

Original Article

DIFFERENCE SPECTROPHOTOMETRIC METHOD FOR SIMULTANEOUS ESTIMATION OF MOXIFLOXACIN AND CEFIXIME TRIHYDRATE IN BULK AND COMBINED DOSAGE FORM

RAVI KANT\*, RAMESH BODLA, RUBINA BHUTANI, GARIMA KAPOOR

Delhi Institute of Pharmaceutical Sciences and Research (DIPSAR), Sector 3 Pushp Vihar, Mehrauli Badarpur Road, New Delhi 110017, India  
Email: ravi.taurean@gmail.com

Received: 17 Jun 2015 Revised and Accepted: 17 Jul 2015

ABSTRACT

**Objective:** To develop rapid, accurate, reproducible, validated and economical difference spectroscopy method for the simultaneous determination of moxifloxacin (MFN) and cefixime (CEF) in tablet dosage forms.

**Methods:** The method comprised the measurement of the absorbance of a solution of the tablet extract in 0.1 M NaOH relative to that of an equimolar solution in 0.1 M HCl at 254 nm for MFN and 292 nm for CEF. The presence of identical isosbestic points for pure drug solutions and tablet extracts indicated the non-interference of excipients in the absorption at these wavelengths.

**Results:** The method was found to be linear over the concentration range of 10-50 µg/ml for CEF and 4-20 µg/ml for MFN. Accuracy was found to be in the range of 99.91-101.18%. Relative standard deviation for precision and intermediate precision was found to be less than 2%. The developed method was successfully applied for the simultaneous estimation of Moxifloxacin and Cefixime in tablet formulation. The results obtained from the validation experiments prove that the developed method is suitable for routine analysis.

**Conclusion:** This method is simple, selective, linear, precise, and accurate and sensitive hence can be successfully employed for the routine quality control of dosage forms containing both the drugs in pharmaceutical industries.

**Keywords:** Moxifloxacin, Cefixime, Difference Spectrophotometry, Method validation.

INTRODUCTION

Cefixime Trihydrate [(6R,7R)-7-(2-(2-Amino-4-thiazolyl) glyoxyl-amido)-8-oxo-3-vinyl-5-thia-1-azabicyclo [4.2.0] oct-2-ene-2-carboxylic acid, 72-(Z)-[O-(carboxymethyl) oxime] trihydrate [fig. 1(a)] is semi synthetic, oral, third-generation cephalosporin antibiotic like ceftriaxone and cefotaxime [1]. It acts by inhibition of muco peptide synthesis in the bacterial cell wall [2].

Cefixime is active against a very wide spectrum of bacteria and is used in the treatment of otitis media, respiratory tract infection, and

typhoid fever, complicated and uncomplicated Urinary Tract Infection [3-10]. It is official in United States Pharmacopoeia (USP) 2015 and British Pharmacopoeia (BP) 2015 [11].

Both the pharmacopoeia describes HPLC method of analysis for Cefixime trihydrate. HPLC method is also official in Indian Pharmacopoeia 2014[12] and Japan Pharmacopoeia 2014[13]. Literature reports many analytical methods for the determination of CEF in single and in combination with other drug, using UV spectroscopy [14-15] spectro fluorometry [16], HPLC [17-24] and HPTLC [25].

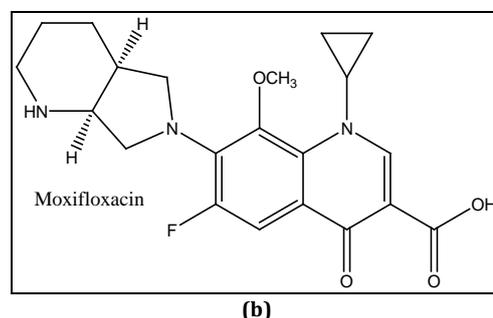
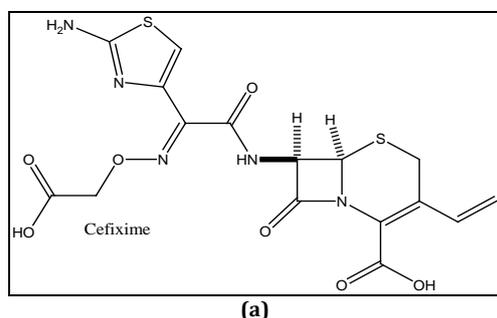


