

International Journal of Pharmacy and Pharmaceutical Sciences

Print ISSN: 2656-0097 | Online ISSN: 0975-1491

Vol 16, Issue 10, 2024

Original Article

DEVELOPMENT AND VERIFICATION OF UV SPECTROPHOTOMETRIC TECHNIQUE FOR DETERMINING N-ACETYLCYSTEINE IN TABLET FORMULATIONS

RAMARAO NADENDLA^{1*}, MADAMANCHI MOUNIKA¹, VENKATA SAI RANJITHADUBBA¹, MANDAVA JAHNAVI¹, SOWJANYASONTI¹, YAKSHITHAYARRA¹, VENKATA SURESH P.², SIVA PRASAD MORLA²

¹Department of Pharmaceutics, Chalapathi Institute of Pharmaceutical Sciences, Chalapathi Nagar, Lam-522034, Guntur, Andhra Pradesh, India. ²Department of Pharmaceutical Analysis, Chalapathi Institute of Pharmaceutical Sciences, Chalapathi Nagar, Lam-522034, Guntur, Andhra Pradesh, India

*Corresponding author: Ramarao Nadendla; *Email: nadendla2000@yahoo.in

Received: 30 Mar 2024, Revised and Accepted: 11 Jun 2024

ABSTRACT

Objective: This study aimed to determine the concentration of N-Acetyl Cysteine (NAC) in tablet formulations using UV-Visiblespectroscopy.

Methods: A precise and accurate UV spectrophotometric method was developed for determining N-acetylcysteine in tablets, using 0.1N NaOH as a diluent. The drug's purity was assessed using UV-visible spectrophotometry, which validated linearity, accuracy, precision, specificity, limit of detection, and quantification.

Results: The calibration curve had a high correlation coefficient ($r^2 = 0.9992$) and maximum absorbance at 235 nm. There was no interference in dosage. The results showed 100.17% mean % recovery and 100.27% tablet assay, with a precision of 0.60% and 0.57%, respectively. The method was robust and rugged.

Conclusion: The method was evaluated using statistical parameters, including precision, accuracy, linearity, recovery, and robustness. The results showed no significant differences compared to other methods. The method can be used to analyse pharmaceutical formulations quickly, and no significant differences in mean values and standard deviations were found.

Keywords: N-acetylcysteine, UV-Visiblespectroscopy, ICH guidelines, Dosage form, Validation

© 2024 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (https://creativecommons.org/licenses/by/4.0/) DOI: https://dx.doi.org/10.22159/ijpps.2024v16i10.51004 Journal homepage: https://innovareacademics.in/journals/index.php/ijpps

INTRODUCTION

N-acetyl cysteine (NAC) is a medication used to decrease mucus thickness in the lungs, which underpins respiratory conditions. It also functions as an antidote for paracetamol toxicity when administered intravenously [1-5]. It is produced from the amino acid l-cysteine and acts as a glutathione (GSH) precursor, which affects various disease mechanisms and oxidative stress. These mechanisms, in turn, affect neurotransmitters glutamate and dopamine and apoptosis, inflammation, and mitochondrial function. NAC is widely available as a nutraceutical in several countries, including the US, Canada, and Australia, and has been studied for its potential therapeutic benefits in laboratory and animal settings. Its antioxidant properties make it a popular choice for treating paracetamol overdose, a common occurrence in countries such as Australia, the United Kingdom, and the United States, where instances of paracetamol poisoning have increased by 40% in the past decade. The NAC market, valued at USD 1083.78 million in 2021, is projected to grow steadily, reaching USD 7598.08 million by 2030, with a robust Compound Annual Growth Rate (CAGR) of 21.5% between 2023 and 2030 [6, 7]. Several methods for measuring N-acetyl-L-cysteine have been documented in the literature, including spectrophotometric and differential pulse polarography [8], liquid chromatography-tandem mass spectrometry [9-13], HPTLC [14, 15], HPLC [16-31], UV-VIS spectroscopy [32-39], chemiluminescence [40-42], Electrochemical [43-48], turbidimetry and nephelometry [49], capillary electrophoresis [50-52], and. These methods are commonly used to quantify N-acetyl-L-cysteine and other drugs, such as clomiphene citrate, arginine, and cefixime trihydrate. Reverse-phase HPLC and ion pair chromatography are commonly used to test substances related to NAC in both bulk and drug products. However, more expensive and less readily available techniques such as LC-UV-MS and capillary electrophoresis-mass spectrometry have also been reported for quantifying substances related to N-acetylcysteine. Our study aimed to develop a costeffective, precise, and specialized UV-Visible spectroscopy method for analysing NAC, as existing methods are expensive and laborious. Our process addresses the market demand for NAC analysis and has been validated by the International Conference on Harmonization's Q2R1 guidelines. Although a literature review showed no UV-visible spectroscopic technique available for the analysis of NAC, our method, which uses 0.1N NaOH as a diluent at a detection wavelength of 235 nm, is more precise than previous studies of a similar nature to the existing methods.



