wide range of pharmacological properties, including antihyperglycemic, antioxidant, anti-inflammatory, analgesic, hepatoprotective, antibacterial, gastroprotective and wound healing properties of Woodfordia fruticosa have been recorded in a recent review by Giri et al. [20]. In vitro cytotoxic potential of methanolic extract of W. fruticosa flowers was reported against liver cancer (PLC/PRF/5) and brine shrimp larvae were reported [22, 23]. To the best of our knowledge, the cytotoxic effect of methanolic extract of S. chinensis and W. fruticosa were not studied against breast cancer-MCF7 cells and pancreatic cancer-PANC-1 cells and, therefore, we have evaluated these parameters in the current study using MTT based colorimetric analysis.

MATERIALS AND METHODS

Chemicals and consumables

All the chemicals and plastic wares used in the experiment were of cell culture grade purchased from Himedia, Mumbai, India and Tarsons, India. Standard Cisplatin was purchased from a medical shop. The solvents used were of analytical grade and procured from Merck, Mumbai, India.

Plant samples and extraction of phytochemicals

S. chinensis and W. fruticosa leaf, along with the young stem samples, were collected from the Arboretum (Latitude: 12° 48' 34.02" N

Longitude: 74° 55' 15.99" E) of Mangalore University Campus, Mangalore, India. The plants were identified using Flora of Presidency of Madras [24] and the voucher specimens (MU/AB/NKC/01 and MU/AB/NKC/02 for *S. chinensis* and *W. fruticosa*) were deposited at the herbarium of the Applied Botany Department, Mangalore University, Karnataka, India. The leaf samples were air-dried under shade for about a week and, extracted in 100% and stored until further use.

The phytochemicals in the dried samples were extracted in methanol using Soxhlet for about 16 h. The extract was collected, filtered and concentrated to dryness using a vacuum concentrator at 45 °C. The dried extract was stored under refrigerated conditions until use.

Cell lines and cell culture

Human breast cancer epithelial cell line-MDA-MB-231 and human pancreatic cancer cell line-PANC-1 was purchased from the National Centre for Cell Science (NCCS), Pune, India and grown in Dulbecco's modified Eagle's medium-DMEM (Himedia, Mumbai, India) supplemented with 10% foetal bovine serum (Himedia, Mumbai, India) and 1% penicillin/streptomycin (Himedia, Mumbai, India), and incubated under 5% CO₂ incubator at 37 °C.

Preparation of test sample to evaluate the cell viability

The methanol extract of *S. chinensis* and *W. fruticosa* (1 mg/ml) was dissolved in DMSO and made up to the final volume using DMEM medium. The concentration of DMSO was maintained at less than 0.1% while preparing the sample. The samples were filtered using 0.22 μ m syringe filters.

Cell viability assay

Cell viability was determined by MTT-(3-[4,5-dimethylthiazole-2-yl]-2,5diphenyltetrazolium bromide)-based assay [25]. A hundred microliters of MDA-MB-231 and PANC-1 cells at a cell density of 1x105cells/ml were seeded in a 96-well microtiter plate and incubated overnight under a 5% CO2 incubator at 37°C. The cells were treated with or without S. chinensis and W. fruticosa samples at varying concentrations of 12.5-200 µg/ml for 48 h. Standard drug Cisplatin was used at a concentration of 3.12-50 µg/ml. Cell viability was assessed after the addition of MTT and recording the absorbance at 570 nm using a microplate reader (Synergy H1, BioTek Instruments Inc, USA).

Statistical analysis

All the experiments were tested in triplicate and the data was expressed as mean \pm standard deviation (SD). Statistical analysis of the data was performed by one-way ANOVA using SPSS 21 software at a significance level of p<0.05.

RESULTS AND DISCUSSION

A significant (p<0.05) dose-dependent cytotoxicity was observed for the standard drug cisplatin and the extracts of both *S. chinensis* and *W. fruticosa* against breast and pancreatic cancers (table 1; fig. 1-3). The IC₅₀ value for cisplatin was 2.54 µg/ml and 7.232 µg/ml against MDA-MB-231 and PANC-1 cells, respectively. The crude extract of *S. chinensis* showed IC₅₀ values of 124 µg/ml ad 230.5 µg/ml against MDA-MB-231 and PANC-1 cells, respectively, while *W. fruticosa* showed an IC₅₀ value of 126.53 µg/ml and 91.15 µg/ml against MDA-MB-231 and PANC 1 cells respectively (table 2).