Fig. 1: Chemical structures of (a) Cefixime and (b) Moxifloxacin

Moxifloxacin (MFN), 1-cyclopropyl-7-[(1S,6S)-2,8-diazabicyclo [4.3.0] non-8-yl]-6 fluoro-8-methoxy-4-oxo-quinoline-3-carboxylic acid [fig.1(b)] is a synthetic fourth generation fluoro quinolone antibiotic[26]. The mechanism of action involve inhibition of an enzyme topoisomerase II (DNA gyrase), which is essential for bacterial DNA replication [27]. It is used in ocular infection (conjunctivitis), acute sinusitis, lower respiratory tract infections and urinary tract infection [28-32]. Moxifloxacin is official in BP 2010[33]. Several analytical methods have been reported for the determination of MFN in formulations and biological fluids, such as UV spectroscopic methods [34-35], Spectro fluorometry [36], RP-HPLC [37-41], and capillary electrophoresis [42-43].

The new combination of MFN and CEF is approved by the Central Drugs Standard Control Organization (CDSCO) India for the treatment of lower respiratory tract infections in adults [44].

Simultaneous determination of these drugs is essential in each step of initial formulation development and screening stage of any solid dosage form. This combination is not official in any of the pharmacopoeia and no official method is available for the simultaneous estimation of Cefixime and Moxifloxacin in the combined dosage forms.

The objective of the current study is to develop a easy, rapid, accurate, reliable, reproducible, validated and economical Difference

Spectroscopy Method for the simultaneous determination of CEF and MFN in tablet dosage forms.

## MATERIALS AND METHODS

### Apparatus

UV/Visible Spectrophotometer: SICAN-2301, Inkar Instruments Pvt Ltd.

Analytical Balance: Sartorius BSA223S-CW

Magnetic Stirrer: REMI 1MLH, Remi Laboratories Limited.

### Chemicals and reagents

Moxifloxacin: Gift sample from Covalent Laboratories Pvt. Ltd., Hyderabad.

Cefixime: Gift sample from Neuland Laboratories Ltd., Hyderabad.

Formulation of Moxifloxacin and Cefixime: Moxicip FC, Cipla Limited and Mahacef, Mankind Ltd.

Solvent: Methanol Analytical Grade, Merck.

Diluent: 0.1N Sodium Hydroxide, 0.1N Hydrochloric acid

### Method development and optimization

#### Preparation of standard stock solution and construction of calibration curve

#### Preparation of Moxifloxacin standard stock solution and plotting overlay UV spectra

The stock solution of MFN was prepared by dissolving 50 mg of pure MFN in 50 ml of methanol. Appropriate aliquots of the stock solution were transferred into two different 25 ml volumetric flasks. The volume was made up with 0.1 N HCl and 0.1 N NaOH to give a series of equimolar solutions of 25 ml each in 0.1 N HCl and 0.1 N NaOH containing 4-20  $\mu\text{g/ml}$  of MFN. The wavelength scan over a range 400-200 nm was taken and overlay spectra was plotted (fig. 2).

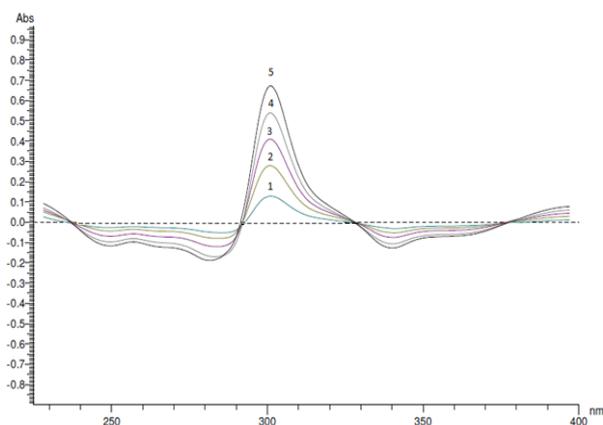


Fig. 2: Overlay UV Spectra of Moxifloxacin (4-20  $\mu\text{g/ml}$ )

#### Preparation of Cefixime standard stock solution and plotting overlay of UV spectra

The stock solution of CEF was prepared by dissolving 50 mg of pure CEF in 50 ml of methanol. Appropriate aliquots were used as for MFN to prepare 25 ml series of equimolar solutions of CEF in 0.1 N HCl and 0.1 N NaOH containing 10-50  $\mu\text{g/ml}$  CEF. The wavelength scan over a range 400-200 nm were taken and overlay spectra was plotted (fig. 3).