Fig. 1: Structure of N-acetylcysteine

MATERIALS AND METHODS

Materials

N-acetylcysteine was generously provided as a gift sample using Alphamed Formulations (Hyderabad). The investigational study utilized tablet formulation of N-acetylcysteine (NAC) manufactured by Sun Glow Ltd (India). Sodium hydroxide was procured from Thermo Fisher Scientific India Pvt., Ltd. (Mumbai, India).

Instrumentation

Experiments were conducted using a UV-Visibledouble-beam spectrophotometer (Labindia 3092, UV-WIN software) and an

analytical balance (Shimadzu AY220) for weighing; we also used an ultrasonicator (Oscar Microclean 103) to sonicate the standard and product sample solutions.

Methods

Method development

Selection of solvent

NAC was dissolved in several solvents to identify the solvent with the optimal solubility. Specifically, methanol, ethanol, acetonitrile, and diluted acidic and basic buffers were used. Consequently, Nacetylcysteine demonstrated solubility more in 0.1N NaOH.

Determination of wavelength of maximum absorbance (λmax)

The standard working solution containing 100 μ g/ml NAC was scanned using the full scan mode and a medium scanning speed across the entire range of UV-visible spectrophotometers, from 200-800 nm, with methanol as the blank. Following the acquisition of the spectrum, the λ max was determined to be 235 nm (fig. 2).

Preparation of the standard solution

Solubility tests confirmed the suitability of a 0.1N NaOH solution as the solvent. NAC (100 mg) was dissolved in 100 ml of 0.1N NaOH, resulting in a 1000 μ g/ml concentration. Next, 10 ml of the resulting solution was dispensed and diluted to 100 ml with NaOH (using 0.1N) NaOH. Afterward, 0.5 ml, 1 ml, 1.5 ml, 2 ml, and 2.5 ml aliquots were pipetted, and each was topped up to 10 ml.

Preparation of sample solutions

Twenty tablets with an average weight were crushed into a fine powder, and 1385.54 mg of this powder was weighed and transferred into a 100 ml volumetric flask. Next, three-quarters of the diluent was added, and the mixture was sonicated for 30 min. The volume was then diluted with the diluent and filtered. From the resulting stock solution, 5 ml was pipetted and adjusted to 25 ml with diluent. Subsequently, 0.1 ml was pipetted and made up to 10 ml with diluent, resulting in a 15 μ g/ml concentration. This study estimated NAC using a UV-Visiblespectrophotometer at a wavelength of maximum absorbance of 235 nanometers in 0.1N NaOH solution.



Fig. 2: N-acetylcysteine wavelength of maximum absorbance at 235 nm

Validation of method parameters

The International Council for Harmonisation (ICH) guidelines were followed to validate the method's suitability, linearity, precision, and accuracy [53].

System suitability

Six identical standard solutions were injected into the UV system at 235 nm, and the percentage Relative Standard Deviation (%RSD) was determined.

Linearity

The N-acetylcysteine working standard solutions were prepared by adding appropriate volumes to several 10 ml volumetric flasks, followed by dilution with distilled water to achieve concentrations of 5, 10, 15, 20, and 25 μ g/ml. Calibration curves were created by relating the absorbance values to the concentrations, and regression equations were established to analyze the behaviour of the drug.

Range

The range of analytical methods was established by determining the interval between the upper and lower levels of the calibration curve plotted on a graph.