Table 1: Cytotoxic effect of S. chinensis and W. fruticosa against MDA-MB-231 and PANC-1 cells

Sample	Concentration (µg/ml)	% Cytotoxicity (mean±SD)	
-		MDA-MB-231	PANC-1
Cisplatin	0	0	0
-	3.12	50.58±4.30	38.63±4.83
	6.25	54.54±2.02	51.12±1.28
	12	61.75±5.44	59.34±3.72
	25	71.83±2.02	73.96±0.25
	50	90.83±5.81	9.37±1.24
S. chinensis	0	0	0
	12.5	40.05±3.69	27.59±1.03
	25	40.23±5.40	36.88±1.91
	50	46.97±4.36	34.52±0.73
	100	49.09±3.22	39.21±2.48
	200	66.77±4.36	46.30±2.21
W. fruticosa	0	0	0
	12.5	14.40±3.11	22.32±2.62
	25	44.38±5.37	29.79±4.77
	50	44.95±5.29	39.91±7.18
	100	50.92±1.51	51.49±4.07
	200	53.09±7.19	58.14±1.27

The mean±SD values are obtained by taking the average and standard deviations of the results from 3 trials for each of the experiments with n=3 wells.

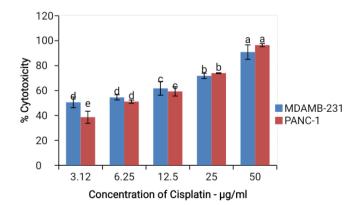


Fig. 1: Effect of Cisplatin on MDA-MB-231 and PANC-1 cells. All the values are obtained by taking the average of the results from 3 trials for each of the experiments with n=3 wells and standard deviations are expressed as error bars. Means with different alphabets (a-d) represent significant differences at 5% level

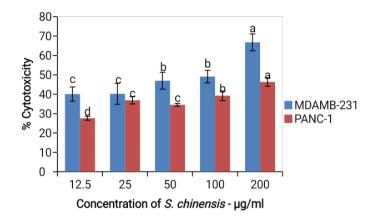


Fig. 2: Effect of *S. chinensis* on MDA-MB-231 and PANC-1. All the values are obtained by taking the average of the results from 3 trials for each of the experiments with n=3 wells and standard deviations are expressed as error bars. Means with different alphabets (a-d) represent significant differences at 5% level

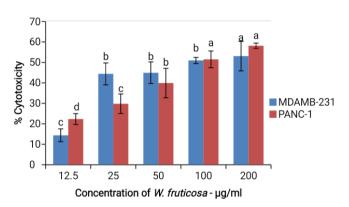


Fig. 3: Effect of *W. fruticosa* on MDA-MB-231 and PANC-1. All the values are obtained by taking the average of the results from 3 trials for each of the experiments with n=3 wells and standard deviations are expressed as error bars. Means with different alphabets (a-d) represent significant differences at 5% level

Sample	IC ₅₀ values* (µg/ml)		
	MDA-MB-231	PANC-1	
Cisplatin	2.54	7.232	
S. chinensis	124	230.5	
W. fruticosa	126.53	91.15	

*All the values are obtained by taking the average of the results from 3 trials for each of the experiments with n=3 wells.

In a study carried out by Khalid *et al.* [25], the anticancer activity of *Sisymbrium officinale* plant extract was noticed at 100 µl of the plant extract on the MCF7 (breast cancer cells) with 6% cancer cell death [26]. However, no results were shown on the 50 µl, 200 µl, or 400 µl concentrations. Musini *et al.* observed the concentration-dependent antiproliferation activity of the methanol extract of *Salacia oblonga* on the breast cancer cell lines (MDA-MB-231) [27]. The results indicated that after treatment with plant extract at a concentration of 30 µg/ml, the cell viability decreased dramatically. Based on their observations, the IC₅₀ value for methanolic aerial and root extracts on breast cancer cells was 35 and 44 µg/ml, respectively after 24 h of incubation. One of the fractions eluted by methanolic extract of *Solonga* also gave a positive cytotoxic effect on EAC with 75% cell death at a concentration of 25 µg/ml and 100% at 50 µg/ml [28].

Ethanolic extract of *W. fruticosa* flowers has been shown to possess anticancer properties in the human liver's PLC/PRF/5 cell lines [22]. On the basis of MTT assay, it was concluded that the synergistic effect of the phytochemicals present in the extract might be responsible for the potential chemoprevention property of *W. fruticosa* flowers in hepatic cancer [22].