#### Calibration curve for moxifloxacin and cefixime trihydrate and their synthetic mixtures

Similar to above, two series of equimolar solutions of mixtures of 25 ml MFN and CEF in 0.1 N HCl and 0.1N NaOH were also prepared

using the stock solutions. The first series contained a constant concentration of CEF (20  $\mu\text{g/ml}$ ) and a varying concentration of MFN (4-20  $\mu\text{g/ml}$ ). The second series contained a constant concentration of MFN (20  $\mu\text{g/ml}$ ) and a varying concentration of CEF (10-50  $\mu\text{g/ml}$ ). The drugs were protected from light throughout the study and the absorbance of the solutions of pure MFN, CEF and their mixtures were taken between 30 and 90 min after preparation. All reagents used were of analytical grade.

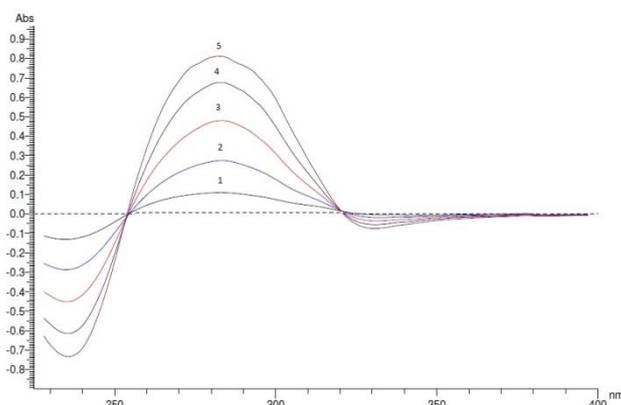


Fig. 3: Overlay UV spectra of cefixime (10-50  $\mu\text{g/ml}$ )

#### Estimation of moxifloxacin and cefixime in combined tablet dosage form

Twenty tablets were accurately weighed, well powdered and a weight of the powder equivalent to 100 mg of MFN (and 100 mg of CEF) was dissolved in 40 ml of methanol by thorough mixing and made up to volume in a 100 ml volumetric flask. The extract was filtered through a Whatman filter paper No. 41. The first and last 5 ml of the filtrate was discarded. The sample solutions of 25 ml of each in 0.1 N HCl and 0.1 N NaOH were prepared using 0.5 ml aliquots of the filtrate using a micropipette (range 100-1000  $\mu\text{l}$ ) so as to obtain equimolar solutions containing approximately 20 $\mu\text{g/ml}$  of MFN and 20 $\mu\text{g/ml}$  of CEF. The absorbance difference ( $\delta A$ ) between the acidic solution and equimolar 0.1 N NaOH solutions of pure drugs and samples were measured from 230 to 400 nm on a SICAN-2301 UV-visible double beam auto scan spectrophotometer by placing the 0.1 N NaOH solutions in the reference compartment and the acidic solutions in the sample compartment (fig. 4).

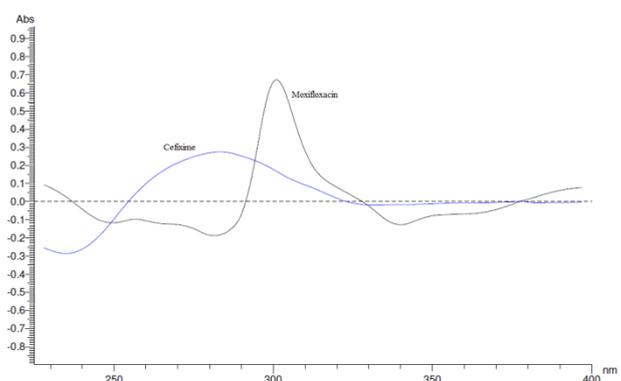


Fig. 4: Overlay spectra of cefixime (20  $\mu\text{g/ml}$ ) and Moxifloxacin (20 $\mu\text{g/ml}$ )

#### Selection of wavelength

The absorbance difference of the analytes at 254 and 292 nm was corrected for the absorbance difference, if any, of 0.1 N NaOH solution relative to 0.1 N HCl at these wavelengths. The difference

absorption spectrum of a solution of MFN in 0.1 N HCl solutions in the reference cell and an equimolar solution of MFN in 0.1 N NaOH solutions in the sample cell compartment showed a maximum value of  $\delta A$  at 301 nm and a minimum value of  $\delta A$  at 283.5 nm. An isosbestic point (a wavelength of zero  $\delta A$  due to equal absorptivities of the two species) occurred at 292 nm (fig. 4). The difference absorption spectrum of solutions of CEF showed maximum values of  $\delta A$  at 297 nm and a minimum value of  $\delta A$  at 268 nm. The isosbestic points of the CEF spectrum were obtained at 254 nm (fig. 4).