Intraday precision (repeatability) and inter-day precision (intermediate precision)

Intraday accuracy involves examining the drug at a concentration of 15 μ g/ml six times throughout the day, whereas inter-day accuracy stretches the analysis over two consecutive days. Repeatability (intraday) was assessed by analyzing six samples with the same drug

concentration (15 $\mu g/ml)$ and reporting the absorbance fluctuations in terms of relative standard deviation. This method allows the evaluation of the dependability and uniformity of the analytical technique.

Accuracy/recovery study

To evaluate the method's reliability, the accuracy of NAC concentration was determined using the standard addition technique at three different concentrations (50%, 100%, and 150%). The mean percentage recoveries were calculated, and the recovery values are summarised in table 4. This approach offers valuable information regarding the ability of the method to measure N-acetylcysteine concentrations at different levels accurately.

Limit of detection and quantification (LOD and LOQ)

LOD

The lowest analyte concentration in a test sample can be clearly distinguished from zero.

LOQ

UV spectroscopy was used to compare the signals from the samples with those of the blank samples to accurately determine the minor concentration of analyte in a test sample while maintaining acceptable repeatability.

RESULTS

Linearity and range

To confirm the dependability of the proposed method, calibration curves were constructed in the concentration range of $5-25 \ \mu g/ml$. NAC followed Beer's law within this $5-25 \ \mu g/ml$ range, demonstrating exceptional adherence, with a regression equation of Y = 0.0347x+0.0029. The correlation coefficient (r) was more significant than 0.9992, indicating the robustness of the standard curve. All stock solutions and working standards were prepared in 0.1N NaOH, and the calibration curve data are presented in table 1 and fig. 3.

Precision

The accuracy of the technique was assessed by conducting six separate tests on the sample preparation, calculating the standard deviation, and evaluating precision and reproducibility. To determine precision and reproducibility, replicate analyses of standard solutions were performed. Concentrations within the calibration range were prepared using 0.1N NaOH and analyzed using respective calibration curves. The results showed that the technique demonstrated excellent precision and accuracy, with low intra-and inter-day variability, as indicated by the Relative Standard Deviation (RSD %) in table 2.

Table 1: Linearity data			
Concentration (µg/ml)	*Absorbance		
5	0.182		
10	0.359		
15	0.508		
20	0.694		
25	0.878		
r ²	0.9992		
Range	5 to 25 μg/ml		
Slope	0.0347		
Intercept	0.0029		

*n=6



Fig. 3: Linearity of N-acetylcysteine

Table 2: Precision table

S. No.	*Intra-day precision	*Inter day precision		
		Day-1	Day-2	
1	0.506	0.507	0.507	
2	0.511	0.512	0.512	
3	0.513	0.516	0.516	
4	0.508	0.512	0.512	
5	0.509	0.511	0.511	
6	0.514	0.513	0.513	
Mean	0.510	0.512	0.5118	
SD	0.0030	0.2930	0.0029	
% RSD	0.60	0.57	0.57	

*n= 6, SD: Standard deviation %RSD= Relative standard deviation, %RSD value for intra-and inter-day precision was<2% (0.60 and 0.57%, respectively), which is within the limit; hence, the precision parameter was validated.

Accuracy/recovery

Experiments involving the addition of known quantities of the pure drug to pre-analyzed formulations were carried out to confirm the accuracy of the developed method. The data obtained for the drug's accuracy demonstrated a high percentage recovery, ranging from 99.97 to 100.53. Notably, the developed method exhibited excellent precision and accuracy, making it applicable for determining NAC tablets in pharmaceuticals without interference from excipients (table 3).

Table 3: Accuracy data

% Level	Standard absorbance	Sample absorbance	% Recovery	*Average % recovery	*mean % recovery
50%	0.509	0.206	100.69	100.53	100.17
		0.207	101.18		
		0.204	99.71		
100%		0.512	100.11	99.97	
		0.508	99.31		
		0.514	100.49		
150%		0.904	100.44	100.02	
		0.902	100.19		
		0.892	99.43		

*n= 3

Stability duration (h)	Concentration (µg/ml)	*Conc. found (mean±SD, μg/ml)	RSD (%)	% Potency
0.0	15	14.96±0.0051	0.234	99.89
1.0		15.06±0.0013	0.532	98.41
5.0		14.83±0.0021	0.631	99.74
10.0		14.92±0.0047	0.832	99.87
24.0		15.09 ± 0.0034	0.725	98.95

Table 4: Short-term stability determined by the proposed method

*n= 6, SD: Standard deviation, %RSD= Relative standard deviation

Table 5: Content of NAC in marketed products determined by the proposed method.