Even in the present study, the cytotoxicity of the extracts of both S. chinensis and W. fruticosa increased with increasing concentrations up to 200 µg. The study revealed promising antiproliferative effect of *W*. fruticosa against PANC-1 cell lines, demonstrating an IC50 value of 91.15 µg/ml, while both W. fruticosa and S. chinensis demonstrated moderate antiproliferative effect against MDA-MB-231 cell lines. Probably, the phytochemicals in the extract might be involved in regulating the molecular pathways that are implicated in the growth and progression of cancer, as mentioned by Choudhari et al. [3]. MDA-MB-231 is a triple-negative breast cancer and phytochemicals in the crude extract of W. fruticosa and S. chinensis are less active compared to PANC-1 cells. Though our study suggests the possible anticancer potential of W. fruticosa and S. chinensis, further experiments on the isolation of individual phytochemicals and evaluation of their cytotoxicity against breast and pancreatic cancers are necessary to take it further to drug development.

CONCLUSION

Salacia chinensis and Woodfordia fruticosa are medicinal plants from the Western Ghats of India traditionally used in the treatment of diabetes, diarrhea and worm infections. The current study aims to evaluate the cytotoxic potential of methanolic extract of *S. chinensis* and *W. fruticosa* against breast and pancreatic cancers *in vitro*. The study revealed a dose-dependent cytotoxicity of both *S. chinensis* and *W. fruticosa* against both MDA-MB-231 and PANC-1 cells. The results were compared with the standard cisplatin. Among the 2 plants used, *W. fruticosa* extract showed a lower IC₅₀ value of 91.15 μ g/ml against pancreatic cancer cells compared to breast cancer. Also, *S. chinensis* showed a higher IC₅₀ value. Probably, the phytochemicals present in the extracts are more active against pancreatic cancer compared to breast cancer calls. However, further experiments on the isolation, characterization, and validation of the phytochemical and its cytotoxicity are necessary for further use of this plant in the pharmaceutical industry to develop an anticancer drug.

ACKNOWLEDGEMENT

The authors express their sincere gratitude to the Department of Applied Botany, Mangalore University for permitting to carry out this work.

FUNDING

The senior author acknowledges the funding from the ICMR in the form of Short-Term Studentship-ID: 2018-06561

AUTHORS CONTRIBUTIONS

All the authors have contributed equally.

CONFLICTS OF INTERESTS

All the authors have none to declare

REFERENCES

- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. CA Cancer J Clin. 2023;73(1):17-48. doi: 10.3322/caac.21763, PMID 36633525.
- Deo SVS, Sharma J, Kumar S. GLOBOCAN 2020 report on global cancer burden: challenges and opportunities for surgical oncologists. Ann Surg Oncol. 2022;29(11):6497-500. doi: 10.1245/s10434-022-12151-6, PMID 35838905.
- Choudhari AS, Mandave PC, Deshpande M, Ranjekar P, Prakash O. Phytochemicals in cancer treatment: from preclinical studies to clinical practice. Front Pharmacol. 2019;10:1614. doi: 10.3389/fphar.2019.01614, PMID 32116665.
- Bhagya N, Chandrashekar KR. Identification and quantification of cytotoxic phenolic acids and flavonoids in *Ixora brachiata* by UHPLC-DAD and UHPLC-ESI-MS/MS. Int J Mass Spectrom. 2020;450:116290. doi: 10.1016/j.ijms.2020.116290.
- Wang X, Zhang H, Chen X. Drug resistance and combating drug resistance in cancer. Cancer Drug Resist. 2019;2(2):141-60. doi: 10.20517/cdr.2019.10, PMID 34322663.
- Housman G, Byler S, Heerboth S, Lapinska K, Longacre M, Snyder N. Drug resistance in cancer: an overview. Cancers (Basel). 2014;6(3):1769-92. doi: 10.3390/cancers6031769, PMID 25198391.
- Rueff J, Rodrigues AS. Cancer drug resistance: a brief overview from a genetic viewpoint. Methods Mol Biol. 2016;1395:1-18. doi: 10.1007/978-1-4939-3347-1_1, PMID 26910065.
- Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. Nat Rev Cancer. 2013;13(10):714-26. doi: 10.1038/nrc3599, PMID 24060863.
- Gordaliza M. Natural products as leads to anticancer drugs. Clin Transl Oncol. 2007;9(12):767-76. doi: 10.1007/s12094-007-0138-9, PMID 18158980.
- Saklani A, Kutty SK. Plant-derived compounds in clinical trials. Drug Discov Today. 2008;13(3-4):161-71. doi: 10.1016/j.drudis.2007.10.010, PMID 18275914.
- 11. Gerson Cwilich R, Serrano Olvera A, Villalobos Prieto A. Complementary and alternative medicine (CAM) in Mexican

patients with cancer. Clin Transl Oncol. 2006;8(3):200-7. doi: 10.1007/s12094-006-0011-2, PMID 16648120.