The wavelength of 254 nm was chosen for the estimation of MFN. For the wavelength of 254 nm, at which the  $\delta A$  value of the MFN difference spectrum was about 0.103 for a concentration of 20  $\mu\text{g/ml}$ , the absorbance value of the CEF difference spectrum was about 0.241 at 292 nm for a concentration of 20  $\mu\text{g/ml}$ . These concentrations were chosen on the basis of the proportions of MFN and CEF in commercial formulations. The proportionality of the  $\delta A$  value and concentration of MFN was found by measuring  $\delta A$  of the 5 pairs of solutions containing 4-20 $\mu\text{g/ml}$  of MFN at 254 nm. The linear regression equation calculated using the method of least squares was  $y = 0.0051x + 0.0007$  (1) with a correlation coefficient of  $r = 0.9994$ . The proportionality of  $\delta A$  and the concentration of CEF were found by measuring the  $\delta A$  values of solutions of CEF containing 10-50  $\mu\text{g/ml}$  at 292 nm. The calculated linear regression equation was  $y = 0.0164x - 0.0698$  (2) with a correlation coefficient of  $r = 0.9994$  (fig. 5 and table 1)

#### Method validation

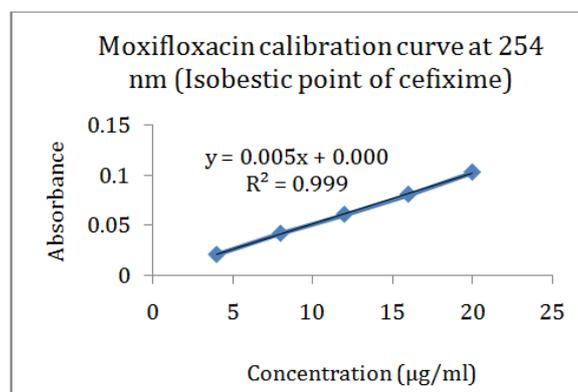
The developed method was validated according to ICH Guidelines [45]. The following parameters were considered: specificity, linearity, limit of detection (LOD), limit of quantitation (LOQ), accuracy, and precision.

#### Specificity

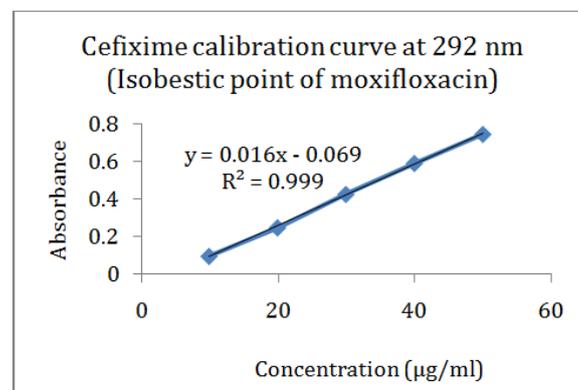
To evaluate further the specificity of the method for samples containing MFN and CEF, two series each of 5 solutions (mentioned under standard preparation) were examined at the isosbestic wavelengths. The solutions of the first series gave a regression equation of  $y = 0.0053x + 0.0017$  (3) with a correlation coefficient of  $r = 0.9995$  at 254 nm, which was similar to that of Eq. (1), suggesting that the presence of CEF did not affect the absorptivity of MFN at 254 nm. The  $\delta A$  values of the second series of solutions gave a regression equation of  $y = 0.0166x - 0.0709$  (4) with a correlation coefficient of  $r = 0.9993$  at 292 nm. Its similarity to Eq. (2) suggests no interference of the absorptivity of MFN with that of CEF at 292 nm. The identical isosbestic points of the two components in the standard and sample difference spectra confirmed the non-interference of the excipients in the measurement of the absorbance values at these wavelengths.

#### Linearity and range

Linearity is expressed in terms of correlation co-efficient of linear regression analysis. The linearity response was determined by analyzing 5 independent levels of calibration curve in the range of 4-20  $\mu\text{g/ml}$  for Moxifloxacin and 10-50  $\mu\text{g/ml}$  for Cefixime. Plot the calibration curve of absorbance v/s concentration and determine correlation coefficient and regression line equations for Moxifloxacin and Cefixime.