Formulation	Drug	Label claim (mg)	*%Amount found±SD	%RSD
Tablet	N-acetylcysteine	600 mg	101.27±0.0019	0.268

*n= 6, SD: Standard deviation, %RSD= Relative standard deviation

Limit of detection (LOD) and quantitation (LOQ)

The sensitivity parameter was determined by the LOD and LOQ for NAC and was found to be $0.35~\mu g/ml$ and $1.0~\mu g/ml$, respectively.

Stability study

Table 4 shows the results of the repeatability study, where the samples were stored at room temperature for 24 h and then analyzed the next day to gauge their short-term stability.

Content of NAC in marketed brands

The proposed method effectively measured NAC in tablet formulations without any interference. The outcomes of our assay were consistent with this claim, which validates our approach [table 5]. To guarantee the long-term dependability of the method, we conducted a stability assessment of NAC in a 0.1N NaOH solution.

DISCUSSION

Developing of a new spectroscopic method for the estimation of drugs in their dosage forms has raised recently due to its important role in pharmaceutical analysis and development [48]. Different solvents like methanol, ethanol, acetonitrile, and diluted acidic and basic buffers were used to determine the studied drug's solubility peak quality and peak shape. Among these, 0.1 NaoH fulfilled all criteria in showing better solubility and giving better peak quality. NAC wavelength was selected at 235 nm, whereas no absorption was detected due to excipients at NAC wavelength. This shows no interference from the excipients at the selected wavelength. This innovation promises to significantly reduce the sample preparation and material costs typically associated with routine analyses.

Lea Kukoc-Modun *et al.* [49] method solutions of NAC used in the method are only stable for a short period of time. They should be analysed within 24 h to ensure method stability and accuracy. In the present study, the solution stability study led that the absorbance of NAC was 98% to 99% of the initial value (table 4) with a %RSD of 0.99, which indicates that the NAC solution in the 0.1N NaoH was stable for at least 24 h. Solution stability is an important criterion to ensure that the prepared samples are stable and that no changes could affect the samples during the test period.

Linearity was calculated using six different 5-25 μ g/ml concentrations. These data showed linear absorbance readings through the selected range. The same R² was found by Swati S. Agawane 1 *et al.* for the determination of Clomiphene citrate and Acetylcysteine in its dosage forms by using the UV-visible spectroscopic method [50].

For comparison, the spectrophotometric methods were reported by Bondarenko N *et al.* [51] and Maja Biocic *et al.* [52]. The articles may not provide results from the analysis of pharmaceutical samples containing NAC and comprehensive information on the validation parameters of the developed method, such as accuracy, precision, linearity, and robustness, which are essential for method reliability. In this contrast, the precision test results are shown in table 2. The % RSD was less than 2%, indicating that the precision values of the validated method were within the accepted limits.

The accuracy of the method was further ascertained through the recovery studies. To the drug solutions of the granules or the tablet powder, the standard solutions of the synthetic NAC were added at four three different concentrations. The recovery of added NAC was 99–100% (table 3), with the relative standard deviation (% RSD) less than 2%. This indicates that the developed method had a higher recovery than the reported method [53, 54].

Unlike some reported methods, the proposed method is free from drastic experimental conditions such as flow injection, solvent effect, heating, and reagents [52-54]. It is also worth mentioning that the proposed method was performed in the UV region ($\lambda = 235$ nm) away from UV-absorbing interfering excipient materials, which might be dissolved from pharmaceutical formulations.

CONCLUSION

Using UV-visible spectroscopy, a method for determining NAC in tablet form was developed and validated. The technique was accurate and cost-effective, making it suitable for the routine quality control analysis of commercially available formulations.