- Tascilar M, De Jong FA, Verweij J, Mathijssen RHJ. Complementary and alternative medicine during cancer treatment: beyond innocence. Oncologist. 2006;11(7):732-41. doi: 10.1634/theoncologist.11-7-732, PMID 16880232.
- Molassiotis A, Browall M, Milovics L, Panteli V, Patiraki E, Fernandez Ortega P. Complementary and alternative medicine use in patients with gynecological cancers in Europe. Int J Gynecol Cancer. 2006;16(1)Suppl 1:219-24. doi: 10.1111/j.1525-1438.2006.00309.x, PMID 16515594.
- Taher ZM, Agouillal F, R LJ, Marof AQ, Dailin DJ, Nurjayadi M. Anticancer molecules from *Catharanthus roseus*. Indonesian J Pharm. 2019;30(3):147. doi: 10.14499/indonesianjpharm30iss3pp147.
- Solowey É, Lichtenstein M, Sallon S, Paavilainen H, Solowey E, Lorberboum-Galski H. Evaluating medicinal plants for anticancer activity. ScientificWorldJournal. 2014;2014:721402. doi: 10.1155/2014/721402, PMID 25478599.
- Kumar D, Sharma M, Sorout A, Saroha K, Verma S. *Woodfordia fruticosa* Kurz: a review on its botany, chemistry and biological activities. J Pharmacogn Phytochem. 2016;5(3):293-8.
- 17. Bhagya N, Chandrashekar KR. *In vitro* pharmacological potential of *Epiprinus mallotiformis* -an endemic species of Western Ghats. IJNPR. 2018;9(2):108-16.
- Deokate UA, Khadabadi SS, Hamza AA, Hassanin SO, Hamza S. Phytopharmacological aspects of *Salacia chinensis*. J Pharmacogn Phytother. 2012;4:1-5.
- Wayal SR, Gurav SS. Pharmacognostic and phytochemical investigation of potentially important plants of Western Ghats, India. Int J Pharm Sci Res. 2019;10(6):3101-8.
- Giri S, Dey G, Sahu R, Paul P, Nandi G, Dua TK. Traditional uses, phytochemistry and pharmacological activities of woodfordia fruticosa (L) kurz: a comprehensive review. Indian J Pharm Sci. 2023;85(1):1-12. doi: 10.36468/pharmaceuticalsciences.1062.
- Minh TT, Hoang Anh NT, Thang VD, Van Sung T. Study on chemical constituents and cytotoxic activities of salacia chinensis growing in vietnam. Zeitschrift fur Naturforschung B. 2010;65(10):1284-8. doi: 10.1515/znb-2010-1017.
- Nitha A, Prabha SP, Ansil PN, Latha MS. Antiproliferative effect of *Woodfordia fruticosa* Kurz flowers on experimentally induced hepatocellular carcinoma in rats and in human hepatoma cell line. J Pharm Res. 2013;6(2):239-48. doi: 10.1016/j.jopr.2013.02.003.
- Baravalia Y, Vaghasiya Y, Chanda S. Brine shrimp cytotoxicity, anti-inflammatory and analgesic properties of *Woodfordia fruticosa* kurz flowers. Iran J Pharm Res. 2012;11(3):851-61. PMID 24250512.
- Gamble JS. Flora of the presidency of madras, Vol. Calcutta, India: Sri Gouranga Press Pvt. Ltd; 1958. p. I-III.
- Khalid M, Amayreh M, Sanduka S, Salah Z, Al-Rimawi F, Al-Mazaideh GM. Assessment of antioxidant, antimicrobial, and anticancer activities of Sisymbrium officinale plant extract. Heliyon. 2022;8(9):e10477. doi: 10.1016/j.heliyon.2022.e10477, PMID 36105455.
- Musini A, Rao JP, Giri A. Phytochemicals of Salacia oblonga responsible for free radical scavenging and antiproliferative activity against breast cancer cell lines (MDA-MB-231). Physiol Mol Biol Plants. 2015;21(4):583-90. doi: 10.1007/s12298-015-0317-z, PMID 26600684.
- Augusti KT, Joseph P, Babu TD. Biologically active principles isolated from *Salacia oblonga* Wall. Indian J Physiol Pharmacol. 1995;39(4):415-7. PMID 8582758.
- Choi J, Park JG, Ali MS, Choi SJ, Baek KH. Systematic analysis of the anticancer agent taxol-producing capacity in *Collectotrichum* species and use of the species for taxol production. Mycobiology. 2016;44(2):105-11. doi: 10.5941/MYCO.2016.44.2.105, PMID 27433121.