(a)



(b)

Fig. 5: Linearity graph for (a) Moxifloxacin (4-20  $\mu\text{g/ml}$ ) and (b) Cefixime (10-50  $\mu\text{g/ml}$ )

Table 1: Data of optical characteristics

Parameters	Observed Value	
	Drugs	
	Moxifloxacin	Cefixime
Beer's Law Limit ( $\mu\text{g/ml}$ )	10-50 $\mu\text{g/ml}$	4-20 $\mu\text{g/ml}$
Correlation Coefficient ( $R^2$ )	0.9994	0.9994
Regression Equation ( $y = mx + c$ )	$y = 0.0051x + 0.0007$	$y = 0.0164x - 0.0698$
Slope	0.0051	0.0164
Intercept	0.0007	-0.0698

#### Accuracy

#### Preparation of sample solution

Twenty tablets were powdered. Powder equivalent to 400 mg of Moxifloxacin and 400 mg of Cefixime was weighed and transferred into 400 ml of the volumetric flask. Then 80 ml of methanol was added and solution was sonicated for 20 minutes and diluted up to mark with Distilled Water. The solution was filtered using Whatman filter paper no.41 and first few drops of filtrate were discarded.

Known amounts of standard solutions of MFN (4.8, 6, 7.2  $\mu\text{g/ml}$ ) and CEF (12, 15, 18 $\mu\text{g/ml}$  for CEF) were added to pre quantified sample solutions of MFN (6 $\mu\text{g/ml}$ ) and CEF (15  $\mu\text{g/ml}$ ) of tablet dosage form. Absorbances of solutions were measured at selected wavelengths for MFN and CEF.

The amounts of MFN and CEF were estimated by applying obtained values ( $n = 6$ ) to the regression equation of the calibration curve. The amount of MFN and CEF was calculated at each level and % recoveries were computed (table 2).

Table 2: Recovery studies

Accuracy (Recovery studies of moxifloxacin)							SD	%RSD
Drug (level of % recovery)	Sample No	Amount Present, B ( $\mu\text{g/ml}$ )	Amount added, C ( $\mu\text{g/ml}$ )	Amount found, A ( $\mu\text{g/ml}$ )	Amount recovered (A-B) ( $\mu\text{g/ml}$ )	% Recovered [(A-B)/C]*100 ( $\mu\text{g/ml}$ )	+	
Moxifloxacin (80%)	1	6	4.8	10.89	4.89	101.88	1.20	1.19
	2	6	4.8	10.79	4.79	99.79		
	3	6	4.8	10.89	4.89	101.88		
					Mean	101.18		
Moxifloxacin (100%)	1	6	6	12.11	6.11	101.83	1.93	1.93
	2	6	6	12.02	6.02	100.33		
	3	6	6	11.88	5.88	98.00		
					Mean	100.06		
Moxifloxacin (120%)	1	6	7.2	13.11	7.11	98.75	1.18	1.18
	2	6	7.2	13.28	7.28	101.11		
	3	6	7.2	13.19	7.19	99.86		
					Mean	99.91		

Accuracy (Recovery studies of cefixime)							SD	%RSD
Drug (level of % recovery)	Sample No	Amount Present, B ( $\mu\text{g/ml}$ )	Amount added, C ( $\mu\text{g/ml}$ )	Amount found, A ( $\mu\text{g/ml}$ )	Amount recovered (A-B) ( $\mu\text{g/ml}$ )	% Recovered [(A-B)/C]*100 ( $\mu\text{g/ml}$ )		
Cefixime (80%)	1	15	12	26.87	11.87	98.92	1.89	1.89
	2	15	12	26.93	11.93	99.42		
	3	15	12	27.29	12.29	102.42		
					Mean	100.25		
Cefixime (100%)	1	15	15	29.87	14.87	99.13	1.21	1.21
	2	15	15	30.21	15.21	101.40		
	3	15	15	29.93	14.93	99.53		
					Mean	100.02		
Cefixime (120%)	1	15	18	32.89	17.89	99.39	1.00	0.99
	2	15	18	33.19	18.19	101.06		
	3	15	18	33.21	18.21	101.17		
					Mean	100.54		

**Precision****Intraday (Repeatability)**

Solutions containing 8, 12, 16  $\mu\text{g/ml}$  MFN and 20, 30, 40  $\mu\text{g/ml}$  CEF in triplicates were analyzed thrice on the same day. The results were reported in terms of relative standard deviation (%RSD) (table 3).

**Interday (Intermediate)**

Solutions containing 8, 12, 16  $\mu\text{g/ml}$  MFN and 20, 30, 40  $\mu\text{g/ml}$  of CEF in triplicates were analyzed for 3 different days. The results were reported in terms of relative standard deviation (%RSD) (table 3).