ACKNOWLEDGEMENT

The authors thank the Chalapathi Institute of Pharmaceutical Sciences, Lam, Guntur, for providing the necessary laboratory facilities. Furthermore, the authors extend their gratitude to the institute's management. In addition, the authors acknowledge the Alphamed Formulations located in Hyderabad for providing the API necessary for this study.

FUNDING

Nil

AUTHORS CONTRIBUTIONS

All the authors played an equal role in this study. Mounika, Venkata Sai Ranjitha Dubba, Mandava Jahnavi, Sowjanya Sonti, and Yakshitha Yarra collected the necessary chemicals and reagents for the experimental work in our laboratories. Dr Ramarao Nadendla contributed to the interpretation of the data. Dr. Venkata Suresh and Siva Prasad Morla drafted and revised the manuscript to ensure its quality.

CONFLICT OF INTERESTS

The authors declare no conflicts of interest.

REFERENCES

- Williams DA, Lenke TL. Foye's principles of medicinal chemistry. 6th ed, Lippincott Williams and Williams. Ltd. Vol. 41(II). India: Wolters Kluwer Health, Pvt; 2008. p. 1127-46.
- 2. https://pubchem.ncbi.nlm.nih.gov/compound/acetylcysteine [Last accessed on 15 Jan 2024].

- Ooi SL, Green R, Pak SC. N-acetylcysteine for the treatment of psychiatric disorders: a review of current evidence. Biomed Res Int. 2018;2018:2469486. doi: 10.1155/2018/2469486, PMID 30426004.
- Kelly GS. Clinical applications of N-acetylcysteine. Altern Med Rev. 1998;3(2):114-27. PMID 9577247.
- Ayene SI, Kale RK, Srivastava PN. Radioprotective effect of 2mercaptopropionyl glycine on radiation-induced lipid peroxidation and enzyme release in erythrocytes. Int J Radiat Biol Relat Stud Phys Chem Med. 1988;53(4):629-39. doi: 10.1080/09553008814550951, PMID 3258297.
- Barbey F, Joly D, Rieu P, Méjean A, Daudon M, Jungers PP. Medical treatment of cystinuria: critical reappraisal of long-term results. J Urol. 2000;163(5):1419-23. doi: 10.1016/s0022-5347(05)67633-1, PMID 10751848.
- Holdiness MR. Clinical pharmacokinetics of N-acetylcysteine. Clin Pharmacokinet. 1991;20(2):123-34. doi: 10.2165/00003088-199120020-00004, PMID 2029805.
- Haggag R. Derivatization with 4-Chloro-7-Nitro-2,1,3benzoxadiazole for the spectrophotometric and differential pulse polarographic determination of acetylcysteine and captopril. Sci Pharm. 2008;76(1):33-48. doi: 10.3797/scipharm.0711-02.
- Toussaint B, Pitti C, Streel B, Ceccato A, Hubert P, Crommen J. Quantitative analysis of N-acetylcysteine and its pharmacopeial impurities in a pharmaceutical formulation by liquid chromatography-UV detection-mass spectrometry. J Chromatogr A. 2000;896(1-2):191-9. doi: 10.1016/S0021-9673(00)00741-X, PMID 11093654.
- Lu C, Liu G, Jia J, Gui Y, Liu Y, Zhang M. Liquid chromatographytandem mass spectrometry method for determination of Nacetylcysteine in human plasma using an isotope-labeled internal standard. Biomed Chromatogr. 2011;25(4):427-31. doi: 10.1002/bmc.1465, PMID 21374646.
- 11. Celma C, Allue JA, Prunonosa J, Peraire C, Obach R. Determination of N-acetylcysteine in human plasma by liquid chromatography coupled to tandem mass spectrometry. J Chromatogr A. 2000 Feb;870(1-2):13-22. doi: 10.1016/s0021-9673(99)01078-x, PMID 10722057.
- Barricklow J, Ryder TF, Furlong MT. Quantitative interference by cysteine and N-acetylcysteine metabolites during the LC-MS/MS bioanalysis of a small molecule. Drug Metab Lett. 2009 Aug;3(3):181-90. doi: 10.2174/187231209789352148, PMID 19702543.
- Longo A, Di Toro M, Galimberti C, Carenzi A. Determination of Nacetylcysteine in human plasma by gas chromatography-mass spectrometry. J Chromatogr. 1991;562(1-2):639-45. doi: 10.1016/0378-4347(91)80614-i, PMID 2026726.
- Magesh AR, Dhanaraju MD. Simultaneous determination of nacetyl cysteine and taurine by HPTLC method in active pharmaceutical ingredient and pharmaceutical dosage form. Am J Anal Chem. 2017;8(11):742-51. doi: 10.4236/ajac.2017.811054.
- Abdelaleem EA, Naguib IA, Zaazaa HE, Hussein EA. Development and validation of HPLC and HPTLC methods for determination of cefoperazone and its related impurities. J Chromatogr Sci. 2016;54(2):179-86. doi: 10.1093/chromsci/bmv125, PMID 26306573.
- Ourique AF, Coradini K, Chaves PS, Garcia SC, Pohlmann AR, Guterres SS. A LC-UV method to assay N-acetylcysteine without derivatization: analyses of pharmaceutical products. Anal Methods. 2013;5(13). doi: 10.1039/c3ay40426a.
- Gowda AP, Schaefer AD, Schuck TK. A simple RP-HPLC method for the stability-indicating determination of N-acetyl-L-cysteine and N, N'-diacetyl-L-cystine in cell culture media. Cell Gene Therapy Insights. 2020;6(2):303-23. doi: 10.18609/cgti.2020.041.
- Mathew EM, Ravi A, Rameshwar N, Sudheer M, Krishnamurthy B. Development and validation of an analytical method for related substances in N-acetyl-L-cysteine effervescent tablets by RP-HPLC. Indian J Pharm Educ Res. 2017;51(4):626-35. doi: 10.5530/ijper.51.4.93.
- 19. Vamseekrishna G, Sanjeev G, Ramanjaneya Varma KJ, Umesh K, Mahesh U, Jayadev B. Stability indicating RP-HPLC method for