Table 3: Precision studies

Intraday analysis of formulation					SD	%RSD
Drug	Sampling Time	Concentration ( $\mu\text{g/ml}$ ) taken	Concentration found ( $\mu\text{g/ml}$ )	%age obtained		
Moxifloxacin	9:00 AM	8	8.07	100.85	0.13	1.54
	1:00 AM	12	12.00	100.03	0.19	1.61
	5:00 PM	16	16.05	100.3	0.15	0.96
Cefixime	9:00 AM	20	20.07	100.36	0.18	0.87
	5:00 PM	40	39.81	99.52	0.48	1.22
Interday analysis of formulation					SD	%RSD
Drug	Sample No.	Concentration ( $\mu\text{g/ml}$ ) taken	Concentration found ( $\mu\text{g/ml}$ )	%age obtained		
Moxifloxacin	Day 1	8	8.03	100.38	0.14	1.68
	Day 2	12	12.03	100.27	0.15	1.28
	Day 3	16	16.05	100.34	0.21	1.29
Cefixime	Day 1	20	20.07	100.36	0.17	0.84
	Day 2	30	30.08	100.25	0.17	0.58
	Day 3	40	40.02	100.05	0.14	0.34

**Limit of detection (LOD) and limit of quantitation (LOQ)**

The LOD and LOQ of MFN and CEF by proposed methods were determined using calibration standards. LOD and LOQ were calculated as  $3.3\sigma/S$  and  $10\sigma/S$  respectively, where S is the slope of the calibration curve and  $\sigma$  is the standard deviation of response.

**RESULTS**

The solubility of Moxifloxacin and Cefixime was studied and distilled water was selected as a choice of solvent. Two drugs individually followed Beer-Lambert's law over the concentration range of 10-50  $\mu\text{g/ml}$  for CEF and 4-20  $\mu\text{g/ml}$  for MFN. Coefficient of correlation for

MFN and CEF were found to be 0.9994 and 0.9994 respectively. The values of correlation coefficient suggest the level of precision of the method. Drug content in tablet (amount present) was directly found from the above mentioned regression equations for both drugs. Standard deviations, and % RSD were calculated and are given in table 4. Percentage estimation in tablet dosage form was 101.07% and 101.06% (%RSD<2) for MFN and CEF respectively (table 4).

## DISCUSSION

In the present study, the difference absorption spectra of MFN in 0.1 N NaOH vs. 0.1 N HCl showed zero crossing point at 292, 328 and 377 nm. The wavelength of 292 nm was chosen for measuring the absorbance of CEF, since the ( $\delta A$ ) values of the CEF difference spectra at this point were more optimal and linear for accurate measurement of different concentrations of CEF.

Similarly, the difference absorption spectra of CEF in 0.1 N NaOH vs. 0.1 N HCl showed zero crossing point at 254 and 320 nm, but the absorbance of MFN was measured at the wavelength of 254 nm due to more linear ( $\delta A$ ) values at this wavelength.

The method was validated according to International Conference on Harmonization guidelines for validation of analytical procedures. Linear regression equations (intercepts and slopes) for MFN and CEF were established. The values of slope, intercept and correlation coefficient values are given in table 1. LOD for TAZ and CEF were found to be 0.92  $\mu\text{g/ml}$  and 0.98  $\mu\text{g/ml}$ , respectively. LOQ for TAZ and CEF were found to be 2.79  $\mu\text{g/ml}$  and 2.97  $\mu\text{g/ml}$ , respectively. To study the validation parameters accuracy, reproducibility, reliability and interference, recovery experiment was carried out by standard addition. The recovery of added standard was calculated at different concentration levels. From the total amount of drug found, the percentage recovery was calculated which was between 98-102 % (RSD<2.0).

Table 4: Analysis of tablet formulation

Brand	Drug	Labeled Claim (mg/tab)	Amount Found (mg/tab)	%Purity	SD	%RSD
Mahacef (Mankind)	Moxifloxacin	400	404.29	101.07	1.35	0.34
	Cefixime	400	404.23	101.06	0.74	0.18
Moxicip (Cipla)	Moxifloxacin	400	404.14	101.04	0.90	0.22
	Cefixime	400	403.92	100.98	0.68	0.17

## CONCLUSION

The proposed method is simple, precise, and accurate for the simultaneous determination of Moxifloxacin and Cefixime in combined tablet dosage forms and this method may be successfully applied in quality control laboratories for their determination in combined dosage form.

## ACKNOWLEDGEMENT

The Authors are thankful to the Neuland Laboratories Ltd, Hyderabad and Covalent Laboratories Pvt. Ltd., Hyderabad for providing gift samples of the drug samples for research. Also we are thankful to the Director, Delhi Institute of Pharmaceutical Sciences and Research for providing the infrastructure and to an University Grants Commission for providing the fund for the research work.