simultaneous estimation of N-acetylcysteine and ascorbic acid. Int J Pharm Pharm Res. 2022;24(2):153-70.

- Petrlova J, Mikelova R, Stejskal K, Kleckerova A, Zitka O, Petrek J. Simultaneous determination of eight biologically active thiol compounds using gradient elution-liquid chromatography with coul-array detection. J Sep. 2006;29(8):1166-73. doi: 10.1002/jssc.200500425, PMID 16830732.
- Ercal N, Oztezcan S, Hammond TC, Matthews RH, Spitz DR. Highperformance liquid chromatography assay for N-acetylcysteine in biological samples following derivatization with N-(1pyrenyl)maleimide. J Chromatogr B Biomed Appl. 1996;685(2):329-34. doi: 10.1016/s0378-4347(96)00196-x, PMID 8953175.
- 22. Johannson M, Lenngren S. Determination of cysteine, glutathione and N-acetylcysteine in plasma by ion-pair reversed-phase liquid chromatography and post-column derivatization. J Chromatogr. 1988;432:65-74. doi: 10.1016/s0378-4347(00)80634-9, PMID 3220916.
- Kumar CH, Uma G, Arcot S. Method development and validation for simultaneous assessment of clomiphene citrate and n-acetyl cysteine in mixed tablet dosage form using RP-UPLC. World J Pharm Pharm Sci. 2014;3:1773-80.
- 24. Jyothi NN, Pasha SI. Development and validation of a new RP-HPLC method for simultaneous estimation of n-acetylcysteine and l-arginine in combined dosage form. Orient J Chem. 2014;30(3):1371-8. doi: 10.13005/ojc/300357.
- Dabir J, Mathew EM, Moorkoth S. Analytical method development and validation of RP-HPLC method for simultaneous estimation of n-acetyl cysteine and cefexime from its fixed-dose combination. Res J Pharm Technol X. 2016;9(7):835-42. doi: 10.5958/0974-360X.2016.00158.X.
- 26. Sana S, Rajania A, Sumedhab N, Pravin P, Shripad N. Development and validation of RP-HPLC method for the estimation of N-acetylcysteine in wet cough syrup. Int J Drug Dev Res. 2012;4:284-93. doi: 10.5937/arhfarm1403271i.
- Farquhar J, Finlay G, Ford PA, Martin Smith M. A reversed-phase high-performance liquid chromatographic assay for the determination of N-acetylcysteine in aqueous formulations. J Pharm Biomed Anal. 1985;3(3):279-85. doi: 10.1016/0731-7085(85)80033-9, PMID 16867688.
- Ourique AF, Coradini K, Chaves PS, Garcia SC, Pohlmann AR, Guterres SS. A LC-UV method to assay N-acetylcysteine without derivatization: analyses of pharmaceutical products. Anal Methods. 2013;5(13):3321-7. doi: 10.1039/C3AY40426A.
- Acheampong A, Gyasi WO, Darko G, Apau J, Addai Arhin S. Validated RP-HPLC method for simultaneous determination and quantification of chlorpheniramine maleate, paracetamol and caffeine in tablet formulation. Springerplus. 2016;5:625. doi: 10.1186/S40064-016-2241-2, PMID 27330891.
- Mamolo MG, Vio L, Maurich V. Simultaneous quantitation of paracetamol, caffeine and propyphenazone by high-pressure liquid chromatography. J Pharm Biomed Anal. 1985;3(2):157-64. doi: 10.1016/0731-7085(85)80019-4, PMID 16867698.
- Houze P, Gamra S, Madelaine I, Bousquet B, Gourmel B. Simultaneous determination of total plasma glutathione, homocysteine, cysteinylglycine, and methionine by highperformance liquid chromatography with electrochemical detection. J Clin Lab Anal. 2001;15(3):144-53. doi: 10.1002/jcla.1018, PMID 11344530.
- Raggi MA, Cavrini V, Di Pietra AM. Colorimetric determination of acetylcysteine, penicillamine, and mercaptopropionylglycine in pharmaceutical dosage forms. J Pharm Sci. 1982;71(12):1384-6. doi: 10.1002/jps.2600711218, PMID 7153888.
- 33. Ogwu V, Cohen G. A simple colorimetric method for the simultaneous determination of N-acetylcysteine and cysteine. Free Radic Biol Med. 1998;25(3):362-4. doi: 10.1016/s0891-5849(98)00024-0, PMID 9680182.
- Gomes GC, Salgado HR. Validation of UV spectrophotometric method for determination of lomefloxacin in pharmaceutical dosage form. Acta Farm Bonaerense. 2005;243:406.
- Kukoc-Modun L, Radić N. Spectrophotometric determination of N-Acetyl-L-cysteine and N-(2-Mercaptopropionyl)-glycine in pharmaceutical preparations. Int J Anal Chem. 2011. doi: 10.1155/2011/140756.

- 36. Agawane SS, Ashpak M, Tamboli MS, Patil SS, Swati T Mane. Simultaneous equation method for the estimation of clomiphene citrate and acetylcysteine by UV-visible spectrophotometry. Int J Pharm Res Appl. 2022;7(2 Mar-Apr):963-71.
- 37. Biocic M, Karabatić D, Tomic D, Kukoc Modun L. Spectrophotometric determination of N-acetyl-L-cysteine in pharmaceutical formulations by flow injection and sequential injection analysis: comparison of the methods. Croat Chem Acta. 2023;96(2):91-7. doi: 10.5562/cca4016.
- 38. Vedangkinjawadekara. Snehalathaboddua, deepalijadhava, sudharathod. Absorption correction method for the simultaneous estimation of n-acetyl-L-cysteine and ambroxol hydrochloride in bulk and in combined tablet dosage form. Int J Pharm Pharm Sci. 2016;8(5):191-5.
- Hasan S, Sultan S. Utility of 2,2'-bipyridyl reagent in spectrophotometric assay of acetylcysteine in pure form and in its pharmaceutical preparations. JES. 2022;31(3):42-53. doi: 10.33899/edusj.2022.134059.1246.
- 40. Li H, Du J. Sensitive chemiluminescence determination of three thiol compounds based on Cu(II)-catalyzing luminol reaction in the absence of an oxidant. Anal Lett. 2009 Aug 21;42(13):2131-40. doi: 10.1080/00032710903082754.
- Samadi Maybodi A, Akhoondi R. Trace analysis of N-acetyl-Lcysteine using luminol-H2O2 chemiluminescence system catalyzed by silver nanoparticles. Luminescence. 2015 Sep 1;30(6):775-9. doi: 10.1002/bio.2819, PMID 25428294.
- Bondarenko NY, Blazheyevskiy MY. Determination of Nacetylcysteine in tablets by means of chemiluminescence inhibition method. MOCA. 2018;13(3):110-4. doi: 10.17721/moca.2018.110-114.
- 43. Foroughi MM, Beitollahi H, Tajik S, Akbari A, Hosseinzadeh R. Electrochemical determination of N-acetylcysteine and folic acid in pharmaceutical and biological samples using a modified carbon nanotube paste electrode. International Journal of Electrochemical Science. 2014 Dec 1;9(12):8407-21. doi: 10.1016/S1452-3981(23)11056-X.
- 44. Da Silva IS, Araújo MF, Ferreira HA, Varela Jde J, Tanaka SM, Tanaka AA. Quantification of N-acetylcysteine in pharmaceuticals using cobalt phthalocyanine modified graphite electrodes. TalantaTalanta. 2011;83(5):1701-6. doi: 10.1016/j.talanta.2010.11.070, PMID 21238771.
- 45. Toito Suarez W, Marcolino LH, Fatibello Filho O. Voltammetric determination of N-acetylcysteine using a carbon paste