## CONFLICT OF INTERESTS

Declared None

## REFERENCES

1. "United States Pharmacopeia" USP 38, United States Pharmacopeia Convention, Inc; 2015. p. 2665.
2. Hooper DC, Wolfson JS. Mechanisms of quinolone action and bacterial killings Quinolone Antimicrobial Agents, American Society for Microbiology. Vol. 1-2<sup>nd</sup> Edition. Washington DC; 1993. p. 53-7.
3. Claudia CO. Analytical profiles of drug substances, excipient and methodology. An Imprint Elsevier 1998;25:39-89.
4. Dhib M, Moulin B, Godin R. Relationship between renal function and disposition of oral cefixime. Eur J Clin Pharmacol 1991;41:579-83.
5. Faulkner RD, Bohaychuk W, Desjardins RE, Look ZM, Haynes JD, Weiss AI, et al. Bioequivalency of solid oral dosage forms of Cefixime. J Clin Pharmacol 1987;27:807.
6. Faulkner RD, Fernandez P, Lawrence G, Sia LL, Falkowski AJ, Weiss AI, et al. Absolute bioavailability of cefixime in man. J Clin Pharmacol 1988;28:700.
7. Faulkner RD, Bohaychuk W, Haynes JD, Desjardins RE, Yacobi A, Silber BM. Influence of an antacid containing aluminium and magnesium on the pharmacokinetics of cefixime. Eur J Clin Pharmacol 1988;34:525-8.
8. Markham A, Brogden RN. Cefixime-a review of its therapeutic efficacy in lower respiratory tract infections. Drugs 1995;49:1007-22.
9. Parfit K. Martindale the complete drug reference. 35th edition. Vol. 1. Pharmaceutical press; 2007. p. 59.
10. <http://www.medlineindia.com/cefixime>. [Last accessed on 05 Jun 2015]
11. British Pharmacopoeia. Vol. I-II. Government British Pharmacopoeial Commission; 2011. p. 1489.
12. Indian pharmacopoeia. Vol. 2. Government of India, the Controller Publication, New Delhi; 2010. p. 1012.
13. Japanese Pharmacopoeia. 15<sup>th</sup> edition. Society of Japanese Pharmacopoeia, Tokyo; 2006;443:938-9.
14. Attimarad M, Anroop B. Simultaneous determination of ofloxacin and cefixime by first and ratio first derivative UV spectrophotometry. Chron Young Sci 2011;2:144-9.
15. Shankar DG, Sushma K, Laxmi RV, Reddy MN, Murthy TK, Rao SY. UV and visible spectrophotometric methods for the determination of cefixime. Indian Drugs 2001;38:617-9.
16. Bukhari N, Al-Warthan A, Wabaidur SM, Othman ZA, Javid M, Haider S. Spectrofluorimetric determination of cefixime in pharmaceutical preparation and biological fluids using calcein as a fluorescence probe. Sens Lett 2010;8:280-4.
17. Dhoka MV, Sandage SJ, Dumbre SC. Simultaneous determination of cefiximetric hydrate and dicloxacillin sodium in pharmaceutical dosage form by reversed phase high performance liquid chromatography. J AOAC Int 2010;93:531-5.
18. Gonzalez-Hernandez R, Nuevas-Paz L, Soto-Mulet L, Lopez-Lopez M, Hoogmartens J. Reversed phase high performance liquid chromatographic determination of cefixime in bulk drugs. J Liq Chromatogr Relat Technol 2001;24:2315-24.
19. Hafiz Muhammad A, Shahnaz G, Raheela B, Muhammad IN. Development of HPLC-UV Method for analysis of cefixime in raw materials and in capsule. Jordan J Pharm Sci 2009;2:53-65.
20. Khan IU, Sharif S, Ashfaq M, Asghar MN. Simultaneous determination of potassium clavulanate and cefixime in synthetic mixtures by high performance liquid chromatography. J AOAC Int 2008;91:744-9.
21. Manna L, Valvo L. Development and validation of a fast reversed-phase ion-pairing liquid chromatographic method for simultaneous determination of eight cephalosporin antibiotics in pharmaceutical formulations. Chromatographia 2004;60:645-9.
22. Rathinavel G, Mukherjee PB, Valarmathy J, Samuel Joshua L, Ganesh M, Sivakumar T, et al. Validated RP-HPLC method for simultaneous estimation of cefixime and cloxacillin in tablets. Eur J Chem 2008;5:648-51.
23. Shah PB, Pundarikakshudu K. Spectrophotometric, difference spectroscopic, and high-performance liquid chromatographic methods for the determination of cefixime in pharmaceutical formulations. J AOAC Int 2006;89:987-94.