electrode modified with copper(II) hexacyanoferrate(III). Microchem J. 2006;82(2):163-7. doi: 10.1016/i.microc.2006.01.007.

- 46. Ensafi AA, Karimi Maleh H, Mallakpour S, Hatami M. Simultaneous determination of N-acetylcysteine and acetaminophen by voltammetric method using N-(3,4dihydroxyphenethyl)-3,5-dinitrobenzamide modified multiwall carbon nanotubes paste electrode. Sens Actuators B. 2011;155(2):464-72. doi: 10.1016/j.snb.2010.12.048.
- Shahrokhian S, Kamalzadeh Z, Bezaatpour A, Boghaei DM. Differential pulse voltammetric determination of Nacetylcysteine by the electrocatalytic oxidation at the surface of carbon nanotube-paste electrode modified with cobalt salophen complexes. Sens Actuators B. 2008;133(2):599-606. doi: 10.1016/j.snb.2008.03.034.
- 48. Shaidarova LG, Gedmina AV, Zhaldak ER, Chelnokova IA, Budnikov GK. Voltammetric determination of acetylcysteine in drugs using an electrode modified by an osmium hexacyanocobaltate film. Pharm Chem J. 2014;47(12):670-4. doi: 10.1007/s11094-014-1029-3.
- 49. Santos VB, Guerreiro TB, Suarez WT, Faria RC, Fatibello Filho O. Evaluation of turbidimetric and nephelometric techniques for analytical determination of n-acetylcysteine and thiamine in pharmaceutical formulations employing a lab-made portable microcontrolled turbidimeter and nephelometer. J Braz Chem Soc. 2011;22(10):1968-78, doi: 10.1590/S0103-50532011001000019.
- Dette C, Watzig H. Separation of enantiomers of N-acetylcysteine by capillary electrophoresis after derivatization by ophthaldialdehyde. Electrophoresis. 1994;15(6):763-8. doi: 10.1002/elps.11501501106, PMID 7982397.
- Jaworska M, Szulinska G, Wilk M, Tautt J. Capillary electrophoretic separation of N-acetylcysteine and its impurities as a method for quality control of pharmaceuticals. J Chromatogr A. 1999;853(1-2):479-85. doi: 10.1016/s0021-9673(99)00727x, PMID 10486756.
- 52. Zinellu A, Sotgia S, Scanu B, Usai MF, Fois AG, Spada V. Simultaneous detection of N-acetyl-L-cysteine and physiological low molecular mass thiols in plasma by capillary electrophoresis. Amino Acids. 2009;37(2):395-400. doi: 10.1007/s00726-008-0167-x, PMID 18695935.
- 53. ICH Q2. Validation of analytical procedures: text and methodology. In: Proceedings of the international conference on harmonization. Vol. R1. Geneva, Switzerland; 2005. p. 1-13.