24. Meng F, Chen X, Zeng Y, Zhong D. Sensitive liquid chromatography tandem mass spectrometry method for the determination of cefixime in human plasma: application to a pharmacokinetic study. *J Chromatogr B: Anal Technol Biomed Life Sci* 2005;819:277–82.
25. Pawar SJ, Kale AP, Amrutkar MP, Jagade JJ, Pore NS, Bhosale AV. HPTLC estimation of cefixime and cloxacillin in tablet dosage form. *Asian J Res Chem* 2010;3:299–301.
26. S. Budavari, Eds. In: *The Merck Index: Encyclopedia of Chemicals, drugs and biological*. 13th Ed. New Jersey: Published by Merck Research Laboratories, Division of Merck and Co., Inc. Whitehouse station; 2001. p. 1097, 1125.
27. Keating GM, Scott LJ. Moxifloxacin: a review of its use in the management of bacterial infections. *Drugs* 2004;64:2347–77.
28. <http://en.wikipedia.org/wiki/Moxifloxacin>. [Last accessed on 10 Jun 2015]
29. <http://www.drugbank.ca/drugs/DB00218>. [Last accessed on 10 Jun 2015]
30. Rang HP, Dale MM, Ritter JM. *Flower. Pharmacology*. 6th Edition. Elsevier publication house; 2001. p. 647-8.
31. <http://www.rxlist.com/vigamox-drug.htm>. [Last accessed on 10 Jun 2015]
32. <http://www.medlineindia.com/eye/moxifloxacin.htm>. [Last accessed on 10 Jun 2015]
33. *British pharmacopeia*. Vol II-III. 6th edition. London: Her Majesty's Stationary Office; 2010. p. 1460.
34. Motwani SK, Chopra S, Ahmad FJ, Khar RK. Validated spectrophotometric methods for the estimation of moxifloxacin in bulk and pharmaceutical formulations. *Spectrochim Acta Part A* 2007;68:250–6.
35. Patel PU, Suhagia BN, Patel MM. Simultaneous spectrophotometric determination of Moxifloxacin and Metronidazole in synthetic mixture by simultaneous equations method. *Indian Drugs* 2005;2:155–7.
36. Ocaña JA, Barragán FJ, Callejón M. Spectrofluorimetric determination of moxifloxacin in tablets, human urine and serum. *Analyst* 2000;125:2322–5.
37. Predrag D, Andrija C, Aleksandra D, Milena Jelicki S. Optimization of separation and determination of moxifloxacin and its related substances by RP-HPLC. *J Pharm Biomed Anal* 2009;50:117–26.
38. Nguyena HA, Grelleta J, Ba BB, Quentin C, Saux MC. Simultaneous determination of levofloxacin, gatifloxacin and moxifloxacin in serum by liquid chromatography with column switching. *J Chromatogr B* 2004;810:77–83.
39. Smet JD, Boussery K, Colpaert K, Suttera PD, Paepe PD, Decruyenaere J, *et al.* Pharmacokinetics of fluoroquinolones in critical care patients: a bio-analytical HPLC method for the simultaneous quantification of ofloxacin, ciprofloxacin and moxifloxacin in human plasma. *J Chromatogr B* 2009;877:961–7.
40. Pranger AD, Alffenaar JW, Wessels AM, Greijdanus B, Uges DR. Determination of moxifloxacin in human plasma, plasma ultra filtrate and cerebrospinal fluid by a rapid and simple liquid chromatography–tandem mass spectrometry method. *J Anal Toxicol* 2010;34:135–41.
41. Vu DH, Koster RA, Alffenaar JW, Brouwers JR, Uges DR. Determination of moxifloxacin in dried blood spots using LC-MS/MS and the impact of the hematocrit and blood volume. *J Chromatogr B: Anal Technol Biomed Life Sci* 2011;879:1063–70.
42. Cruz LA, Hall R. Enantiomeric purity assay of moxifloxacin hydrochloride by capillary electrophoresis. *J Pharm Biomed Anal* 2005;38:8–13.
43. Moller JG, Stass H, Heining R, Blaschke G. Capillary electrophoresis with laser induced fluorescence: a routine method to determine moxifloxacin in human body fluids in very small sample volumes. *J Chromatogr B* 1998;716:325–34.
44. <http://cdsco.nic.in/writereaddata/Approved%20FDC%20list%20till%20november%202014.pdf> [Last accessed on 15 Jun 2015]
45. ICH Harmonized Tripartite Guideline. Validation of Analytical Procedures: Text and Methodology Q2 (R1), International Conference on Harmonization, Geneva, Switzerland Nov; 2005.